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                German (DE) application and patent publication number format
                changes
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NEWS 24 MAR 03 MEDLINE file segment of TOXCENTER reloaded
NEWS 25 MAR 03 FRANCEPAT now available on STN
NEWS 26 MAR 29 Pharmaceutical Substances (PS) now available on STN
NEWS 27 MAR 29 WPIFV now available on STN
NEWS 28 MAR 29 No connect hour charges in WPIFV until May 1, 2004
NEWS 29 MAR 29 New monthly current-awareness alert (SDI) frequency in RAPRA
NEWS EXPRESS MARCH 5 CURRENT WINDOWS VERSION IS V7.00A, CURRENT
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             AND CURRENT DISCOVER FILE IS DATED 3 MARCH 2004
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L3 1238 CRE LOX

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=> s FRT () FLP

L5 96 FRT (W) FLP

=> s 15 and recombination

L6 82 L5 AND RECOMBINATION

=> s 16 and stability

L7 27 L6 AND STABILITY

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L7 ANSWER 1 OF 27 MEDLINE on STN

TI High-level heterologous gene expression in Saccharomyces cerevisiae from a

stable 2 microns plasmid system.

The best candidate for a high-copy-number and mitotic stability AB expression system in yeast is the endogenous 2 microns plasmid. Nevertheless, derivatives of the 2 microns plasmid typically exhibit lower copy numbers and require selection for adequate maintenance within cells. We report the construction and utilization of an efficient heterologous gene expression system containing a 4.5-kb inducible expression cassette inserted into the 2 microns plasmid and selected in cells utilizing a carrier plasmid which is subsequently lost via FRT/Flp recombination. The non-selectable 2 micron plasmid, containing the cassette, was found to be stably maintained in cells, without selection, at high copy number. The dynamics of resolution and partitioning of this plasmid were analyzed during the course of 50 generations of growth under non-selective conditions. The heterologous lacZ reporter gene coding for beta-galactosidase (beta Gal) is driven by the hybrid, galactose-inducible promoter GAL10::pMF alpha 1. Upon induction, beta Gal was secreted into the periplasm and culture supernatant at levels which could be detected directly from Coomassie blue-stained SDS-PAGE. Furthermore, plasmid-containing cells could be maintained directly on rich YPD medium and identified either by utilizing XGal or by observing inhibition of colony growth on YPGal solid medium. The cassette was designed for direct, high-level, inducible expression of cloned genes downstream from the MF alpha 1 signal sequence, with or without a C-terminal lacZ fusion. This vector represents the first demonstration of a non-selectable, mitotically stable, episomal plasmid system capable of expressing recombinant proteins at high levels. (ABSTRACT TRUNCATED AT 250 WORDS)

ACCESSION NUMBER: 94010342 MEDLINE DOCUMENT NUMBER: PubMed ID: 8406040

TITLE: High-level heterologous gene expression in Saccharomyces

cerevisiae from a stable 2 microns plasmid system.

AUTHOR: Ludwig D L; Ugolini S; Bruschi C V

CORPORATE SOURCE: Microbiology Department, International Centre for Genetic

Engineering and Biotechnology, Trieste, Italy.

SOURCE: Gene, (1993 Sep 30) 132 (1) 33-40.

Journal code: 7706761. ISSN: 0378-1119.

PUB. COUNTRY: Netherlands

DOCUMENT TYPE: Journal; Article; (JOURNAL ARTICLE)

LANGUAGE: English

FILE SEGMENT: Priority Journals; Space Life Sciences

ENTRY MONTH: 199311

ENTRY DATE: Entered STN: 19940117

Last Updated on STN: 19940117 Entered Medline: 19931119

L7 ANSWER 2 OF 27 USPATFULL on STN

TI Modified starch metabolism enzymes and encoding genes for improvement

and optimization of plant phenotypes

The invention provides methods for generating, identifying, and selecting polynucleotides encoding novel starch metabolizing enzymes (NSME), NSME-encoding polynucleotides, compositions of recombinant shuffled NSME protein, plant cells and microbes containing a shuffled NSME polynucleotide in expressible form, plants containing a shuffled NSME polynucleotide in expressible form, novel starch compositions produced by plants and cells, uses of such plants, cells, and starch compositions.

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

ACCESSION NUMBER: 2004:59856 USPATFULL

Modified starch metabolism enzymes and encoding genes TITLE:

> for improvement and optimization of plant phenotypes Stemmer, Willem P. C., Los Gatos, CA, United States Subramanian, Venkitswaran, San Diego, CA, United States

Raillard, Sun Ai, Mountain View, CA, United States

Huisman, Gjalt, San Carlos, CA, United States

Maxygar, Inc., Redwood City, CA, United States (U.S. PATENT ASSIGNEE(S):

corporation)

NUMBER KIND DATE -----PATENT INFORMATION: US 6703240 B1 20040309 US 2000-547844 20000412

20000412 (9) APPLICATION INFO.:

NUMBER DATE _____

US 1999-129009P 19990413 (60) PRIORITY INFORMATION:

DOCUMENT TYPE: Utility GRANTED FILE SEGMENT:

PRIMARY EXAMINER: ASSISTANT EXAMINER: Reynolds, Deborah J. Woitach, Joseph

LEGAL REPRESENTATIVE: Holmer, Christopher, Kruse, Norman J., Townsend &

Townsend & Crew

82 NUMBER OF CLAIMS: EXEMPLARY CLAIM: 1

INVENTOR(S):

3 Drawing Figure(s); 3 Drawing Page(s) NUMBER OF DRAWINGS:

LINE COUNT: 2972

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

L7 ANSWER 3 OF 27 USPATFULL on STN

TIIn vitro mutagenesis, phenotyping, and gene mapping

Cellular libraries useful for in vitro phenotyping and gene mapping. In AB a representative approach, a method for preparing a homozygous cellular library includes the steps of providing a heterozygous cellular library comprising a plurality of isolated parent cells; inducing site-specific mitotic recombination in the plurality of isolated parent cells; culturing the plurality of isolated parent cells, whereby a population of daughter cells is produced; and selecting daughter cells comprising a homozygous genetic modification, whereby a homozygous

cellular library is prepared.

CAS INDEXING IS AVAILABLE FOR THIS PATENT. 2004:44603 USPATFULL ACCESSION NUMBER:

In vitro mutagenesis, phenotyping, and gene mapping TITLE: Threadgill, David W., Chapel Hill, NC, UNITED STATES INVENTOR(S):

Lee, Daekee, Chapel Hill, NC, UNITED STATES

NUMBER KIND DATE PATENT INFORMATION: US 2004033596 A1 20040219 US 2003-428977 A1 20030502 (10) APPLICATION INFO.:

> NUMBER DATE -----

PRIORITY INFORMATION: US 2002-377864P 20020502 (60)

DOCUMENT TYPE: Utility FILE SEGMENT: APPLICATION

LEGAL REPRESENTATIVE: JENKINS & WILSON, PA, 3100 TOWER BLVD, SUITE 1400,

DURHAM, NC, 27707

NUMBER OF CLAIMS: 71 EXEMPLARY CLAIM: 1

NUMBER OF DRAWINGS: 6 Drawing Page(s)

LINE COUNT: 3125

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

ANSWER 4 OF 27 USPATFULL on STN **L**7

ΤI Self-rearranging DNA vectors

AB Disclosed are replicatable viral DNA vectors encoding a site-specific DNA-altering enzyme and a DNA target recognized by the enzyme, the enzyme selectively converting, in a cell expressing the enzyme, the DNA vector to a rearranged form. The invention further relates to methods for assembling recombinant adenoviral DNAs. These methods include the steps of: (a) providing a first linearized DNA vector including a restriction site and a cos site and a second linearized DNA vector including the restriction site, an adenoviral nucleic acid molecule, and a cos site; and (b) ligating the first and second linearized DNA vectors, the ligation assembling a recombinant adenoviral DNA.

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

ACCESSION NUMBER: 2004:38117 USPATFULL

TITLE:

Self-rearranging DNA vectors

INVENTOR(S):

Seed, Brian, Boston, MA, UNITED STATES

Freeman, Mason Wright, Lincoln, MA, UNITED STATES

Kovtun, Alexander, Acton, MA, UNITED STATES Murakawa, Masahiro, Fukuoka City, JAPAN

Park, Eun-Chung, Washington, DC, UNITED STATES Wang, Xinzhong, Framingham, MA, UNITED STATES

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US 2004028653 A1 20040212 US 2003-384136 A1 20030307

PATENT INFORMATION:

20030307 (10)

RELATED APPLN. INFO.:

Continuation of Ser. No. WO 2001-US27682, filed on 7

Sep 2001, PENDING

DATE NUMBER _____

PRIORITY INFORMATION:

US 2000-246904P 20001108 (60)

US 2000-231053P 20000908 (60) DOCUMENT TYPE: Utility

FILE SEGMENT:

APPLICATION

LEGAL REPRESENTATIVE:

CLARK & ELBING LLP, 101 FEDERAL STREET, BOSTON, MA,

02110

NUMBER OF CLAIMS: EXEMPLARY CLAIM:

34 1

NUMBER OF DRAWINGS:

40 Drawing Page(s)

LINE COUNT:

3577

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

L7ANSWER 5 OF 27 USPATFULL on STN

Compositions and methods for making mutations in cell lines and animals ΤI The present invention is directed generally to reduction or inactivation AΒ of gene function or gene expression in cells in vitro and in multicellular organisms. The invention encompasses methods for mutating cells using a combination of mutagens, particularly wherein at least one mutagen is an insertional mutagen, to achieve homozygous gene mutation or mutation of multiple genes required cumulatively to achieve a phenotype to create knock-outs, knock-downs, and other modifications in the same cell. The invention is also directed to cells (and libraries thereof) and organisms created by the methods of the invention, including those in which at least one of the genes created by insertional mutagenesis is tagged by means of the insertion sequences thereby allowing identification of the mutated gene(s). The invention is also directed to libraries of mutated cells and their uses. The invention is also directed to methods of identifying mutations with

methods of the invention, in cells (and libraries thereof) and organisms, by means of the insertional tag.

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

ACCESSION NUMBER: 2004:24785 USPATFULL

Compositions and methods for making mutations in cell TITLE:

lines and animals

Harrington, John Joseph, Mentor, OH, UNITED STATES INVENTOR(S):

Jackson, Paul David, Shaker Heights, OH, UNITED STATES

Jiang, Li, Hudson, OH, UNITED STATES

Athersys, Inc., Cleveland, OH, 44115 (U.S. corporation) PATENT ASSIGNEE(S):

> NUMBER KIND DATE ______

PATENT INFORMATION:

US 2004018624 A1 20040129 US 2002-277612 A1 20021022 (10)

RELATED APPLN. INFO.:

Continuation-in-part of Ser. No. US 2002-196721, filed

on 15 Jul 2002, ABANDONED

NUMBER DATE _____

PRIORITY INFORMATION:

US 2001-336497P 20011022 (60)

DOCUMENT TYPE:

Utility

FILE SEGMENT:

APPLICATION

LEGAL REPRESENTATIVE: LAHIVE & COCKFIELD, 28 STATE STREET, BOSTON, MA, 02109

NUMBER OF CLAIMS: 170

1

EXEMPLARY CLAIM:

NUMBER OF DRAWINGS: 21 Drawing Page(s)

LINE COUNT:

4151

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

ANSWER 6 OF 27 USPATFULL on STN L7

Compositions and methods for the targeted insertion of a nucleotide TI

sequence of interest into the genome of a plant

Methods for the targeted integration of nucleotide sequences into a AB plant are provided. Transfer cassettes comprising nucleotide sequences of interest flanked by non-identical recombination sites are used to transform a plant comprising a target site. The target site contains at least a set of non-identical recombination sites corresponding to those on the transfer cassette. Exchange of the

nucleotide sequences flanked by the recombination sites is

effected by a recombinase.

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

ACCESSION NUMBER: 2004:3409 USPATFULL

TITLE: Compositions and methods for the targeted insertion of

a nucleotide sequence of interest into the genome of a

plant

INVENTOR (S): Baszczynski, Christopher L., Urbandale, IA, UNITED

Bowen, Benjamin A., Berkeley, CA, UNITED STATES Peterson, David J., Ames, IA, UNITED STATES Tagliani, Laura, Zionsville, IN, UNITED STATES

PATENT ASSIGNEE(S): Pioneer Hi-Bred International, Inc. (U.S. corporation)

> NUMBER KIND DATE ------US 2004003435 A1 20040101 US 2003-440030 A1 20030516 (10)

PATENT INFORMATION: RELATED APPLN. INFO.:

Continuation of Ser. No. US 1999-455050, filed on 6 Dec

1999, GRANTED, Pat. No. US 6624297 Division of Ser. No. US 1998-193502, filed on 17 Nov 1998, GRANTED, Pat. No.

US 6187994

DATE NUMBER

US 1997-65627P 19971118 (60) US 1997-65613P 19971118 (60) PRIORITY INFORMATION:

Utility DOCUMENT TYPE: APPLICATION FILE SEGMENT:

ALSTON & BIRD LLP, PIONEER HI-BRED INTERNATIONAL, INC., LEGAL REPRESENTATIVE:

BANK OF AMERICA PLAZA, 101 SOUTH TYRON STREET, SUITE

4000, CHARLOTTE, NC, 28280-4000

NUMBER OF CLAIMS: 22 EXEMPLARY CLAIM: 1

INVENTOR(S):

2 Drawing Page(s) NUMBER OF DRAWINGS:

1546 LINE COUNT:

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

L7 ANSWER 7 OF 27 USPATFULL on STN

Compositions and methods to reduce the complexity of transgene TI

integration into the genome of a plant

Methods for the targeted integration of nucleotide sequences into a AB plant are provided. Transfer cassettes comprising nucleotide sequences of interest flanked by non-identical recombination sites are used to transform a plant comprising a target site. The target site contains at least a set of non-identical recombination sites corresponding to those on the transfer cassette. Exchange of the nucleotide sequences flanked by the recombination sites is effected by a recombinase.

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

ACCESSION NUMBER: 2003:336227 USPATFULL

Compositions and methods to reduce the complexity of TITLE:

transgene integration into the genome of a plant Baszczynski, Christopher L., Urbandale, IA, UNITED

STATES

Bowen, Benjamin A., Berkeley, CA, UNITED STATES Peterson, David J., Ames, IA, UNITED STATES Tagliani, Laura, Zionsville, IN, UNITED STATES

PATENT ASSIGNEE(S): Pioneer Hi-Bred International, Inc. (U.S. corporation)

> NUMBER KIND DATE -----

US 2003237107 A1 20031225 US 2003-430908 A1 20030507 (10) PATENT INFORMATION: APPLICATION INFO.:

RELATED APPLN. INFO.: Continuation of Ser. No. US 1999-455050, filed on 6 Dec 1999, GRANTED, Pat. No. US 6624297 Division of Ser. No.

US 1998-193502, filed on 17 Nov 1998, GRANTED, Pat. No.

US 6187994

NUMBER DATE

PRIORITY INFORMATION:

US 1997-65627P 19971118 (60) US 1997-65613P 19971118 (60)

DOCUMENT TYPE: Utility FILE SEGMENT: APPLICATION

LEGAL REPRESENTATIVE: ALSTON & BIRD LLP, PIONEER HI-BRED INTERNATIONAL, INC.,

BANK OF AMERICA PLAZA, 101 SOUTH TYRON STREET, SUITE

4000, CHARLOTTE, NC, 28280-4000

NUMBER OF CLAIMS: 23 EXEMPLARY CLAIM:

NUMBER OF DRAWINGS: 2 Drawing Page(s)

LINE COUNT: 1556

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

ANSWER 8 OF 27 USPATFULL on STN L7

Compositions and methods for locating preferred integration sites within TΙ

the genome of a plant

Methods to find optimal integration sites within a plant genome are AB provided. More particularly, a plant is transformed with a target site having an expression cassette comprising a nucleotide sequence operably linked to a promoter active in the plant. The target site is flanked by non-identical recombination sites. Transformed protoplast, tissues, or whole plants can be tested to determine the levels of activity of the inserted gene. By comparison of cellular activities of the gene in different insertion sites, preferred integration sites may be found wherein the gene is expressed at high or acceptable levels. These plants can then be utilized with subsequent retargeting techniques to replace the nucleotide sequence with other genes or nucleotide sequences of interest contained in a transfer cassette.

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

ACCESSION NUMBER: 2003:320407 USPATFULL

Compositions and methods for locating preferred TITLE:

integration sites within the genome of a plant

Baszczynski, Christopher L., Urbandale, IA, UNITED INVENTOR(S):

STATES

Bowen, Benjamin A., Berkeley, CA, UNITED STATES Peterson, David J., Ames, IA, UNITED STATES Tagliani, Laura, Zionsville, IN, UNITED STATES

Pioneer Hi-Bred International, Inc. (U.S. corporation) PATENT ASSIGNEE(S):

> NUMBER KIND DATE

_____ US 2003226160 A1 20031204 US 2003-430907 A1 20030507 (10) PATENT INFORMATION:

APPLICATION INFO.:

Continuation of Ser. No. US 1999-455050, filed on 6 Dec RELATED APPLN. INFO.:

1999, GRANTED, Pat. No. US 6624297 Division of Ser. No. US 1998-193502, filed on 17 Nov 1998, GRANTED, Pat. No.

US 6187994

NUMBER DATE

US 1997-65627P 19971118 (60) US 1997-65613P 19971118 (60) PRIORITY INFORMATION:

DOCUMENT TYPE: Utility

APPLICATION FILE SEGMENT:

ALSTON & BIRD LLP, PIONEER HI-BRED INTERNATIONAL, INC., LEGAL REPRESENTATIVE:

BANK OF AMERICA PLAZA, 101 SOUTH TYRON STREET, SUITE

4000, CHARLOTTE, NC, 28280-4000

NUMBER OF CLAIMS: 17 EXEMPLARY CLAIM:

NUMBER OF DRAWINGS: 2 Drawing Page(s)

LINE COUNT: 1537

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

ANSWER 9 OF 27 USPATFULL on STN L7

ΤI Compositions and methods for making mutations in cell lines and animals The present invention is directed generally to reduction or inactivation AΒ of gene function or gene expression in cells in vitro and in multicellular organisms. The invention encompasses methods for mutating cells using a combination of mutagens, particularly wherein at least one mutagen is an insertional mutagen, to achieve homozygous gene mutation or mutation of multiple genes required cumulatively to achieve a phenotype to create knock-outs, knock-downs, and other modifications in the same cell. The invention is also directed to cells (and libraries thereof) and organisms created by the methods of the invention, including those in which at least one of the genes created by insertional mutagenesis is tagged by means of the insertion sequences thereby allowing identification of the mutated gene(s). The invention is also directed to libraries of mutated cells and their uses. The

invention is also directed to methods of identifying mutations with methods of the invention, in cells (and libraries thereof) and organisms, by means of the insertional tag.

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

ACCESSION NUMBER: 2003:318774 USPATFULL

Compositions and methods for making mutations in cell TITLE:

lines and animals

Harrington, John Joseph, Mentor, OH, UNITED STATES INVENTOR(S):

Jackson, Paul David, Shaker Heights, OH, UNITED STATES

Jiang, Li, Hudson, OH, UNITED STATES

PATENT ASSIGNEE(S): Athersys, Inc., Cleveland, OH (U.S. corporation)

NUMBER KIND DATE

PATENT INFORMATION: US 2003224519 A1 20031204 US 2003-345115 A1 20030115 (10)

RELATED APPLN. INFO.: Continuation-in-part of Ser. No. US 2002-277612, filed

on 22 Oct 2002, PENDING Continuation-in-part of Ser. No. US 2002-196721, filed on 15 Jul 2002, ABANDONED

DATE NUMBER -----

PRIORITY INFORMATION: US 2001-336497P 20011022 (60)

DOCUMENT TYPE: Utility APPLICATION FILE SEGMENT:

LEGAL REPRESENTATIVE: LAHIVE & COCKFIELD, 28 STATE STREET, BOSTON, MA, 02109

NUMBER OF CLAIMS: 210

NUMBER OF DRAWINGS: 21 Drawing Page(s)
LINE COUNT: 4363

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

ANSWER 10 OF 27 USPATFULL on STN L7

DNA sequences comprising gene transcription regulatory qualities and TI

methods for detecting and using such DNA sequences

The invention is concerned with the systematic elucidation and AΒ identification of regulatory sequences. The invention provides among others screenings and detection methods with which regulatory sequences can be identified. The invention further provides regulatory sequences and use thereof in various fields such as, but not limited to protein production, diagnostics, transgenic plants and animals, and the therapeutic field.

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

2003:283124 USPATFULL ACCESSION NUMBER:

TITLE:

DNA sequences comprising gene transcription regulatory qualities and methods for detecting and using such DNA

sequences

Otte, Arie Peter, Purmerend, NETHERLANDS INVENTOR(S):

Kruckeberg, Arthur Leo, Amsterdam, NETHERLANDS

NUMBER KIND DATE ______ US 2003199468 A1 20031023 US 2002-190312 A1 20020705 (10) PATENT INFORMATION: APPLICATION INFO.:

NUMBER DATE

______ PRIORITY INFORMATION: US 2001-303199P 20010705 (60)

DOCUMENT TYPE: Utility FILE SEGMENT: APPLICATION

LEGAL REPRESENTATIVE: TRASK BRITT, P.O. BOX 2550, SALT LAKE CITY, UT, 84110

NUMBER OF CLAIMS: 77 EXEMPLARY CLAIM:

NUMBER OF DRAWINGS: 65 Drawing Page(s)

LINE COUNT:

4902

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

ANSWER 11 OF 27 USPATFULL on STN L7

Compositions for the genetic modification of plants TI

Methods and compositions for the targeted integration of nucleotide AB sequences into a plant are provided. Particularly, the present invention is drawn to compositions comprising polynucleotide sequences having the following operably linked components: an intron, a nucleotide sequence of interest, and a terminator region, wherein the polynucleotide sequence comprises one or more recombination sites. The recombination sites are non-identical to one another and one of the recombination sites is contained within the intron.

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

INVENTOR(S):

ACCESSION NUMBER: 2003:253688 USPATFULL

TITLE:

Compositions for the genetic modification of plants Baszczynski, Christopher L., Urbandale, IA, United

States

Bowen, Benjamin A., Des Moines, IA, United States Peterson, David J., Ames, IA, United States Tagliani, Laura A., Ankeny, IA, United States

PATENT ASSIGNEE(S):

Pioneer Hi-Bred International, Inc., Des Moines, IA,

United States (U.S. corporation)

NUMBER KIND DATE

PATENT INFORMATION:

US 6624297 B1 20030923 US 1999-455050 19991206 (9)

RELATED APPLN. INFO.:

Division of Ser. No. US 1998-193502, filed on 17 Nov 1998, now patented, Pat. No. US 6187994, issued on 13

Feb 2001

NUMBER DATE -----

PRIORITY INFORMATION:

US 1997-65627P 19971118 (60) US 1997-65613P 19971118 (60)

DOCUMENT TYPE: Utility FILE SEGMENT: PRIMARY EXAMINER: Fox, David T.
ASSISTANT EXAMINER: Kruse, David H

Utility GRANTED LEGAL REPRESENTATIVE: Alston & Bird LLP

NUMBER OF CLAIMS:

15

EXEMPLARY CLAIM: NUMBER OF DRAWINGS:

2 Drawing Figure(s); 2 Drawing Page(s)

1599 LINE COUNT:

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

ANSWER 12 OF 27 USPATFULL on STN L7

Compositions and methods to reduce the complexity of transgene TI integration in the genome of a plant

AB Methods for reducing the complexity of integration of nucleotide sequences into a plant are provided. Transfer cassettes comprising nucleotide sequences of interest flanked by non-identical recombination sites are used to transform a plant comprising a target site. The target site contains at least a set of non-identical recombination sites corresponding to those on the transfer cassette. Exchange of the nucleotide sequences flanked by the recombination sites is effected by a recombinase.

CAS INDEXING IS AVAILABLE FOR THIS PATENT. 2003:149071 USPATFULL ACCESSION NUMBER:

Compositions and methods to reduce the complexity of TITLE:

transgene integration in the genome of a plant

Baszczynski, Christopher L., Urbandale, IA, United INVENTOR(S):

States

Bowen, Benjamin A., Des Moines, IA, United States

Peterson, David J., Ames, IA, United States Tagliani, Laura A., Ankeny, IA, United States

Pioneer Hi-Bred International, Inc., Des Moines, IA, PATENT ASSIGNEE(S):

United States (U.S. corporation)

KIND DATE NUMBER ______

PATENT INFORMATION:

US 6573425 B1 20030603 US 1999-439042 19991112

APPLICATION INFO.: RELATED APPLN. INFO.: (9)

Division of Ser. No. US 1998-193502, filed on 17 Nov

1998, now patented, Pat. No. US 6187994

NUMBER DATE _____

PRIORITY INFORMATION:

US 1997-65627P 19971118 (60) US 1997-65613P 19971118 (60)

DOCUMENT TYPE: Utility

GRANTED

FILE SEGMENT: PRIMARY EXAMINER: Fox, David T.
ASSISTANT EXAMINER: Kruse, David H

LEGAL REPRESENTATIVE: Alston & Bird LLP

NUMBER OF CLAIMS: EXEMPLARY CLAIM:

37

NUMBER OF DRAWINGS:

2 Drawing Figure(s); 2 Drawing Page(s)

LINE COUNT:

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

L7 ANSWER 13 OF 27 USPATFULL on STN

Universal stem cells ΤI

The subject invention pertains to materials and methods for preparing AΒ multi-potential stem cells having a pre-selected expression of MHC antiqens. Stem cells of the subject invention can be used to generate histocompatible tissues/organs for transplantation. The process of the subject invention comprises the use of targeting vectors capable of gene knockout, insertion of site-specific recombination cassettes, and the replacement of histocompatibility alleles in the stem cell. Novel knockout vectors are used to delete designated regions of one chromosome. Recombination cassette vectors are then used to delete the same region on the second chromosome and deposit a site-specific recombination cassette which can be utilized by replacement vectors for inserting the new MHC genes on the chromosome of the engineered cell. The subject invention also pertains to cells, tissues, and transgenic mammal prepared using the methods and materials of the invention.

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

ACCESSION NUMBER:

2003:147717 USPATFULL Universal stem cells

INVENTOR(S):

TITLE:

Lawman, Patricia, Chipley, FL, UNITED STATES Lawman, Michael J.P., Chipley, FL, UNITED STATES

NUMBER KIND DATE

PATENT INFORMATION: APPLICATION INFO .:

US 2003101465 A1 20030529 US 2002-186231 A1 20020628 (10)

RELATED APPLN. INFO.:

Continuation of Ser. No. US 1998-47769, filed on 25 Mar

1998, ABANDONED

NUMBER DATE

US 1997-42358P 19970325 (60)

PRIORITY INFORMATION: Utility DOCUMENT TYPE: APPLICATION FILE SEGMENT:

LEGAL REPRESENTATIVE: SALIWANCHIK LLOYD & SALIWANCHIK, A PROFESSIONAL

ASSOCIATION, 2421 N.W. 41ST STREET, SUITE A-1,

GAINESVILLE, FL, 326066669

18 NUMBER OF CLAIMS: EXEMPLARY CLAIM: 1

6 Drawing Page(s) NUMBER OF DRAWINGS:

1852 LINE COUNT:

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

ANSWER 14 OF 27 USPATFULL on STN

Compositions and methods for locating preferred integration sites within ΤI a plant genome

Methods to find optimal integration sites within a plant genome are AB provided. More particularly, a plant is transformed with a target site having an expression cassette comprising a nucleotide sequence operably linked to a promoter active in the plant. The target site is flanked by non-identical recombination sites, Transformed protoplast, tissues, or whole plants can be tested to determine the levels of activity of the inserted gene, By comparison of cellular activities of the gene in different insertion sites, preferred integration sites may be found wherein the gene is expressed at high or acceptable levels. These plants can then be utilized with subsequent retargeting techniques to replace the nucleotide sequence with other genes or nucleotide sequences of interest contained in a transfer cassette.

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

2003:109223 USPATFULL ACCESSION NUMBER:

Compositions and methods for locating preferred TITLE:

integration sites within a plant genome

Baszczynski, Christopher L., Urbandale, IA, United INVENTOR(S):

States

Bowen, Benjamin A., Des Moines, IA, United States Peterson, David J., Ames, IA, United States Tagliani, Laura A., Ankeny, IA, United States

Pioneer Hi-Bred International, Inc., Des Moines, IA, PATENT ASSIGNEE(S):

United States (U.S. corporation)

NUMBER KIND DATE _____ US 6552248 B1 20030422 US 1999-438239 19991112 (9) PATENT INFORMATION: APPLICATION INFO.:

Division of Ser. No. US 1998-193502, filed on 17 Nov RELATED APPLN. INFO.: 1998, now patented, Pat. No. US 6187994, issued on 13

Feb 2001

NUMBER DATE ______

PRIORITY INFORMATION:

US 1997-65627P 19971118 (60) US 1997-65613P 19971118 (60)

DOCUMENT TYPE: Utility FILE SEGMENT: GRANTED

PRIMARY EXAMINER: Fox, David T. ASSISTANT EXAMINER: Kruse, David H LEGAL REPRESENTATIVE: Alston & Bird LLP

49 NUMBER OF CLAIMS: EXEMPLARY CLAIM:

2 Drawing Figure(s); 2 Drawing Page(s) NUMBER OF DRAWINGS:

LINE COUNT:

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

1.7 ANSWER 15 OF 27 USPATFULL on STN

TT Methods for improving a photosynthetic carbon fixation enzyme

The invention relates to methods and compositions for generating, AB modifying, adapting, and optimizing polynucleotide sequences that encode proteins having photosynthetic carbon fixation activities, including Rubisco and Rubisco activase activities, which are useful for introduction into plant species, agronomically-important microorganisms, and other hosts, and related aspects.

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

ACCESSION NUMBER: 2003:106224 USPATFULL

TITLE:

Methods for improving a photosynthetic carbon fixation

enzyme

INVENTOR(S):

Zhu, Genhai, San Jose, CA, UNITED STATES

PATENT ASSIGNEE(S):

Maxygen, Inc. (U.S. corporation)

NUMBER KIND DATE ______

PATENT INFORMATION: APPLICATION INFO.:

US 2003073135 A1 20030417 US 2002-271019 A1 20021015 (10)

DATE NUMBER ------

PRIORITY INFORMATION:

US 2001-328871P 20011012 (60)

DOCUMENT TYPE: Utility FILE SEGMENT:

APPLICATION

LEGAL REPRESENTATIVE: MAXYGEN, INC., INTELLECTUAL PROPERTY DEPARTMENT, 515

GALVESTON DRIVE, RED WOOD CITY, CA, 94063

NUMBER OF CLAIMS: EXEMPLARY CLAIM:

20 1

LINE COUNT:

2569

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

ANSWER 16 OF 27 USPATFULL on STN L7

Modified ADP-glucose pyrophosphorylase for improvement and optimization TI

of plant phenotypes

The invention provides methods and compositions relating to AB sequence-shuffled variants of ADP-glucose pyrophosphorylase.

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

ACCESSION NUMBER: 2002:304117 USPATFULL

TITLE:

Modified ADP-glucose pyrophosphorylase for improvement

and optimization of plant phenotypes

INVENTOR (S):

Stemmer, Willem P. C., Los Gatos, CA, United States Subramanian, Venkiteswaran, San Diego, CA, United

PATENT ASSIGNEE(S):

Maxygen, Inc., Redwood City, CA, United States (U.S.

corporation)

NUMBER KIND DATE -----

PATENT INFORMATION:

US 6483011 B1 20021119 US 2000-721540 20001122 (9)

APPLICATION INFO.: RELATED APPLN. INFO.: Continuation of Ser. No. US 1999-437725, filed on 9 Nov

1999, now abandoned

NUMBER DATE ______

PRIORITY INFORMATION:

US 1998-107782P 19981110 (60)

DOCUMENT TYPE: FILE SEGMENT:

Utility GRANTED

PRIMARY EXAMINER: Horlick, Kenneth R. ASSISTANT EXAMINER: Strzelecka, Teresa

LEGAL REPRESENTATIVE: Townsend and Townsend and Crew LLP

NUMBER OF CLAIMS: 17 EXEMPLARY CLAIM:

1 EXEMPLARY CLAIM: 1

NUMBER OF DRAWINGS: 2 Drawing Figure(s); 2 Drawing Page(s)

LINE COUNT: 2975

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

ANSWER 17 OF 27 USPATFULL on STN L7

Alpha-tocopherol transfer protein knockout animals TI

This invention provides knockout animals comprising a disruption in one AB or both alleles of the gene encoding alpha-tocopherol transfer protein (TTP). The knockout animals provide good model systems for

atherosclerosis.

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

ACCESSION NUMBER: 2002:302526 USPATFULL

Alpha-tocopherol transfer protein knockout animals TITLE:

Farese, Robert V., JR., San Francisco, CA, UNITED INVENTOR(S):

Terasawa, Yuko, Campbell, CA, UNITED STATES Traber, Maret G., Corvallis, OR, UNITED STATES

The Regents of the University of California (U.S. PATENT ASSIGNEE(S):

corporation)

NUMBER KIND DATE _______ US 2002170080 A1 20021114 US 2001-1278 A1 20011101 (10) PATENT INFORMATION: APPLICATION INFO.:

NUMBER DATE ______

PRIORITY INFORMATION: US 2000-245302P 20001102 (60)

DOCUMENT TYPE: Utility
FILE SEGMENT: APPLICAT FILE SEGMENT: APPLICATION

LEGAL REPRESENTATIVE: QUINE INTELLECTUAL PROPERTY LAW GROUP, P.C., P O BOX

458, ALAMEDA, CA, 94501

NUMBER OF CLAIMS: 56
EXEMPLARY CLAIM: 1
NUMBER OF DRAWINGS: 4 Drawing Page(s)
LINE COUNT: 1912

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

ANSWER 18 OF 27 USPATFULL on STN L7

METHODS FOR OBTAINING A POLYNECLEOTIDE ENCODING A POLYPEPTIDE HAVING A TΤ RUBISCO ACTIVITY

The invention relates to methods and compositions for generating, AΒ modifying, adapting, and optimizing polynucleotide sequences that encode proteins having Rubisco biosynthetic enzyme activities which are useful for introduction into plant species, agronomically-important microorganisms, and other hosts, and related aspects.

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

2002:272896 USPATFULL ACCESSION NUMBER:

METHODS FOR OBTAINING A POLYNECLEOTIDE ENCODING A TITLE:

POLYPEPTIDE HAVING A RUBISCO ACTIVITY

STEMMER, WILLEM P. C., LOS GATOS, CA, UNITED STATES INVENTOR(S):

SUBRAMANIAN, VENKITSWARAN, SAN DIEGO, CA, UNITED STATES

ZHU, GENHAI, SUNNYVALE, CA, UNITED STATES LIU, LU, REDWOOD CITY, CA, UNITED STATES

SELIFONOV, SERGEY A., LOS ALTOS, CA, UNITED STATES

NUMBER KIND DATE PATENT INFORMATION: APPLICATION INFO.: US 2002151017 A1 20021017 US 1999-437726 A1 19991109 (9)

DATE NUMBER ______

PRIORITY INFORMATION:

US 1999-153093P 19990909 (60)

US 1998-107756P 19981110 (60)

DOCUMENT TYPE:

Utility APPLICATION

FILE SEGMENT: LEGAL REPRESENTATIVE:

TOWNSEND AND TOWNSEND AND CREW, LLP, TWO EMBARCADERO

CENTER, EIGHTH FLOOR, SAN FRANCISCO, CA, 94111-3834

NUMBER OF CLAIMS:

1

EXEMPLARY CLAIM: NUMBER OF DRAWINGS:

5 Drawing Page(s)

LINE COUNT:

3434

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

ANSWER 19 OF 27 USPATFULL on STN

Compositions and methods for the targeted removal of a nucleotide TI

sequence from the genome of a plant

Methods and compositions to remove a nucleotide sequence of interest in AB a plant and plant cell are provided. In particular the methods of the invention comprise providing a plant cell having stably incorporated into its genome a transfer cassette comprising a nucleotide sequence of interest flanked by non-identical recombination sites and introducing into the plant cell a chimeric RNA-DNA oligonucleotide molecule. The chimeric RNA-DNA oligonucleotide is capable of recognizing and implementing a nucleotide conversion in one of the non-identical recombination sites so as to create two identical recombination sites. An appropriate recombinase is provided which excises the sequences between the two identical recombination sites.

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

ACCESSION NUMBER:

PATENT ASSIGNEE(S):

2002:254223 USPATFULL

TITLE:

Compositions and methods for the targeted removal of a

nucleotide sequence from the genome of a plant Baszczynski, Christopher L., Urbandale, IA, United

INVENTOR(S):

States

Bowen, Benjamin A., Berkeley, CA, United States

Peterson, David J., Ames, IA, United States

Tagliani, Laura A., Zionsville, IN, United States Pioneer Hi-Bred International, Inc., Des Moines, IA,

United States (U.S. corporation)

NUMBER KIND DATĒ US 6458594 B1 20021001 US 1999-439158 19991112 (9) PATENT INFORMATION: APPLICATION INFO.:

RELATED APPLN. INFO.: Division of Ser. No. US 1998-193502, filed on 17 Nov 1998, now patented, Pat. No. US 6187994, issued on 13

Feb 2001

	NUMBER	DATE	
PRIORITY INFORMATION:	US 1997-65627P	19971118	(60)
	US 1997-65613P	19971118	(60)
	US 1998-98235P	19980828	(60)
	US 1997-65628P	19971118	(60)
DOCUMENT TYPE:	Utility		
FILE SEGMENT:	GRANTED		
PRIMARY EXAMINER:	Fox, David T.		

ASSISTANT EXAMINER: Kruse, David H LEGAL REPRESENTATIVE: Alston & Bird LLP

30 NUMBER OF CLAIMS: EXEMPLARY CLAIM: 1

2 Drawing Figure(s); 2 Drawing Page(s) NUMBER OF DRAWINGS:

1619 LINE COUNT:

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

ANSWER 20 OF 27 USPATFULL on STN L7

Compositions and methods to stack multiple nucleotide sequences of ΤI

interest in the genome of a plant

Methods and compositions for the stacking of multiple nucleotide AΒ sequences at precise locations in the genome of a plant or plant cell are provided, Specifically, transfer cassettes comprising nucleotide sequences of interest flanked by non-identical recombination sites are used to transform a plant comprising a target site. The target site contains at least a set of non-identical recombination sites corresponding to those on the transfer cassette. exchange of the nucleotide sequences flanked by the recombination sites is effected by a recombinase. The transfer cassettes and target sites are designed so as to allow for the stacking or ordering of nucleotide sequences at precise locations in the plant genome.

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

2002:246589 USPATFULL ACCESSION NUMBER:

Compositions and methods to stack multiple nucleotide TITLE:

sequences of interest in the genome of a plant

Baszczynski, Christopher L., Urbandale, IA, United INVENTOR(S):

States

Bowen, Benjamin A., Des Moines, IA, United States

Peterson, David J., Ames, IA, United States Tagliani, Laura A., Ankeny, IA, United States

Pioneer Hi-Bred International, Inc., Des Moines, IA, PATENT ASSIGNEE(S):

United States (U.S. corporation)

NUMBER KIND DATE _____

APPLICATION INFO.:

PATENT INFORMATION: US 6455315 B1 20020924 APPLICATION INFO.: US 1999-438874 19991112 (9)

RELATED APPLN. INFO.:

Division of Ser. No. US 1998-193502, filed on 17 Nov 1998, now patented, Pat. No. US 6187994, issued on 13

Feb 2000

NUMBER DATE _____

PRIORITY INFORMATION:

US 1997-65627P 19971118 (60) US 1997-65613P 19971118 (60)

DOCUMENT TYPE:

Utility GRANTED

FILE SEGMENT: FILE SEGMENT: GRANTED
PRIMARY EXAMINER: Fox, David T.
ASSISTANT EXAMINER: Kruse, David H

LEGAL REPRESENTATIVE: Alston & Bird LLP

44 NUMBER OF CLAIMS: EXEMPLARY CLAIM:

NUMBER OF DRAWINGS:

2 Drawing Figure(s); 2 Drawing Page(s)

1695 LINE COUNT:

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

ANSWER 21 OF 27 USPATFULL on STN L7

Modified ribulose 1,5-bisphosphate carboxylase/oxygenase for improvement ΤI

and optimization of plant phenotypes

The invention relates to methods and compositions for generating, AB modifying, adapting, and optimizing polynucleotide sequences that encode proteins having Rubisco biosynthetic enzyme activities which are useful for, introduction into plant species, agronomically-important microorganisms, and other hosts, and related aspects.

2001:183179 USPATFULL ACCESSION NUMBER:

Modified ribulose 1,5-bisphosphate TITLE:

carboxylase/oxygenase for improvement and optimization

of plant phenotypes

Stemmer, Willem P.C., Los Gatos, CA, United States INVENTOR (S):

Subramanian, Venkitswaran, San Diego, CA, United States

Zhu, Genhai, Sunnyvale, CA, United States Liu, Lu, Redwood City, CA, United States

Selifonov, Sergey A., Los Altos, CA, United States

Maxygen. Inc. (U.S. corporation) PATENT ASSIGNEE(S):

> NUMBER KIND DATE -----

US 2001032342 A1 20011018 US 2001-800123 A1 20010305 (9) PATENT INFORMATION: APPLICATION INFO.:

Continuation of Ser. No. US 1999-437726, filed on 9 Nov RELATED APPLN. INFO.:

1999, PENDING

DATE NUMBER ______

US 1999-153093P 19990909 (60) PRIORITY INFORMATION:

US 1998-107756P 19981110 (60)

DOCUMENT TYPE: Utility APPLICATION FILE SEGMENT:

LEGAL REPRESENTATIVE: LAW OFFICES OF JONATHAN ALAN QUINE, P O BOX 458,

ALAMEDA, CA, 94501

NUMBER OF CLAIMS: 26 EXEMPLARY CLAIM:

5 Drawing Page(s) NUMBER OF DRAWINGS:

3440 LINE COUNT:

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

ANSWER 22 OF 27 USPATFULL on STN Ь7

Compositions and methods for genetic modification of plants TI

Methods for the targeted integration of nucleotide sequences into a AB plant are provided. Transfer cassettes comprising nucleotide sequences of interest flanked by non-identical recombination sites are used to transform a plant comprising a target site. The target site contains at least a set of non-identical recombination sites corresponding to those on the transfer cassette. Exchange of the nucleotide sequences flanked by the recombination sites is

effected by a recombinase.

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

ACCESSION NUMBER: 2001:22439 USPATFULL

Compositions and methods for genetic modification of TITLE:

plants

Baszczynski, Christopher L., Urbandale, IA, United INVENTOR(S):

States

Bowen, Benjamin A., Des Moines, IA, United States

Peterson, David J., Ames, IA, United States Tagliani, Laura A., Ankeny, IA, United States

Pioneer Hi-Bred International, Inc., Des Moines, IA, PATENT ASSIGNEE(S):

United States (U.S. corporation)

NUMBER KIND DATE ______

PATENT INFORMATION: US 6187994 B1 20010213 APPLICATION INFO.: US 1998-193502 19981117 (9)

Continuation of Ser. No. US 1997-65627, filed on 18 Nov RELATED APPLN. INFO.: 1997 Continuation of Ser. No. US 1997-65613, filed on

18 Nov 1997

NUMBER DATE

______ US 1997-45121P 19970430 (60) PRIORITY INFORMATION:

DOCUMENT TYPE: Utility Granted FILE SEGMENT:

PRIMARY EXAMINER: McElwain, Elizabeth F. ASSISTANT EXAMINER: Mehta, Ashwin D. LEGAL REPRESENTATIVE: Alston & Bird LLP

NUMBER OF CLAIMS: 28 EXEMPLARY CLAIM:

2 Drawing Figure(s); 2 Drawing Page(s) NUMBER OF DRAWINGS:

LINE COUNT: 1628

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

ANSWER 23 OF 27 USPATFULL on STN 1.7

Mammalian artificial chromosomes and methods of using same ΤI

The present invention provides a mammalian artificial chromosome (MAC), AΒ comprising a centromere and a unique cloning site, said MAC containing less than 0.1% of the DNA present in a normal haploid genome of the mammalian cell from which the centromere was obtained. The invention further provides a MAC, wherein the unique cloning site is a nucleic acid sequence encoding a selectable marker. The invention also provides methods of preparing a MAC. In addition, the invention provides methods of stably expressing a selectable marker in a cell, comprising introducing a MAC containing the selectable marker into the cell. The invention also provides a cell containing a MAC expressing an exogenous nucleic acid sequence and a transgenic mammal expressing a selectable marker.

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

2000:138587 USPATFULL ACCESSION NUMBER:

Mammalian artificial chromosomes and methods of using TITLE:

Scheffler, Immo E., Del Mar, CA, United States INVENTOR(S):

The Regents of the University of California, Oakland, PATENT ASSIGNEE(S):

CA, United States (U.S. corporation)

NUMBER KIND DATE US 6133503 20001017 PATENT INFORMATION: US 1998-24472 19980217 (9) APPLICATION INFO.:

Division of Ser. No. US 1996-741406, filed on 29 Oct RELATED APPLN. INFO.:

1996, now patented, Pat. No. US 5721118

DATE NUMBER

US 1995-39256P 19951031 (60) PRIORITY INFORMATION: Utility

DOCUMENT TYPE: Granted FILE SEGMENT:

PRIMARY EXAMINER: Campell, Bruce R. ASSISTANT EXAMINER: Woitach, Joseph LEGAL REPRESENTATIVE: Campbell & Flores LLP

14 NUMBER OF CLAIMS: EXEMPLARY CLAIM:

6 Drawing Figure(s); 3 Drawing Page(s) NUMBER OF DRAWINGS:

1897 LINE COUNT:

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

ANSWER 24 OF 27 USPATFULL on STN L7

Mice homozygous for an inactivated α 1,3-galactosyl transferase TI

Human pre-formed xenoantibodies play an important role in the hyperacute AB rejection response in human xenotransplantation. Disclosed are materials and methods for removing or neutralizing such antibodies. Also disclosed are materials and methods for reducing or eliminating the epitopes in

the donor organs that are recognized by such antibodies. Such epitopes are formed as the result of activity by the enzyme $\alpha\text{-1,3}$ galactosyltransferase. The porcine gene encoding α -1,3 galactosyltransferase is disclosed, as are materials and methods for inactivating ("knocking out") the α -1,3 galactosyltransferase gene in mammalian cells and embryos. Included are nucleic acid constructs useful for inactivating the α -1,3 galactosyltransferase gene in a target cell. Also disclosed is a novel leukemia inhibitory factor (T-LIF) that is useful for maintenance of embryonic stem cells and primordial germ cells in culture.

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

ACCESSION NUMBER: 1998:157587 USPATFULL

TITLE:

Mice homozygous for an inactivated α

1,3-galactosyl transferase gene

INVENTOR (S):

d'Apice, Anthony J. F., Balwyn, Australia Pearse, Martin J., Mordialloc, Australia Robins, Allan J., Waterloo Corner, Australia Crawford, Robert J., West Lake Shores, Australia

Rathjen, Peter D., Blackwood, Australia

PATENT ASSIGNEE(S):

Bresatch Limited, Adelaide, Australia (non-U.S.

corporation)

St. Vincent's Hospital, Victoria, Australia (non-U.S.

corporation)

KIND DATE NUMBER _____

PATENT INFORMATION:

19981215

APPLICATION INFO.:

US 5849991 US 1995-378617 19950126

RELATED APPLN. INFO.:

Continuation-in-part of Ser. No. US 1994-188607, filed

on 27 Jan 1994, now abandoned

DOCUMENT TYPE:

Utility Granted

FILE SEGMENT: PRIMARY EXAMINER:

Crouch, Deborah

LEGAL REPRESENTATIVE: Fish & Richardson P.C., P.A.

NUMBER OF CLAIMS:

13

EXEMPLARY CLAIM:

47 Drawing Figure(s); 42 Drawing Page(s)

NUMBER OF DRAWINGS: LINE COUNT:

4190

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

ANSWER 25 OF 27 USPATFULL on STN L7

Mammalian artificial chromosomes and methods of using same TI

The present invention provides a mammalian artificial chromosome (MAC), ΔR comprising a centromere and a unique cloning site, said MAC containing less than 0.1% of the DNA present in a normal haploid genome of the mammalian cell from which the centromere was obtained. The invention further provides a MAC, wherein the unique cloning site is a nucleic acid sequence encoding a selectable marker. The invention also provides methods of preparing a MAC. In addition, the invention provides methods of stably expressing a selectable marker in a cell, comprising introducing a MAC containing the selectable marker into the cell. The invention also provides a cell containing a MAC expressing an exogenous nucleic acid sequence and a transgenic mammal expressing a selectable marker.

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

ACCESSION NUMBER:

1998:19593 USPATFULL

TITLE:

Mammalian artificial chromosomes and methods of using

INVENTOR(S):

Scheffler, Immo E., Del Mar, CA, United States

PATENT ASSIGNEE(S):

The Regents of the University of California, San Diego,

Alameda, CA, United States (U.S. corporation)

NUMBER KIND DATE _______

PATENT INFORMATION: 19980224 US 5721118 19961029 (8) US 1996-741406 APPLICATION INFO .:

DATE NUMBER ______

US 1995-39256P 19951031 (60) PRIORITY INFORMATION:

Utility DOCUMENT TYPE: FILE SEGMENT: Granted

PRIMARY EXAMINER: Chambers, Jasemine C. ASSISTANT EXAMINER: Schmuck, Jill D. LEGAL REPRESENTATIVE: Campbell & Flores LLP

NUMBER OF CLAIMS: 17 EXEMPLARY CLAIM: 2,7,17

6 Drawing Figure(s); 3 Drawing Page(s) NUMBER OF DRAWINGS:

LINE COUNT: 1797

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

ANSWER 26 OF 27 EMBASE COPYRIGHT 2004 ELSEVIER INC. ALL RIGHTS RESERVED. L7on STN

High-level heterologous gene expression in Saccharomyces cerevisiae from a TΙ

stable 2µm plasmid system.

The best candidate for a high-copy-number and mitotic stability AB expression system in yeast is the endogenous 2 µm plasmid. Nevertheless, derivatives of the 2µm plasmid typically exhibit lower copy numbers and require selection for adequate maintenance within cells. We report the construction and utilization of an efficient heterologous gene expression system containing a 4.5-kb inducible expression cassette inserted into the 2μm plasmid and selected in cells utilizing a carrier plasmid which is subsequently lost via FRT/Flp recombination.

The non-selectable 2µm plasmid, containing the cassette, was found to be stably maintained in cells, without selection, at high copy number. The dynamics of resolution and partitioning of this plasmid were analyzed during the course of 50 generations of growth under non-selective conditions. The heterologous lacZ reporter gene coding for

 β -galactosidase (β Gal) is driven by the hybrid,

galactose-inducible promoter GAL10::pMF α 1. Upon induction, β Gal was secreted into the periplasm and culture supernatant at levels which could be detected directly from Coomassie blue-stained SDS-PAGE. Furthermore, plasmid-containing cells could be maintained directly on rich YPD medium and identified either by utilizing XGal or by observing inhibition of colony growth on YPGal solid medium. The cassette was designed for direct, high-level, inducible expression of cloned genes downstream from the MF α 1 signal sequence, with or without a C-terminal lacZ fusion. This vector represents the first demonstration of a non-selectable, mitotically stable, episomal plasmid system capable of expressing recombinant proteins at high levels. By supplanting the need for synthetic medium, this system could provide both an efficient and

cost-effective means of generating recombinant protein at either the laboratory or large-scale level.

ACCESSION NUMBER: DOCUMENT NUMBER:

93310911 EMBASE 1993310911

TITLE:

High-level heterologous gene expression in Saccharomyces

cerevisiae from a stable 2µm plasmid system.

AUTHOR:

Ludwig D.L.; Ugolini S.; Bruschi C.V.

CORPORATE SOURCE: Microbiology Department, Internat. Ct. Gen.

Eng./Biotechnol., Padriciano 99, I-34012 Trieste, Italy

SOURCE:

Gene, (1993) 132/1 (33-40).

ISSN: 0378-1119 CODEN: GENED6

COUNTRY:

Netherlands

DOCUMENT TYPE: FILE SEGMENT:

Journal; Article 004 Microbiology

LANGUAGE:

English

SUMMARY LANGUAGE: English

L4

L5

L6

96 S FRT () FLP

82 S L5 AND RECOMBINATION

ANSWER 27 OF 27 BIOSIS COPYRIGHT 2004 BIOLOGICAL ABSTRACTS INC. on STN **L**7 High-level heterologous gene expression in Saccharomyces cerevisiae from a TIstable 2-mu-m plasmid system. The best candidate for a high-copy-number and mitotic stability AB expression system in yeast is the endogenous 2-mu-m plasmid. Nevertheless, derivatives of the 2-mu-m plasmid typically exhibit lower copy numbers and require selection for adequate maintenance within cells. We report the construction and utilization of an efficient heterologous gene expression system containing a 4.5-kb inducible expression cassette inserted into the 2-mu-m plasmid and selected in cells utilizing a carrier plasmid which is subsequently lost via FRT/Flp recombination. The non-selectable 2-mu-m plasmid, containing the cassette, was found to be stably maintained in cells, without selection, at high copy number. The dynamics of resolution and partitioning of this plasmid were analyzed during the course of 50 generations of growth under non-selective conditions. The heterologous lacZ reporter gene coding for beta-galactosidase (beta-Gal) is driven by the hybrid, galactose-inducible promoter GAL10::pMF-alpha-1. Upon induction, beta-Gal was secreted into the periplasm and culture supernatant at levels which could be detected directly from Coomassie blue-stained SDS-PAGE. Furthermore, plasmid-containing cells could be maintained directly on rich YPD medium and identified either by utilizing XGal or by observing inhibition of colony growth on YPGal solid medium. The cassette was designed for direct, high-level, inducible expression of cloned genes downstream from the MF-alpha-1 signal sequence, with or without a C-terminal lacZ fusion. This vector represents the first demonstration of a non-selectable, mitotically stable, episomal plasmid system capable of expressing recombinant proteins at high levels. By supplanting the need for synthetic medium, this system could provide both an efficient and cost-effective means of generating recombinant protein at either the laboratory or large-scale level. ACCESSION NUMBER: 1993:585676 BIOSIS PREV199497005046 DOCUMENT NUMBER: High-level heterologous gene expression in Saccharomyces TITLE: cerevisiae from a stable 2-mu-m plasmid system. Ludwig, Dale L.; Ugolini, Simone; Bruschi, Carlo V. AUTHOR (S): [Reprint author] Microbiol. Dep., Int. Centre Genetic Eng. and Biotechnol., CORPORATE SOURCE: Padriciano 99, I-34012 Trieste, Italy Gene (Amsterdam), (1993) Vol. 132, No. 1, pp. 33-40. SOURCE: CODEN: GENED6. ISSN: 0378-1119. DOCUMENT TYPE: Article English LANGUAGE: Entered STN: 28 Dec 1993 ENTRY DATE: Last Updated on STN: 28 Dec 1993 => s his 583163 HIS L8 => d his (FILE 'HOME' ENTERED AT 17:55:13 ON 29 MAR 2004) FILE 'MEDLINE, USPATFULL, DGENE, EMBASE, WPIDS, FSTA, CEN, BIOSIS' ENTERED AT 17:55:41 ON 29 MAR 2004 0 S FRT/FLP L16180 S FRT OR FLP L21238 S CRE LOX L3 0 S L2 AND STABLE RECOMBINATION

```
27 S L6 AND STABILITY
L7
         583163 S HIS
L8
=> s 17 and ARS
             0 L7 AND ARS
L9
=> s autonomous replicating sequence
   4 FILES SEARCHED...
           217 AUTONOMOUS REPLICATING SEQUENCE
L10
=> s 110 and 17
             0 L10 AND L7
1.11
=> s "1400 LNH-ST"
             0 "1400 LNH-ST"
L12
=> s plasmid and "1400"
          5601 PLASMID AND "1400"
L13
=> s XR or XD or XK
         16569 XR OR XD OR XK
L14
=> s 114 and 17
             0 L14 AND L7
L15
=> d his
      (FILE 'HOME' ENTERED AT 17:55:13 ON 29 MAR 2004)
     FILE 'MEDLINE, USPATFULL, DGENE, EMBASE, WPIDS, FSTA, CEN, BIOSIS'
     ENTERED AT 17:55:41 ON 29 MAR 2004
              0 S FRT/FLP
L1
           6180 S FRT OR FLP
L2
            1238 S CRE LOX
L3
              0 S L2 AND STABLE RECOMBINATION
L4
              96 S FRT () FLP
L_5
              82 S L5 AND RECOMBINATION
L6
              27 S L6 AND STABILITY
 L7
         583163 S HIS
 L8
              0 S L7 AND ARS
 L9
             217 S AUTONOMOUS REPLICATING SEQUENCE
 L10
               0 S L10 AND L7
 L11
               0 S "1400 LNH-ST"
 L12
            5601 S PLASMID AND "1400"
 L13
           16569 S XR OR XD OR XK
 L14
               0 S L14 AND L7
 L15
 => s 13 and 110
              7 L3 AND L10
 L16
 => d l16 ti abs ibib tot
 L16 ANSWER 1 OF 7 USPATFULL on STN
        Methods for isolating centromere DNA
 TТ
        The invention provides efficient methods for the isolation of
        centromeres from potentially any organism. The methods represents an
 AB
        advance over the prior art in that costly and labor intensive mapping
        programs are not required. Using the technique, methylated centromere
        DNA may be isolated from potentially any centromere in an organism. The
        technique is amenable to mass screenings employing use of arrays
        comprising libraries of DNA from a target species.
```

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

ACCESSION NUMBER:

2004:31144 USPATFULL

TITLE: INVENTOR(S): Methods for isolating centromere DNA Luo, Song, Chicago, IL, UNITED STATES

Copenhaver, Gregory, Oak Park, IL, UNITED STATES

Keith, Kevin, Chicago, IL, UNITED STATES Preuss, Daphne, Chicago, IL, UNITED STATES

KIND DATE NUMBER _______

PATENT INFORMATION: APPLICATION INFO.: RELATED APPLN. INFO.: US 2004023282 A1 20040205 US 2003-620924 A1 20030716 (10)

Division of Ser. No. US 2001-888220, filed on 22 Jun 2001, GRANTED, Pat. No. US 6649347

DATE NUMBER _____

PRIORITY INFORMATION:

US 2000-228793P 20000623 (60)

DOCUMENT TYPE: FILE SEGMENT:

Utility APPLICATION

LEGAL REPRESENTATIVE: BELL, BOYD & LLOYD LLC, P.O. Box 1135, Chicago, IL,

60690-1135

NUMBER OF CLAIMS: EXEMPLARY CLAIM:

119

NUMBER OF DRAWINGS:

40 Drawing Page(s)

LINE COUNT:

4102

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

L16 ANSWER 2 OF 7 USPATFULL on STN

Methods for generating or increasing revenues from crops

TIThe present invention provides methods of doing business and providing AB services. For example, methods of increasing the revenue of crops are provided. To this end, the method includes the use of a nucleic acid sequences of plant centromeres. This will permit construction of stably inherited recombinant DNA constructs and mini chromosomes which can serve as vectors for the construction of transgenic plant and animal cells.

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

ACCESSION NUMBER:

2003:189578 USPATFULL

TITLE:

Methods for generating or increasing revenues from

INVENTOR (S):

Copenhaver, Gregory, Chapel Hill, NC, UNITED STATES

Keith, Kevin, Chicago, IL, UNITED STATES Preuss, Daphne, Chicago, IL, UNITED STATES

NUMBER KIND DATE

PATENT INFORMATION: APPLICATION INFO.: RELATED APPLN. INFO.: US 2003131372 A1 20030710 US 2002-170944 A1 20020612 (10)

Continuation-in-part of Ser. No. US 2000-553231, filed on 19 Apr 2000, PENDING Continuation of Ser. No. US 1998-90051, filed on 3 Jun 1998, GRANTED, Pat. No. US

6156953 Continuation-in-part of Ser. No. US 2000-531120, filed on 17 Mar 2000, PENDING

			NUMBER	DATE	
PRIORITY	INFORMATION:	US	1997-48451P	19970603	(60)
		US	1998-73741P	19980205	(60)
		US	1999-125219P	19990318	(60)
		US	1999-127409P	19990401	(60)
		US	1999-134770P	19990518	(60)
		US	1999-153584P	19990913	(60)
		US	1999-154603P	19990917	(60)
		US	1999-172493P	19991216	(60)

DOCUMENT TYPE:

Utility

FILE SEGMENT:

APPLICATION

LEGAL REPRESENTATIVE: Bell, Boyd & Lloyd LLC, P.O. Box 1135, Chicago, IL,

60690-1135

NUMBER OF CLAIMS: EXEMPLARY CLAIM:

NUMBER OF DRAWINGS:

46 Drawing Page(s)

LINE COUNT:

4575

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

L16 ANSWER 3 OF 7 USPATFULL on STN

Plant centromere compositions ΤT

The present invention provides for the nucleic acid sequences of plant AΒ centromeres. This will permit construction of stably inherited recombinant DNA constructs and minichromosomes which can serve as vectors for the construction of transgenic plant and animal cells.

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

ACCESSION NUMBER: 2003:180731 USPATFULL

TITLE:

Plant centromere compositions

INVENTOR (S):

Mach, Jennifer, Chicago, IL, UNITED STATES Zieler, Helge, Chicago, IL, UNITED STATES Jin, RongGuan, Chicago, IL, UNITED STATES Keith, Kevin, Chicago, IL, UNITED STATES

Copenhaver, Gregory, Chapel Hill, NC, UNITED STATES

Preuss, Daphne, Chicago, IL, UNITED STATES

KIND DATE NUMBER -----

PATENT INFORMATION:

APPLICATION INFO.:

US 2003124561 A1 20030703 US 2002-170912 A1 20020612

RELATED APPLN. INFO.:

Continuation-in-part of Ser. No. US 2000-553231, filed on 19 Apr 2000, PENDING Continuation of Ser. No. US 1998-90051, filed on 3 Jun 1998, GRANTED, Pat. No. US

(10)

6156953 Continuation-in-part of Ser. No. US 2000-531120, filed on 17 Mar 2000, PENDING

DATE NUMBER _____

PRIORITY INFORMATION:

US 1997-48451P 19970603 (60) US 1998-73741P 19980205 (60)

US 1997-48451P 19970603 (60)
US 1998-73741P 19980205 (60)
US 1999-125219P 19990318 (60)
US 1999-127409P 19990401 (60)
US 1999-134770P 19990518 (60)
US 1999-153584P 19990913 (60)
US 1999-154603P 19990917 (60)
US 1999-172493P 19991216 (60)

DOCUMENT TYPE:

Utility

FILE SEGMENT:

APPLICATION

LEGAL REPRESENTATIVE:

BELL, BOYD & LLOYD, LLC, PO BOX 1135, CHICAGO, IL,

60690-1135

NUMBER OF CLAIMS:

127

EXEMPLARY CLAIM:

1

NUMBER OF DRAWINGS:

46 Drawing Page(s)

LINE COUNT: 4478

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

L16 ANSWER 4 OF 7 USPATFULL on STN

Methods for isolating centromere DNA

ΤI The invention provides efficient methods for the isolation of AB centromeres from potentially any organism. The methods represents an advance over the prior art in that costly and labor intensive mapping programs are not required. Using the technique, methylated centromere DNA may be isolated from potentially any centromere in an organism. The technique is amenable to mass screenings employing use of arrays comprising libraries of DNA from a target species.

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

ACCESSION NUMBER: 2002:227900 USPATFULL

TITLE:

Methods for isolating centromere DNA

INVENTOR (S):

Luo, Song, Chicago, IL, UNITED STATES Copenhaver, Gregory, Oak Park, IL, UNITED STATES

Keith, Kevin, Chicago, IL, UNITED STATES Preuss, Daphne, Chicago, IL, UNITED STATES

PATENT ASSIGNEE(S):

University of Chicago (U.S. corporation)

NUMBER KIND DATE ______ US 2002123053 A1 20020905 US 6649347 B2 20031118 US 2001-888220 A1 20010622 (9) PATENT INFORMATION: APPLICATION INFO.:

> NUMBER DATE _____

PRIORITY INFORMATION: US 2000-228793P 20000623 (60)

DOCUMENT TYPE: Utility FILE SEGMENT: APPLICATION

LEGAL REPRESENTATIVE: BELL, BOYD & LLOYD, LLC, PO BOX 1135, CHICAGO, IL,

60690-1135

NUMBER OF CLAIMS: 119
EXEMPLARY CLAIM: 1
NUMBER OF DRAWINGS: 40 Drawing Page(s)
LINE COUNT: 4078

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

L16 ANSWER 5 OF 7 USPATFULL on STN

TI

Telomerase compositions and methods

AB

Disclosed are various methods, compositions and screening assays connected with telomerase, including genes encoding the template RNA of S. cerevisiae telomerase and various telomerase-associated polypeptides.

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

ACCESSION NUMBER: 2002:108815 USPATFULL

TITLE:

Telomerase compositions and methods

INVENTOR(S):

Gottschling, Daniel E., Chicago, IL, United States

Singer, Miriam S., Chicago, IL, United States

PATENT ASSIGNEE(S):

Arch Development, Chicago, IL, United States (U.S.

corporation)

NUMBER KIND DATE _____

PATENT INFORMATION: US 6387619 B1 20020514 APPLICATION INFO.: US 1999-345294 19990630 (9)

RELATED APPLN. INFO.: Division of Ser. No. US 1997-938534, filed on 26 Sep 1997, now patented, Pat. No. US 5916752 Division of Ser. No. US 1995-431080, filed on 28 Apr 1995, now patented, Pat. No. US 5698686 Division of Ser. No. US 345294 Continuation-in-part of Ser. No. US 1994-326781,

filed on 20 Oct 1994, now abandoned

DOCUMENT TYPE:

Utility GRANTED

FILE SEGMENT: PRIMARY EXAMINER: Fredman, Jeffrey

LEGAL REPRESENTATIVE: Fulbright & Jaworski, LLP

NUMBER OF CLAIMS:

10

EXEMPLARY CLAIM:

1 NUMBER OF DRAWINGS: 15 Drawing Figure(s); 11 Drawing Page(s) LINE COUNT: 6648

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

L16 ANSWER 6 OF 7 USPATFULL on STN

Telomerase screening methods TΤ

Disclosed are various methods, compositions and screening assays AB connected with telomerase, including genes encoding the template RNA of S. cerevisiae telomerase and various telomerase-associated polypeptides.

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

ACCESSION NUMBER:

1999:72446 USPATFULL

TITLE:

Telomerase screening methods

INVENTOR(S):

Gottschling, Daniel E., Chicago, IL, United States

Singer, Miriam S., Chicago, IL, United States

PATENT ASSIGNEE(S):

Arch Development Corporation, Chicago, IL, United

States (U.S. corporation)

KIND DATE NUMBER _____

PATENT INFORMATION:

US 5916752 19990629 US 1997-938534 19970926

APPLICATION INFO.:

19970926 (8)

RELATED APPLN. INFO.:

Division of Ser. No. US 1995-431080, filed on 28 Apr 1995, now patented, Pat. No. US 5698686 which is a

continuation-in-part of Ser. No. US 1994-326781, filed

on 20 Oct 1994, now abandoned

DOCUMENT TYPE:

Utility

FILE SEGMENT:

Granted Fredman, Jeffrey

PRIMARY EXAMINER:

LEGAL REPRESENTATIVE: Arnold, White & Durkee

NUMBER OF CLAIMS:

EXEMPLARY CLAIM:

NUMBER OF DRAWINGS:

15 Drawing Figure(s); 15 Drawing Page(s)

LINE COUNT:

7780

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

L16 ANSWER 7 OF 7 USPATFULL on STN

Yeast telomerase compositions ΤI

Disclosed are various methods, compositions and screening assays AB

connected with telomerase, including genes encoding the template RNA of S. cerevisiae telomerase and various telomerase-associated polypeptides.

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

ACCESSION NUMBER: 97:118172 USPATFULL

TITLE:

Yeast telomerase compositions

INVENTOR (S):

Gottschling, Daniel E., Chicago, IL, United States

Singer, Miriam S., Chicago, IL, United States

PATENT ASSIGNEE(S):

Arch Development Corporation, Chicago, IL, United

States (U.S. corporation)

KIND DATE NUMBER

PATENT INFORMATION: US 5698686 APPLICATION INFO.: US 1995-431080 19971216

19950428 (8)

RELATED APPLN. INFO.: Continuation-in-part of Ser. No. US 1994-326781, filed

on 20 Oct 1994, now abandoned

DOCUMENT TYPE: FILE SEGMENT:

Utility Granted

PRIMARY EXAMINER: PRIMARY EXAMINER: Jones, W. Gary ASSISTANT EXAMINER: Fredman, Jeffrey

Jones, W. Gary

LEGAL REPRESENTATIVE: Arnold, White & Durkee

NUMBER OF CLAIMS:

71

EXEMPLARY CLAIM:

NUMBER OF DRAWINGS:

15 Drawing Figure(s); 15 Drawing Page(s)

LINE COUNT:

7319

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

(FILE 'HOME' ENTERED AT 17:55:13 ON 29 MAR 2004)

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FILE 'MEDLINE, USPATFULL, DGENE, EMBASE, WPIDS, FSTA, CEN, BIOSIS' ENTERED AT 17:55:41 ON 29 MAR 2004
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L1 0 S FRT/FLP L2 6180 S FRT OR FLP

L3 1238 S CRE LOX

L4 0 S L2 AND STABLE RECOMBINATION

L5 96 S FRT () FLP

L6 82 S L5 AND RECOMBINATION

L7 27 S L6 AND STABILITY

L8 583163 S HIS

L9 0 S L7 AND ARS

L10 217 S AUTONOMOUS REPLICATING SEQUENCE

L11 0 S L10 AND L7

L12 0 S "1400 LNH-ST"

L13 5601 S PLASMID AND "1400"

L14 16569 S XR OR XD OR XK

L15 0 S L14 AND L7

L16 7 S L3 AND L10

=> s 12 and 110

L17 8 L2 AND L10

=> d l17 ti abs ibib tot

L17 ANSWER 1 OF 8 USPATFULL on STN

TI Methods for isolating centromere DNA

The invention provides efficient methods for the isolation of centromeres from potentially any organism. The methods represents an advance over the prior art in that costly and labor intensive mapping programs are not required. Using the technique, methylated centromere DNA may be isolated from potentially any centromere in an organism. The technique is amenable to mass screenings employing use of arrays comprising libraries of DNA from a target species.

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

ACCESSION NUMBER: 2004:31144 USPATFULL

ACCESSION NUMBER: 2004:31144 USPAIRUDD
TITLE: Methods for isolating centromere DNA

INVENTOR(S): Luo, Song, Chicago, IL, UNITED STATES

Copenhaver, Gregory, Oak Park, IL, UNITED STATES

Keith, Kevin, Chicago, IL, UNITED STATES Preuss, Daphne, Chicago, IL, UNITED STATES

APPLICATION INFO.: US 2003-620924 A1 20030716 (10)
RELATED APPLN. INFO.: Division of Ser. No. US 2001-888220, filed on 22 Jun

2001, GRANTED, Pat. No. US 6649347

NUMBER DATE

PRIORITY INFORMATION: US 2000-228793P 20000623 (60)

DOCUMENT TYPE: Utility
FILE SEGMENT: APPLICATION

LEGAL REPRESENTATIVE: BELL, BOYD & LLOYD LLC, P.O. Box 1135, Chicago, IL,

60690-1135

NUMBER OF CLAIMS: 119
EXEMPLARY CLAIM: 1

PATENT INFORMATION:

NUMBER OF DRAWINGS: 40 Drawing Page(s)

4102 LINE COUNT:

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

L17 ANSWER 2 OF 8 USPATFULL on STN

Methods for generating or increasing revenues from crops TI

The present invention provides methods of doing business and providing AB services. For example, methods of increasing the revenue of crops are provided. To this end, the method includes the use of a nucleic acid sequences of plant centromeres. This will permit construction of stably inherited recombinant DNA constructs and mini chromosomes which can serve as vectors for the construction of transgenic plant and animal cells.

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

ACCESSION NUMBER: 2003:189578 USPATFULL

TITLE:

Methods for generating or increasing revenues from

crops

INVENTOR(S):

Copenhaver, Gregory, Chapel Hill, NC, UNITED STATES

Keith, Kevin, Chicago, IL, UNITED STATES Preuss, Daphne, Chicago, IL, UNITED STATES

NUMBER KIND DATE -----US 2003131372 A1 20030710 US 2002-170944 A1 20020612 PATENT INFORMATION: (10)

APPLICATION INFO.: RELATED APPLN. INFO.:

Continuation-in-part of Ser. No. US 2000-553231, filed

on 19 Apr 2000, PENDING Continuation of Ser. No. US 1998-90051, filed on 3 Jun 1998, GRANTED, Pat. No. US

6156953 Continuation-in-part of Ser. No. US 2000-531120, filed on 17 Mar 2000, PENDING

DATE NUMBER _____ US 1997-48451P 19970603 (60) US 1998-73741P 19980205 (60) PRIORITY INFORMATION: US 1998-73741P 19980205 (60)
US 1999-125219P 19990318 (60)
US 1999-127409P 19990401 (60)
US 1999-134770P 19990518 (60)
US 1999-153584P 19990913 (60)
US 1999-154603P 19990917 (60)
US 1999-172493P 19991216 (60)

DOCUMENT TYPE:

Utility

FILE SEGMENT:

APPLICATION

LEGAL REPRESENTATIVE: Bell, Boyd & Lloyd LLC, P.O. Box 1135, Chicago, IL,

60690-1135

NUMBER OF CLAIMS:

91

EXEMPLARY CLAIM:

1

NUMBER OF DRAWINGS:

46 Drawing Page(s)

LINE COUNT:

4575

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

L17 ANSWER 3 OF 8 USPATFULL on STN

Plant centromere compositions ΤI

The present invention provides for the nucleic acid sequences of plant AB centromeres. This will permit construction of stably inherited recombinant DNA constructs and minichromosomes which can serve as vectors for the construction of transgenic plant and animal cells.

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

ACCESSION NUMBER:

2003:180731 USPATFULL

TITLE:

Plant centromere compositions

INVENTOR (S):

Mach, Jennifer, Chicago, IL, UNITED STATES Zieler, Helge, Chicago, IL, UNITED STATES Jin, RongGuan, Chicago, IL, UNITED STATES

Keith, Kevin, Chicago, IL, UNITED STATES Copenhaver, Gregory, Chapel Hill, NC, UNITED STATES Preuss, Daphne, Chicago, IL, UNITED STATES

	NUMBER	KIND	DATE
PATENT INFORMATION: APPLICATION INFO.:	US 2003124561 US 2002-170912	A1 A1	20030703
APPLICATION INFO.:	05 2002 170712		2002002

Continuation-in-part of Ser. No. US 2000-553231, filed on 19 Apr 2000, PENDING Continuation of Ser. No. US 1998-90051, filed on 3 Jun 1998, GRANTED, Pat. No. US 6156953 Continuation-in-part of Ser. No. US

(10)

6156953 Continuation-in-part of Ser. No. US 2000-531120, filed on 17 Mar 2000, PENDING

			NUMBER	DATE	
PRIORITY	INFORMATION:	US	1997-48451P	19970603	(60)
		US	1998-73741P	19980205	(60)
		US	1999-125219P	19990318	(60)
		US	1999-127409P	19990401	(60)
		US	1999-134770P	19990518	(60)
		US	1999-153584P	19990913	(60)
		US	1999-154603P	19990917	(60)
			1999-172493P	19991216	(60)
DOCUMENT	TYPE:		ility		•

DOCUMENT TYPE: Utility
FILE SEGMENT: APPLICATION

LEGAL REPRESENTATIVE: BELL, BOYD & LLOYD, LLC, PO BOX 1135, CHICAGO, IL,

60690-1135

NUMBER OF CLAIMS: 127 EXEMPLARY CLAIM: 1

RELATED APPLN. INFO.:

NUMBER OF DRAWINGS: 46 Drawing Page(s)

LINE COUNT: 4478

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

L17 ANSWER 4 OF 8 USPATFULL on STN

TI Rapid creation of gene targeting vectors using homologous recombination in yeast

The present invention provides methods of preparing gene targeted mammalian cells having a targeted gene mutation methods of making gene targeted mice, and gene targeting vectors that are useful in these methods.

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

ACCESSION NUMBER: 2003:134098 USPATFULL

TITLE: Rapid creation of gene targeting vectors using

homologous recombination in yeast

INVENTOR(S): Fisher, Katherine E., Old Lyme, CT, UNITED STATES
Reaume, Andrew G., Waterford, CT, UNITED STATES

DOCUMENT TYPE: Utility
FILE SEGMENT: APPLICATION

LEGAL REPRESENTATIVE: Gregg C. Benson, Pfizer Inc., Patent Department, MS

4159, Eastern Point Road, Groton, CT, 06340

NUMBER OF CLAIMS: 20 EXEMPLARY CLAIM: 1

NUMBER OF DRAWINGS: 2 Drawing Page(s)

LINE COUNT: 1249

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

L17 ANSWER 5 OF 8 USPATFULL on STN

Methods for isolating centromere DNA

The invention provides efficient methods for the isolation of centromeres from potentially any organism. The methods represents an advance over the prior art in that costly and labor intensive mapping programs are not required. Using the technique, methylated centromere DNA may be isolated from potentially any centromere in an organism. The technique is amenable to mass screenings employing use of arrays comprising libraries of DNA from a target species.

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

2002:227900 USPATFULL ACCESSION NUMBER:

TITLE:

Methods for isolating centromere DNA

INVENTOR(S):

TТ

 $\mathbf{A}\mathbf{B}$

Luo, Song, Chicago, IL, UNITED STATES Copenhaver, Gregory, Oak Park, IL, UNITED STATES

Keith, Kevin, Chicago, IL, UNITED STATES Preuss, Daphne, Chicago, IL, UNITED STATES

PATENT ASSIGNEE(S):

University of Chicago (U.S. corporation)

KIND DATE NUMBER _____ US 2002123053 A1 20020905 US 6649347 B2 20031118 US 2001-888220 A1 20010622 (9) PATENT INFORMATION: APPLICATION INFO.:

> NUMBER DATE ______

PRIORITY INFORMATION:

US 2000-228793P 20000623 (60)

DOCUMENT TYPE:

Utility

FILE SEGMENT:

APPLICATION

LEGAL REPRESENTATIVE: BELL, BOYD & LLOYD, LLC, PO BOX 1135, CHICAGO, IL,

60690-1135

NUMBER OF CLAIMS: 119
EXEMPLARY CLAIM: 1
NUMBER OF DRAWINGS: 40 Drawing Page(s)
4078

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

L17 ANSWER 6 OF 8 USPATFULL on STN

Methods for the production of gelatin and full-length triple helical TI

collagen in recombinant cells

Methods are disclosed for simplified recombinant production of fibrillar AΒ collagens. DNAs encoding fibrillar collagen monomers lacking the N propeptide, the C propeptide, or both propeptides are introduced into recombinant host cells and expressed. Trimeric collagen is recovered from the recombinant host cells.

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

ACCESSION NUMBER: 2002:194714 USPATFULL

TITLE:

INVENTOR(S):

Methods for the production of gelatin and full-length

triple helical collagen in recombinant cells Olsen, David R., Menlo Park, CA, United States Chang, Robert, Hillsborough, CA, United States McMullin, Hugh, Menlo Park, CA, United States Hitzeman, Ronald A., Pacifica, CA, United States Chisholm, George, San Mateo, CA, United States Cohesion Technologies, Inc., Palo Alto, CA, United

PATENT ASSIGNEE(S):

States (U.S. corporation)

Genotypes, Inc., Pacifica, CA, United States (U.S.

corporation)

KIND DATE NUMBER _____ US 6428978 B1 20020806 US 1999-289578 19990409 (9) PATENT INFORMATION: APPLICATION INFO.:

NUMBER DATE -----

US 1998-84828P 19980508 (60) PRIORITY INFORMATION:

Utility DOCUMENT TYPE:

FILE SEGMENT: GRANTED
PRIMARY EXAMINER: Whisenant, Ethan C. LEGAL REPRESENTATIVE: Osman, Richard Aron

NUMBER OF CLAIMS: 31 EXEMPLARY CLAIM: 1

14 Drawing Figure(s); 14 Drawing Page(s) NUMBER OF DRAWINGS:

1945 LINE COUNT:

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

L17 ANSWER 7 OF 8 USPATFULL on STN

Recombinant gelatin and full-length triple helical collagen ΤI

Methods are disclosed for simplified recombinant production of fibrillar AB collagens. DNAs encoding fibrillar collagen monomers lacking the N propeptide, the C propeptide, or both propeptides are introduced into recombinant host cells and expressed. Trimeric collagen is recovered from the recombinant host cells.

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

2002:160549 USPATFULL ACCESSION NUMBER:

Recombinant gelatin and full-length triple helical TITLE:

collagen

Olsen, David R., Menlo Park, CA, United States Chang, Robert, Hillsborough, CA, United States INVENTOR (S):

McMullin, Hugh, Menlo Park, CA, United States Hitzeman, Ronald A., Pacifica, CA, United States Chisholm, George, San Mateo, CA, United States

Cohesion Technologies, Inc., Palo Alto, CA, United PATENT ASSIGNEE(S):

States (U.S. corporation)

Genotypes, Inc., Pacifica, CA, United States (U.S.

(9)

corporation)

NUMBER KIND DATE ______ US 6413742 B1 20020702 US 2000-585887 20000531 20000531

APPLICATION INFO .: Division of Ser. No. US 1999-289578, filed on 9 Apr RELATED APPLN. INFO.:

1999

DATE NUMBER ______

US 1998-84828P 19980508 (60) PRIORITY INFORMATION:

Utility DOCUMENT TYPE:

FILE SEGMENT: GRANTED PRIMARY EXAMINER: Whisenant, Ethan C. LEGAL REPRESENTATIVE: Osman, Richard Aron

29 NUMBER OF CLAIMS: EXEMPLARY CLAIM:

PATENT INFORMATION:

14 Drawing Figure(s); 14 Drawing Page(s) NUMBER OF DRAWINGS:

LINE COUNT: 1573

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

L17 ANSWER 8 OF 8 WPIDS COPYRIGHT 2004 THOMSON DERWENT on STN

Recombinant DNA construct comprising a plant centromere, useful for TТ producing stably inherited michrosomes which can serve as vectors for the construction of transgenic plant and animal cells.

2000-587529 [55] AΝ WPIDS

1999-080832 [07]; 2000-587463 [55]; 2003-829605 [77] CR

WO 200055325 A UPAB: 20031128 AB NOVELTY - A recombinant DNA construct (I) comprising a plant centromere, is new.

DETAILED DESCRIPTION - INDEPENDENT CLAIMS are also included for the following:

(1) a minichromosome vector comprising a plant centromere and telomere sequence;

(2) a cell transformed with a recombinant DNA construct comprising a plant centromere;

(3) a plant comprising the cell of (2);

(4) a method of preparing a transgenic plant cell comprising contacting a starting plant cell with a recombinant DNA construct comprising a plant centromere;

(5) a transgenic plant comprising a minichromosome vector;

- (6) a method for producing a minichromosome vector comprising:
- (a) obtaining a first and second vector, where either comprises a selectable or screenable marker, an origin of replication, a telomere, and a plant centromere, and where the vectors comprise a site for site-specific recombination; and

(b) contacting the first vector with the second vector;

(7) a method of screening a candidate centromere sequence for plant centromere activity comprising:

(a) obtaining an isolated nucleic acid sequence comprising a candidate centromere sequence;

(b) integratively transforming plant cells with the isolated nucleic

(c) screening for centromere activity;

(8) a recombinant DNA construct comprising an Arabidopsis polyubiquitin 11 promoter comprising a defined 2000 bp sequence (given in the specification), or its fragments;

(9) a recombinant DNA construct comprising an Arabidopsis 40S ribosomal protein S16 promoter comprising a defined 2000 bp sequence

(given in the specification);

(10) a recombinant DNA construct comprising an Arabidopsis polyubiquitin 11 3' regulatory sequence comprising a defined 2001 bp sequence (given in the specification); and

(11) a recombinant DNA construct comprising an Arabidopsis 40S ribosomal protein S16 3' regulatory sequence comprising a defined 2000 bp

sequence (given in the specification).

USE - The constructs are useful for producing stably inherited michrosomes which can serve as vectors for the construction of transgenic plant and animal cells expressing selected proteins such as hormones, enzymes, interleukins, clotting factors, cytokines, antibodies, and growth factors.

Dwq.0/23

ACCESSION NUMBER:

2000-587529 [55] WPIDS 1999-080832 [07]; 2000-587463 [55]; 2003-829605 [77] CROSS REFERENCE:

C2000-175305 DOC. NO. CPI:

Recombinant DNA construct comprising a plant centromere, TITLE: useful for producing stably inherited michrosomes which can serve as vectors for the construction of transgenic

plant and animal cells.

B04 C06 D16 P13 DERWENT CLASS:

COPENHAVER, G; KEITH, K; PREUSS, D; JIN, R; MACH, J; INVENTOR(S):

(UYCH-N) UNIV CHICAGO; (COPE-I) COPENHAVER G; (JINR-I) PATENT ASSIGNEE(S):

JIN R; (KEIT-I) KEITH K; (MACH-I) MACH J; (PREU-I) PREUSS

D; (ZIEL-I) ZIELER H

91 COUNTRY COUNT:

PATENT INFORMATION:

WEEK LA PG PATENT NO KIND DATE -----

WO 2000055325 A2 20000921 (200055)* EN

RW: AT BE CH CY DE DK EA ES FI FR GB GH GM GR IE IT KE LS LU MC MW NL OA PT SD SE SL SZ TZ UG ZW

W: AE AL AM AT AU AZ BA BB BG BR BY CA CH CN CR CU CZ DE DK DM EE ES
FI GB GD GE GH GM HR HU ID IL IN IS JP KE KG KP KR KZ LC LK LR LS
LT LU LV MA MD MG MK MN MW MX NO NZ PL PT RO RU SD SE SG SI SK SL
TJ TM TR TT TZ UA UG US UZ VN YU ZA ZW

AU 2000037649 A 20001004 (200101)

BR 2000009119 A 20011226 (200206)

EP 1165792 A2 20020102 (200209) EN

R: AT BE CH CY DE DK ES FI FR GB GR IE IT LI LU MC NL US 2003124561 A1 20030703 (200345)

APPLICATION DETAILS:

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FILING DETAILS:

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PRIORITY APPLN. INFO: US 1999-172493P 19991216; US 1999-125219P

19990318; US 1999-127409P 19990401; US 1999-134770P 19990518; US 1999-153584P 19990913; US 1999-154603P 19990917; US 1997-48451P 19970603; US 1998-73741P 19980205; US 1998-90051 19980603; US 2000-531120 20000317; US 2000-553231 20000419; US 2002-170912 20020612

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Search History

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L6: Entry 1 of 1

File: USPT

Aug 4, 1998

DOCUMENT-IDENTIFIER: US 5789210 A

** See image for Certificate of Correction **

TITLE: Recombinant yeasts for effective fermentation of glucose and xylose

Drawing Description Text (4):

FIG. 3 shows the genes cloned on and the restriction map of the plasmid pLSK15.

Drawing Description Text (5):

FIG. 4 shows the genes cloned on and the restriction map of the plasmid pUCKm10.

Drawing Description Text (6):

FIG. 5 shows the genes cloned on and the restriction map of the plasmid pLNH21.

Drawing Description Text (10):

FIG. 7 shows the genes cloned on and the restriction map of plasmid pLNH33.

Drawing Description Text (13):

FIG. 9 is a schematic diagram outlining the construction of pBluescript II KS(-) containing the cloned XR, XD, and XK genes: four such <u>plasmids</u> were constructed: pKS(-)-KK-A*R-KD-1; pKS(-)-KK-A*R-KD-2; pKS(-)-KK-AR-KD-3; and pKS(-)-KK-AR-KD-4, as further described in Example 4.

Drawing Description Text (16):

FIG. 12 is a schematic diagram outlining the construction of the plasmid pLNH21.

Detailed Description Text (5):

The particular XR gene used in the applicants' studies herein was cloned from P. stipitis by Polymerase Chain Reaction (PCR) (Chen and Ho, 1993). The oligonucleotides required for the amplification of XR from the chromosomal DNA by PCR were synthesized according to the published sequence of the P. stipitis XR gene (Takama et al., 1991). The amplified XR was first cloned and stored into plasmid pUC19. The cloned XR was then fused to different promotors including the promotors of yeast TRP5 gene (Zalkin and Yanofsky, 1982) and yeast alcohol dehydrogenase I gene (ADC1) (Ammerer, 1983; Bennetzen and Hall, 1982).

Detailed Description Text (10):

The fusion of XR, XD, and XK to intact promoters from ADC1, PYK, GPD, etc., was carried out by cloning both the fragment containing the specific promoter and the structural gene of XR, XD, or XK on one of the Bluescript KS plasmids (Stratagene, La Jolla, Calif.), followed by the removal of the extra unwanted nucleotides by site-specific mutagenesis (Kunkel et al., 1987). The invention thus also provides several pBluescript II KS(-) (hereinafter pKS(-)) derivatives containing the cloned XD (fused to the pyruvate dehydrogenase promoter), XR (fused to the ADC1 promoter), and XK (fused to the pyruvate kinase promoter). These recombinant plasmids are designated as pKS(-) KD-AR (or A*R) -KK. Four such plasmids were constructed as outlined in FIG. 9. These plasmids have similar but not identical structures. The XR, XD, and XK (or KD-AR (or A*R) -KK) cloned on these plasmids can be separated from the parent pKS(-) plasmid by a single XhoI restriction digestion.

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Detailed Description Text (11):

The XR, XD, and XK genes fused to the proper promoters were then cloned on pLSK15 (FIG. 3) or pUCKm10 (FIG. 4). pLSK15, a derivative of pLX10-14 (Stevis and Ho, 1985), is a low copy number plasmid with a copy number of approximately 10 in yeast (S. cerevisiae). It contains the yeast 2.mu. replicon which enables the plasmid to be replicated autonomously in S. cerevisiae and closely related species. pLSK15 also contains the geneticin (kanamycin) resistance gene (Km.sup.R) and ampicillin resistance gene (Ap.sup.R and also amp.sup.r) which serve as selection markers in S. cerevisiae and other yeasts. pLSK15 also contains the XK gene fused to the yeast TRP-5 promoter. Thus, XR and XD genes fused to proper 5' noncoding sequences containing suitable promoters were inserted into pLSK15 to demonstrate the effect of the resulting plasmids on yeast xylose fermentation. To compare the effect of the presence of different genes on yeast xylose fermentation, a plasmid containing only XR and XD was also used to transform S. cerevisiae and the resulting yeast used in comparative fermentations. Results of the fermentation of xylose by unengineered S. cerevisiae, yeast containing the cloned XR, XD, and XK (SC(pLNH21)), and yeast containing the cloned XR and XD but not XK (SC(pLNH13-32)) genes are shown in FIG. 6A, 6B, and 6C.

Detailed Description Text (12):

pUCKm10 (FIG. 4) is a high copy-number plasmid (i.e. plasmid with a copy number of about 50 or more) with a copy number close to 100 in S. cerevisiae. pUCKm10 is a pUC9 derivative containing the identical 2.mu. replicon, and the Km.sup.R, and Ap.sup.R genes present in pLSK15. These specific DNA fragments serve as the replicon and selection markers that enable the plasmid to be replicated autonomously in S. cerevisiae (and in related yeasts) and also enable the yeast transformants containing the plasmid to be distinguished from the untransformed host cells.

Detailed Description Text (13):

The applicants have constructed pUCKm10 based recombinant plasmids that contain the same XR, XD, and XK fused to 5' proper noncoding sequences containing suitable promotors. These vectors are designed to be useful to transform all S. cerevisiae strains and strains of related species. No special mutants are required to act as the recipient strains. Thus plasmids such as pLNH33 (FIG. 7), as well as pLNH21 (FIG. 5), can be used to transform industrial S. cerevisiae and other strains.

Detailed Description Text (14):

Yeast transformation with derivatives of either pLSK15 or pUCKm10 was carried out by electroporation generally using the the procedure described by Becker and Guarente (1991). Authentic yeast transformants containing derivatives of either pLSK15 or pUCKm10 were isolated as further described below. Km.sup.R present in the plasmids served as the primary selection marker which renders any host cells obtaining one of these plasmids resistant to a much higher concentration of geneticin present in the medium. However, some yeast cells can be induced to become resistant to the same level of geneticin of the transformants containing the plasmid. Thus, not every geneticin resistant colony is a true transformant. It has been reported that Ap.sup.R can be expressed in S. cerevisiae but the latter is resistant to ampicillin without the presence of Ap.sup.R. Thus, Ap.sup.R cannot serve as a selection marker for yeast plasmid-mediated transformation. Nevertheless, yeasts that contain the highly expressed Ap.sup.R will produce sufficient penicillinase and make it possible to identify colonies containing such yeasts on special solid plates by the penicillinase test (Chevallier and Aigle, 1979). The latter test has provided a technique to identify the true transformants of S. cerevisiae and other yeasts from the geneticin resistant colonies.

Detailed Description Text (18):

pLNH33 is a more effective plasmid than pLNH21 for xylose fermentation because it is a higher copy-number plasmid. Furthermore, the XK in pLNH33 is fused to a more efficient promoter than the XK in pLNH21. S. cerevisiae has also been transformed with pLNH33, designated SC(pLNH33). Although SC(pLNH33) is much more effective in fermenting xylose or mixtures of xylose and glucose than SC(pLNH21), 1400(pLNH33) was found to be more effective in fermenting mixtures of glucose and xylose than SC (pLNH33). Thus, individual strains also affect the efficiency of fermentation. Similar to S. cerevisiae, the unengineered strain 1400 cannot use or ferment xylose (alone or in a mixture of glucose and xylose) as shown in FIG. 8B.

Detailed Description Text (30):

Oligonucleotides I and II were used to synthesize the intact XD gene and oligonucleotides II and III were used to synthesize the promotorless XD as shown in FIG. 10. The intact XD and the promotorless XD were first cloned in pKS(-) plasmid. The intact XR was then subcloned on pUCKm10 (FIG. 4) and the resulting plasmid pUCKm10-XD, was used to transform S. cerevisiae by electroporation as described in Example 5. The yeast transformants were used to assay the xylitol dehydrogenase activity to demonstrate that the cloned gene is intact and can be expressed in S. cerevisiae.

Detailed Description Text (34):

The promoter fragment of yeast pyruvate kinase from -910 to +23 (Burke et al., 1983) was synthesized by PCR as described in Example 1 for the synthesis of the XD gene. Both the P.sub.PK fragment and the promotorless XD were subcloned on pKS(-) plasmid and the undesired nucleotides between the P.sub.PK and the intact XD structural gene were removed by site-specific mutagenesis according to the procedure of Kunkel (Kunkel, 1987). The resulting fused gene contains -910 to -1 promoter fragments from the pyruvate kinase gene and +1 to +1963 nucleotides from the Pichia XD gene. The resulting pKS(-) plasmid containing P.sub.PK -XD (or KD) is designated pKS(-)-KD or pKD2.

Detailed Description Text (40):

Plasmid pMA56 (Ammerer, 1983) contains the yeast alcohol dehydrogenase I promoter (P.sub.ADCl). The applicants have used this promoter to modify some of the genes in their work. For example, P.sub.ADCl has been fused to XR, and the resulting gene has been designated P.sub.ADCl -XR or AR. Nevertheless, this P.sub.ADCl is not intact and does not contain the -1 to -14 nucleotides of the intact ADCl promoter (Bennetzen and Hall, 1982). The -1 to -14 region of a gene is usually very significant for controlling protein synthesis. Any gene fused to such a promoter has to rely on its original genetic signal for controlling the synthesis of its protein product.

Detailed Description Text (43):

Construction of plasmid pLNH21 (also designated as pLSK15-KD-AR) and transformation of S. cerevisiae and 1400 with pLNH21

Detailed Description Text (44):

The construction of pLNH21 is outlined in FIG. 12. pLNH21 was used to transform S. cerevisiae and strain 1400 by electroporation under the following conditions. Fifty ml yeast cells, grown to early log phase (Klett Unit (KU) 130), were centrifuged to remove the medium, washed twice with cold water, once with cold 1M sorbitol, and resuspended in 200 .mu.l 1M sorbitol. Sixty .mu.l of the cells were transferred into a 4 ml presterilized plastic tube (with cap) and to which 0.1 .mu.g to 1 .mu.g plasmid DNA was added. Fifty .mu.l of the resulting cells and plasmid mixture were pipetted into a precooled gene pulser cuvette with a 0.2 cm electrode gap and the content in the cuvette was subjected to pulse by the gene pulser with a pulse controller (BioRad) at 2.0 KV, 25 .mu.F, 200 ohms.

Detailed Description Text (46):

Transformation of strain 1400 with pLNH21 or other <u>plasmids</u> was carried out using a similar procedure to that described above, except that the cells were grown to 140-190 KU rather than 130 KU and the YEPD plates for the initial selection of

transformants after electroporation contained 40 .mu.g/ml geneticin G418 rather than 50. Transformation of strain 1400 by the above described procedures was not as effective as transformation of S. cerevisiae.

Detailed Description Text (49):

These three yeasts were cultured in rich medium YEPD aerobically under identical conditions (SC(pLNH13-32) was constructed by transforming S. cerevisiae with a plasmid, designated pLNH13-32, which contains only the XR and XD gene/promotor combinations). These yeast cells were then used to ferment 5% xylose in YEP (1% yeast extract, 2% peptone) medium anaerobically also under identical conditions. The consumption of xylose and the formation of ethanol and xylitol were followed during fermentation by taking samples at proper intervals and analyzing them by HPLC under the following conditions.

Detailed Description Text (61):

When the cell density reached mid-log phase (400 Klett units), 12.5 ml (40%) glucose and 6.25 ml (40%) xylose were added to each flask. After thorough mixing, 1 ml of the culture mixture was removed from the flask to serve as the zero sample. The flask was then sealed with Saran wrap to allow fermentation to be carried out anaerobically. One ml samples of the fermentation broth (with some cells) were removed at proper intervals (every 24 hr.) to serve as samples for measuring the sugar and ethanol contents of the broth during fermentation. The ethanol, glucose, xylose, and xylitol contents of the samples were analyzed by HPLC as described in Example 6. The results, shown in FIGS. 8A and 8B, demonstrate that the genetically engineered yeast 1400(pLNH33) can ferment 10% glucose and 5% xylose to ethanol simultaneously in two to four days without requiring high cell density. On the other hand, the parent strain 1400 can only convert glucose to ethanol but not xylose. The fermentation was carried out under normal conditions, without requiring special medium, special pH, and also without requiring growth of yeast to high cell density. Thus the genetically engineered 1400(pLNH33) containing the XR, XD, and XK, all fused to glycolytic promotors and cloned on a high copy-number plasmid pUCKm10, can ferment high concentrations of both glucose and xylose simultaneously to ethanol in two to four days with very little xylitol produced as a by-product.

Detailed Description Text (78): Chevallier, M. R. and M. Aigle, "Qualitative detection of penicillinase produced by yeast strains carrying chimeric yeast-coli plasmids," FEBS Letters, 108(1) 179-184 (1979).

Other Reference Publication (6):

Chevallier, M. R. and M. Aigle, "Qualitative detection of penicillinase produced by yeast strains carrying chimeric yeast-coli plasmids," FEBS Letters, 108(1) 179-184 (1979).

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REFERENCE
               127:131709
            4:
REFERENCE
            5:
                124:252209
REFERENCE
            6: 111:34409
L51 ANSWER 9 OF 14 REGISTRY COPYRIGHT 2004 ACS on STN
RN
     104118-53-8 REGISTRY
     Reductase, D-xylose (reduced nicotinamide adenine dinucleotide phosphate)
CN
            (CA INDEX NAME)
     (9CI)
OTHER NAMES:
     D-Xylose reductase
CN
     D-Xylose reductase (NADPH)
CN
     NADPH-dependent xylose reductase
CN
     Reductase, xylose (reduced nicotinamide adenine dinucleotide phosphate)
CN
     Xylose reductase
CN
     163913-54-0
DR
     Unspecified
MF
CI
     MAN
SR
                  AGRICOLA, BIOBUSINESS, BIOSIS, CA, CAPLUS, CIN, PIRA,
LC
     STN Files:
       TOXCENTER
*** STRUCTURE DIAGRAM IS NOT AVAILABLE ***
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29 REFERENCES IN FILE CA (1907 TO DATE)

29 REFERENCES IN FILE CAPLUS (1907 TO DATE)

REFERENCE 1: 140:107416

REFERENCE 2: 140:14605

REFERENCE 3: 138:400479

REFERENCE 4: 138:86217

REFERENCE 5: 137:335006

REFERENCE 6: 135:223422

REFERENCE 7: 135:136473

REFERENCE 8: 133:360695

REFERENCE 9: 132:11668

REFERENCE 10: 131:227744

L51 ANSWER 10 OF 14 REGISTRY COPYRIGHT 2004 ACS on STN

RN 99775-25-4 REGISTRY

CN Reductase, D-xylose (9CI) (CA INDEX NAME)

OTHER NAMES:

CN D-Xylose reductase

CN NADH-dependent xylose reductase

CN Xylose reductase

MF Unspecified

CI MAN

SR CA

LC STN Files: AGRICOLA, BIOBUSINESS, BIOSIS, CA, CAPLUS, CIN, PIRA, TOXCENTER, USPATFULL

*** STRUCTURE DIAGRAM IS NOT AVAILABLE ***

- 51 REFERENCES IN FILE CA (1907 TO DATE)
- 1 REFERENCES TO NON-SPECIFIC DERIVATIVES IN FILE CA
- 51 REFERENCES IN FILE CAPLUS (1907 TO DATE)

REFERENCE 1: 139:225259

REFERENCE 2: 139:178733

REFERENCE 3: 139:65901

REFERENCE 4: 138:283215

REFERENCE 5: 138:166431

REFERENCE 6: 138:54597

REFERENCE 7: 138:54591

REFERENCE 8: 137:139426

REFERENCE 9: 137:30383

REFERENCE 10: 136:147534

L51 ANSWER 11 OF 14 REGISTRY COPYRIGHT 2004 ACS on STN

RN 95829-40-6 REGISTRY

CN Reductase, D-xylose (reduced nicotinamide adenine dinucleotide

(phosphate)) (9CI) (CA INDEX NAME) OTHER NAMES: D-Xylose reductase CN NAD(P)H-dependent aldose reductase CNNAD(P)H-dependent xylose reductase CNNADPH-D-xylose reductase CN CN Xylose reductase Unspecified MF CI MAN AGRICOLA, BIOBUSINESS, BIOSIS, CA, CAPLUS, CIN, PIRA, PROMT, STN Files: LC TOXCENTER, USPATFULL *** STRUCTURE DIAGRAM IS NOT AVAILABLE *** 79 REFERENCES IN FILE CA (1907 TO DATE) 2 REFERENCES TO NON-SPECIFIC DERIVATIVES IN FILE CA 79 REFERENCES IN FILE CAPLUS (1907 TO DATE) 1: 140:107416 REFERENCE 140:55544 REFERENCE 140:14605 REFERENCE 3: 139:394971 4: REFERENCE 139:394931 REFERENCE 5: 139:360858 REFERENCE 6: 139:260078 REFERENCE 7: 139:260077 REFERENCE 8: REFERENCE 9: 139:226255 REFERENCE 10: 139:176481 L51 ANSWER 12 OF 14 REGISTRY COPYRIGHT 2004 ACS on STN 9030-58-4 REGISTRY RNKinase (phosphorylating), xylulo- (9CI) (CA INDEX NAME) CNOTHER NAMES: **D-Xylulokinase** CND-Xylulose kinase CN E.C. 2.7.1.17 CNXylulokinase CNXylulose kinase CN57127-28-3 DR Unspecified MF MAN AGRICOLA, BIOBUSINESS, BIOSIS, BIOTECHNO, CA, CAPLUS, CIN, CI STN Files: LC EMBASE, PIRA, TOXCENTER, USPATFULL *** STRUCTURE DIAGRAM IS NOT AVAILABLE *** 154 REFERENCES IN FILE CA (1907 TO DATE) 1 REFERENCES TO NON-SPECIFIC DERIVATIVES IN FILE CA 154 REFERENCES IN FILE CAPLUS (1907 TO DATE) 1: 140:144753 REFERENCE

REFERENCE 2: 140:127260 REFERENCE 3: 140:107886

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139:347843
REFERENCE
           4:
               139:260078
REFERENCE
           5:
           6: 139:260077
REFERENCE
REFERENCE
           7: 139:226997
REFERENCE
           8:
              139:225259
          9: 139:178733
REFERENCE
REFERENCE 10: 139:163668
L51 ANSWER 13 OF 14 REGISTRY COPYRIGHT 2004 ACS on STN
     9028-31-3 REGISTRY
RN
     Reductase, aldose (9CI) (CA INDEX NAME)
CN
OTHER NAMES:
    Aldose reductase
CN
     D-Ribose reductase
CN
CN E.C. 1.1.1.21
    L-Arabinose reductase
CN
    NADPH-aldopentose reductase
CN
     NADPH-aldose reductase
CN
     NADPH-dependent aldose reductase
CN
     NADPH-L-arabinose reductase
CN
     Xylose reductase
CN
     Unspecified
MF
CI
     MAN
     STN Files: ADISNEWS, AGRICOLA, ANABSTR, BIOBUSINESS, BIOSIS, BIOTECHNO,
LC
       CA, CAPLUS, CASREACT, CEN, CIN, EMBASE, IFICDB, IFIPAT, IFIUDB,
       NAPRALERT, PIRA, PROMT, TOXCENTER, USPAT2, USPATFULL
*** STRUCTURE DIAGRAM IS NOT AVAILABLE ***
            2523 REFERENCES IN FILE CA (1907 TO DATE)
              20 REFERENCES TO NON-SPECIFIC DERIVATIVES IN FILE CA
            2531 REFERENCES IN FILE CAPLUS (1907 TO DATE)
            1: 140:146002
REFERENCE
            2: 140:127260
REFERENCE
            3: 140:111411
REFERENCE
            4: 140:108742
REFERENCE
            5: 140:108579
REFERENCE
            6: 140:106718
REFERENCE
            7: 140:104477
REFERENCE
            8: 140:104459
REFERENCE
            9: 140:104246
REFERENCE
REFERENCE 10: 140:94037
L51 ANSWER 14 OF 14 REGISTRY COPYRIGHT 2004 ACS on STN
     9028-16-4 REGISTRY
RN
     Reductase, D-xylulose (9CI) (CA INDEX NAME)
CN
```

OTHER NAMES:

2,3-cis-Polyol dehydrogenase

```
CN D-Xylulose reductase
```

CN Dehydrogenase, 2,3-cis-polyol

CN E.C. 1.1.1.9

CN Erythritol dehydrogenase

CN NAD-dependent meso-erythritol dehydrogenase

CN NAD-dependent xylitol dehydrogenase

CN Polyol dehydrogenase

CN Xylitol dehydrogenase

DR 9032-74-0

MF Unspecified

CI MAN

LC STN Files: AGRICOLA, BIOBUSINESS, BIOSIS, BIOTECHNO, CA, CAPLUS, CASREACT, CIN, EMBASE, PIRA, TOXCENTER, USPATFULL

*** STRUCTURE DIAGRAM IS NOT AVAILABLE ***

213 REFERENCES IN FILE CA (1907 TO DATE)

1 REFERENCES TO NON-SPECIFIC DERIVATIVES IN FILE CA

214 REFERENCES IN FILE CAPLUS (1907 TO DATE)

REFERENCE 1: 140:127260

REFERENCE 2: 140:24162

REFERENCE 3: 140:14605

REFERENCE 4: 139:260078

REFERENCE 5: 139:260077

REFERENCE 6: 139:226997

REFERENCE 7: 139:225259

REFERENCE 8: 139:178733

REFERENCE 9: 139:176481

REFERENCE 10: 139:163668

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FILE COVERS 1907 - 4 Mar 2004 VOL 140 ISS 10 FILE LAST UPDATED: 3 Mar 2004 (20040303/ED)

This file contains CAS Registry Numbers for easy and accurate substance identification.

=> d all tot 149 L49 ANSWER 1 OF 33 HCAPLUS COPYRIGHT 2004 ACS on STN 2003:486085 HCAPLUS ANDN 139:225259 Entered STN: 26 Jun 2003 EDDNA microarray analysis of the expression of the genes TI encoding the major enzymes in ethanol production during glucose and xylose co-fermentation by metabolically engineered Saccharomyces yeast Sedlak, Miroslav; Edenberg, Howard J.; Ho, Nancy W. Y. ΑU School of Engineering, Laboratory of Renewable Resources Engineering, CS Purdue University, West Lafayette, IN, 47907-2022, USA Enzyme and Microbial Technology (2003), 33(1), 19-28 SO CODEN: EMTED2; ISSN: 0141-0229 Elsevier Science PBDTJournal LA English 3-3 (Biochemical Genetics) CC Section cross-reference(s): 10, 16 Lignocellulosic biomass, which contains large amts. of glucose and xylose, AB is the new ideal feedstock for ethanol production used as renewable liquid fuel for transportation. The naturally occurring Saccharomyces yeasts traditionally used for industrial ethanol production are unable to ferment xylose. The authors have successfully developed genetically engineered Saccharomyces yeasts that can effectively co-ferment both glucose and xylose simultaneously to ethanol. The engineered yeast contains three xylose metabolizing genes, the xylose reductase (XR), xylitol dehydrogenase (XD) and xylulokinase (XK) genes , fused to glycolytic promoters, on high copy plasmids or integrated into the yeast chromosome in multiple copies. Although the glucose/xylose co-fermenting yeasts are currently the most effective yeast for producing ethanol from cellulosic biomass, they still utilize glucose more efficiently than xylose. The authors believe that carefully analyzing gene expression during co-fermentation of glucose and xylose to ethanol, using the genetically modified strains, will reveal ways to optimize xylose fermentation In this paper, the authors report the results on analyzing the expression of genes in the glycolytic and alc. fermentation pathways using microarray technol. The authors also report the results on the anal. of the activities of the selected enzymes for ethanol production during co-fermentation of glucose and xylose to ethanol by one of the effective glucose/xylose co-fermenting yeasts 424A(LNH-ST). DNA microarray ethanol glucose xylose fermn Saccharomyces ST Saccharomyces cerevisiae IT (424A(LNH-ST); DNA microarray anal. of genes encoding enzymes involved in glycolysis and alc. fermentation glucose and xylose co-fermentation by metabolically engineered Saccharomyces yeast 424A(LNH-ST)) IT Gene, microbial RL: BSU (Biological study, unclassified); BIOL (Biological study) (ADH1; DNA microarray anal. of genes encoding enzymes involved in glycolysis and alc. fermentation glucose and xylose co-fermentation by metabolically engineered Saccharomyces yeast 424A (LNH-ST)) Gene, microbial ΙT RL: BSU (Biological study, unclassified); BIOL (Biological study)

(ADH2; DNA microarray anal. of genes encoding

424A(LNH-ST))

enzymes involved in glycolysis and alc. fermentation glucose and xylose

co-fermentation by metabolically engineered Saccharomyces yeast

```
Gene, microbial
IT
     RL: BSU (Biological study, unclassified); BIOL (Biological study)
        (ADH3; DNA microarray anal. of genes encoding
        enzymes involved in glycolysis and alc. fermentation glucose and xylose
        co-fermentation by metabolically engineered Saccharomyces yeast
        424A(LNH-ST))
     Gene, microbial
IT
     RL: BSU (Biological study, unclassified); BIOL (Biological study)
        (ADH4; DNA microarray anal. of genes encoding
        enzymes involved in glycolysis and alc. fermentation glucose and xylose
        co-fermentation by metabolically engineered Saccharomyces yeast
        424A (LNH-ST))
     DNA microarray technology
IT
        (Affymetrix; DNA microarray anal. of genes encoding
        enzymes involved in glycolysis and alc. fermentation glucose and xylose
        co-fermentation by metabolically engineered Saccharomyces yeast
        424A(LNH-ST))
     Fermentation
IT
       Gene expression profiles, microbial
     Genetic engineering
     Glycolysis
        (DNA microarray anal. of genes encoding enzymes
        involved in glycolysis and alc. fermentation glucose and xylose
co-fermentation by
        metabolically engineered Saccharomyces yeast 424A(LNH-ST))
IT
     Gene, microbial
     RL: BSU (Biological study, unclassified); BIOL (Biological study)
        (ENO1; DNA microarray anal. of genes encoding
        enzymes involved in glycolysis and alc. fermentation glucose and xylose
        co-fermentation by metabolically engineered Saccharomyces yeast
        424A (LNH-ST))
     Gene, microbial
IT
     RL: BSU (Biological study, unclassified); BIOL (Biological study)
        (ENO2; DNA microarray anal. of genes encoding
        enzymes involved in glycolysis and alc. fermentation glucose and xylose
        co-fermentation by metabolically engineered Saccharomyces yeast
        424A(LNH-ST))
     Gene, microbial
IT
     RL: BSU (Biological study, unclassified); BIOL (Biological study)
        (FBI1; DNA microarray anal. of genes encoding
        enzymes involved in glycolysis and alc. fermentation glucose and xylose
        co-fermentation by metabolically engineered Saccharomyces yeast
        424A(LNH-ST))
IT
     Gene, microbial
     RL: BSU (Biological study, unclassified); BIOL (Biological study)
         (GLK1; DNA microarray anal. of genes encoding
        enzymes involved in glycolysis and alc. fermentation glucose and xylose
        co-fermentation by metabolically engineered Saccharomyces yeast
        424A (LNH-ST))
IT
     Gene, microbial
     RL: BSU (Biological study, unclassified); BIOL (Biological study)
         (GPM1; DNA microarray anal. of genes encoding
        enzymes involved in glycolysis and alc. fermentation glucose and xylose
        co-fermentation by metabolically engineered Saccharomyces yeast
        424A(LNH-ST))
IT
     Gene, microbial
     RL: BSU (Biological study, unclassified); BIOL (Biological study)
         (HXK1; DNA microarray anal. of genes encoding
        enzymes involved in glycolysis and alc. fermentation glucose and xylose
        co-fermentation by metabolically engineered Saccharomyces yeast
        424A (LNH-ST))
     Gene, microbial
IT
```

RL: BSU (Biological study, unclassified); BIOL (Biological study)

(HXK2; **DNA** microarray anal. of **genes** encoding enzymes involved in glycolysis and alc. fermentation glucose and xylose co-fermentation by metabolically engineered Saccharomyces **yeast** 424A(LNH-ST))

IT Gene, microbial

RL: BSU (Biological study, unclassified); BIOL (Biological study)
(PDC1; **DNA** microarray anal. of **genes** encoding
enzymes involved in glycolysis and alc. fermentation glucose and xylose
co-fermentation by metabolically engineered Saccharomyces **yeast**424A(LNH-ST))

IT Gene, microbial

RL: BSU (Biological study, unclassified); BIOL (Biological study)
(PDC5; DNA microarray anal. of genes encoding
enzymes involved in glycolysis and alc. fermentation glucose and xylose
co-fermentation by metabolically engineered Saccharomyces yeast
424A(LNH-ST))

IT Gene, microbial

RL: BSU (Biological study, unclassified); BIOL (Biological study)
(PDC6; **DNA** microarray anal. of **genes** encoding
enzymes involved in glycolysis and alc. fermentation glucose and xylose
co-fermentation by metabolically engineered Saccharomyces **yeast**424A(LNH-ST))

IT Gene, microbial

RL: BSU (Biological study, unclassified); BIOL (Biological study)
(PFK1; DNA microarray anal. of genes encoding
enzymes involved in glycolysis and alc. fermentation glucose and xylose
co-fermentation by metabolically engineered Saccharomyces yeast
424A(LNH-ST))

IT Gene, microbial

RL: BSU (Biological study, unclassified); BIOL (Biological study)
(PFK2; DNA microarray anal. of genes encoding
enzymes involved in glycolysis and alc. fermentation glucose and xylose
co-fermentation by metabolically engineered Saccharomyces yeast
424A(LNH-ST))

IT Gene, microbial

RL: BSU (Biological study, unclassified); BIOL (Biological study)
(PGI1; **DNA** microarray anal. of **genes** encoding
enzymes involved in glycolysis and alc. fermentation glucose and xylose
co-fermentation by metabolically engineered Saccharomyces **yeast**424A(LNH-ST))

IT Gene, microbial

RL: BSU (Biological study, unclassified); BIOL (Biological study)
(PGK1; DNA microarray anal. of genes encoding
enzymes involved in glycolysis and alc. fermentation glucose and xylose
co-fermentation by metabolically engineered Saccharomyces yeast
424A(LNH-ST))

IT Gene, microbial

RL: BSU (Biological study, unclassified); BIOL (Biological study)
(PYK1; **DNA** microarray anal. of **genes** encoding
enzymes involved in glycolysis and alc. fermentation glucose and xylose
co-fermentation by metabolically engineered Saccharomyces **yeast**424A(LNH-ST))

IT Gene, microbial

RL: BSU (Biological study, unclassified); BIOL (Biological study)
(PYK2; **DNA** microarray anal. of **genes** encoding
enzymes involved in glycolysis and alc. fermentation glucose and xylose
co-fermentation by metabolically engineered Saccharomyces **yeast**424A(LNH-ST))

IT Gene, microbial

RL: BSU (Biological study, unclassified); BIOL (Biological study)
(TDH1; DNA microarray anal. of genes encoding
enzymes involved in glycolysis and alc. fermentation glucose and xylose
co-fermentation by metabolically engineered Saccharomyces yeast

```
424A(LNH-ST))
    Gene, microbial
ΙT
    RL: BSU (Biological study, unclassified); BIOL (Biological study)
        (TDH2; DNA microarray anal. of genes encoding
        enzymes involved in glycolysis and alc. fermentation glucose and xylose
        co-fermentation by metabolically engineered Saccharomyces yeast
        424A(LNH-ST))
    Gene, microbial
IT
    RL: BSU (Biological study, unclassified); BIOL (Biological study)
        (TDH3; DNA microarray anal. of genes encoding
        enzymes involved in glycolysis and alc. fermentation glucose and xylose
        co-fermentation by metabolically engineered Saccharomyces yeast
        424A(LNH-ST))
     Gene, microbial
IT
     RL: BSU (Biological study, unclassified); BIOL (Biological study)
        (TPI1; DNA microarray anal. of genes encoding
        enzymes involved in glycolysis and alc. fermentation glucose and xylose
        co-fermentation by metabolically engineered Saccharomyces yeast
        424A (LNH-ST))
     Enzymes, biological studies
IT
     RL: BSU (Biological study, unclassified); BIOL (Biological study)
        (glycolytic; DNA microarray anal. of genes encoding
        enzymes involved in glycolysis and alc. fermentation glucose and xylose
        co-fermentation by metabolically engineered Saccharomyces yeast
        424A(LNH-ST))
                                                        9001-04-1, Pyruvate
     64-17-5, Ethanol, biological studies
                                            591-59-3
IT
                                                  9001-83-6, Phosphoglycerate
                    9001-59-6, Pyruvate kinase
     decarboxylase
              9031-72-5, Alcohol dehydrogenase
     RL: BSU (Biological study, unclassified); BIOL (Biological study)
        (DNA microarray anal. of genes encoding enzymes
        involved in glycolysis and alc. fermentation glucose and xylose
co-fermentation by
        metabolically engineered Saccharomyces yeast 424A(LNH-ST))
                                             58-86-6, Xylose, biological
     50-99-7, D-Glucose, biological studies
IT
     studies
     RL: BUU (Biological use, unclassified); BIOL (Biological study); USES
     (Uses)
        (DNA microarray anal. of genes encoding enzymes
        involved in glycolysis and alc. fermentation glucose and xylose
co-fermentation by
        metabolically engineered Saccharomyces yeast 424A(LNH-ST))
     9028-16-4, Xylitol dehydrogenase
IT
     9030-58-4, Xylulokinase 99775-25-4,
     Xylose reductase
     RL: BUU (Biological use, unclassified); BIOL (Biological study); USES
         (DNA microarray anal. of genes encoding enzymes
        involved in glycolysis and alc. fermentation glucose and xylose
co-fermentation by
        metabolically engineered Saccharomyces yeast 424A(LNH-ST)
        containing)
              THERE ARE 11 CITED REFERENCES AVAILABLE FOR THIS RECORD
RE.CNT 11
RE
(1) Bradford, M; Anal Chem 1976, V72, P248 HCAPLUS
(2) Ho, N; WO 97/42307 HCAPLUS
(3) Ho, N; US 08/148581 1998
(4) Ho, N; US 5789210 1998 HCAPLUS
(5) Ho, N; ACS Symposium Series 2000, V767, P142
(6) Ho, N; Advances in biochemical engineering/biotechnology 1999
(7) Ho, N; Appl Environ Microbiol 1998, V64, P1852 MEDLINE
(8) Kotter, P; Appl Microbiol Biotechnol 1993, V38, P776
(9) Li, R; A laboratory guide to RNA: isolation analysis, and synthesis 1996,
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P43

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(10) Maitra, P; J Biol Chem 1971, V246, P475 HCAPLUS
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(11) Walfridsson, M; Appl Microbiol Biotechnol 1997, V48, P218 HCAPLUS

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L49 ANSWER 2 OF 33 HCAPLUS COPYRIGHT 2004 ACS on STN
```

AN 2003:282043 HCAPLUS

DN 138:302748

ED Entered STN: 11 Apr 2003

TI Manufacture of five-carbon sugars and sugar alcohols

IN Miasnikov, Andrei; Ojamo, Heikki; Povelainen, Mira; Gros, Hakan; Toivari, Mervi; Richard, Peter; Ruohonen, Laura; Koivuranta, Kari; Londesborough, John; Aristidou, Aristos; Penttila, Merja; Plazanet-Menut, Claire; Deutscher, Josef

PA Xyrofin Oy, Finland

SO U.S. Pat. Appl. Publ., 96 pp., Cont.-in-part of U.S. Ser. No. 488,581, abandoned.

CODEN: USXXCO

DT Patent

LA English

IC ICM C12P007-18

ICS C12N001-21; C12N001-18

NCL 435158000; 435252300; 435254200

CC 16-2 (Fermentation and Bioindustrial Chemistry)

Section cross-reference(s): 3

FAN.CNT 3 PATENT NO.				KIND		DATE		APPLICATION NO.					o. :	DATE					
	ΡI	US 2003068791				 Δ1		20030410					01-9			20010	720	<	
	PI	HU 72187						19960328								19931105		<	
		HU 219016						20010129											
AT 184917					19991015			AT 1993-924615					1993	1105	<				
		ES 2139024						20000201									19931105		
					A										1995	0103	<		
		WO 2001053306								00 2333 00,000					20010122				
	WO 2001053306 WO 2001053306							,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,											
		WO								ΔII.	Δ7.	BA.	BB.	BG.	BR.	BY,	BZ.	CA,	CH,
			VV -	CN	CD	CII	C7.	CZ,	DE,	DE,	DK.	DK.	DM.	DZ.	EE.	EE,	ES.	FI.	FI.
				CN,	CR,	CU,	CH,	CM,	HD,	HII	TD.	TI.	TN.	TS.	JP.	KE,	KG.	KP.	KR.
				GB,	UD,	LK	T.D	T.C	T.T	T.II	LV	MA.	MD.	MG.	MK.	MN,	MW.	MX.	MZ.
																SL,			
				TYCT,	T7	IIA	IIC	IIC,	IIZ	VNI	VII	ZA	ZW.	AM.	A7.	BY,	KG.	KZ.	MD.
					TJ,			05,	02,	V1.,	10,	,	2,	,	,	,	,	,	,
	•		DW.					MW	M 7.	GD.	ST.	S7.	Т7.	IIG.	ZW.	AT,	BE.	CH.	CY.
			RW:	On,	עם,	EC,	ът,	ED.	CB	GP,	TE.	TT	T.II	MC.	NT.	PT,	SE.	TR.	BF.
				DE,	OE,	cc,	CT,	CM	GD,	GN,	GM,	MT.	MD,	NE.	SN	TD,	TG	,	,
	DD3.		1000									νш,	riic,	нь,	0117	12,			
	PRAI	US 1992-973325																	
		US 1993-110672																	
		US 1995-368395																	
		US 1997-790585																	
		US 2000-488581 WO 2001-FI51																	
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AB The invention relates to the methods of manufacturing C5 sugars and sugar alcs. as well as other compds. derived from the pentose phosphate pathway from readily available substrates such as hexoses using metabolically engineered microbial hosts.

pentose pentitol fermn bacteria genetic engineering; gene sequence xylitol arabitol phosphate dehydrogenase

IT Gene, microbial

RL: BSU (Biological study, unclassified); BIOL (Biological study)
(APDH; manufacture of five-carbon sugars and sugar alcs.)

IT Gene, microbial

RL: BSU (Biological study, unclassified); BIOL (Biological study) (DOG1; manufacture of five-carbon sugars and sugar alcs.)

IT Gene, microbial

```
RL: BSU (Biological study, unclassified); BIOL (Biological study)
        (LTP1; manufacture of five-carbon sugars and sugar alcs.)
     Gene, microbial
IT
     RL: BSU (Biological study, unclassified); BIOL (Biological study)
        (XPDH; manufacture of five-carbon sugars and sugar alcs.)
IT
     DNA sequences
     Protein sequences
        (bacterial xylitol and arabitol phospphate dehydrogenases)
IT
     Bacillus subtilis
     Genetic engineering
     Pentose phosphate pathway
       Saccharomyces cerevisiae
        (manufacture of five-carbon sugars and sugar alcs.)
IT
     Alditols
     Pentoses
     RL: BMF (Bioindustrial manufacture); BPN (Biosynthetic preparation); BIOL
     (Biological study); PREP (Preparation)
        (manufacture of five-carbon sugars and sugar alcs.)
IT
     Bacillus halodurans
     Enterococcus avium
     Lactobacillus rhamnosus
        (manufacture of five-carbon sugars and sugar alcs. with enzyme from)
     Gene, microbial
TT
     RL: BSU (Biological study, unclassified); BIOL (Biological study)
        (rpi; manufacture of five-carbon sugars and sugar alcs.)
                   510787-76-5
     510787-74-3
IT
     RL: BSU (Biological study, unclassified); PRP (Properties); BIOL
     (Biological study)
        (amino acid sequence; manufacture of five-carbon sugars and sugar alcs.)
     9035-82-9, Dehydrogenase
IT
     RL: BSU (Biological study, unclassified); BIOL (Biological study)
        (for arabitol phosphate; manufacture of five-carbon sugars and sugar alcs.)
                                              488-82-4P, D-Arabitol
     87-99-0P, Xylitol
                        488-81-3P, Ribitol
IT
                                                       1114-34-7P, D-Lyxose
                              551-84-8P, D-Xylulose
     488-84-6P, D-Ribulose
                             10323-20-3P, D-Arabinose
     6917-36-8P, Pentitol
     RL: BMF (Bioindustrial manufacture); BPN (Biosynthetic preparation); BIOL
     (Biological study); PREP (Preparation) (manufacture of five-carbon sugars and sugar alcs.)
     9028-16-4, Xylitol dehydrogenase
                                         64886-68-6,
IT
     Xylitol phosphate dehydrogenase
     RL: BSU (Biological study, unclassified); BIOL (Biological study)
         (manufacture of five-carbon sugars and sugar alcs.)
                   510787-77-6
ΙT
     510787-75-4
     RL: BSU (Biological study, unclassified); PRP (Properties); BIOL
     (Biological study)
        (nucleotide sequence; manufacture of five-carbon sugars and sugar alcs.)
                                                510945-01-4
                                                               510945-02-5
                   510944-99-7
IT
     510944-98-6
                                  510945-00-3
                                                510945-06-9
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                   510945-46-7
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                                                               510945-54-7
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                   510945-51-4
     510945-55-8
     RL: PRP (Properties)
         (unclaimed nucleotide sequence; manufacture of five-carbon sugars and sugar
        alcs.)
                                                               510945-29-6
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                    510945-24-1
                                  510945-27-4
IT
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                   510945-41-2
     510945~30-9
     RL: PRP (Properties)
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(unclaimed protein sequence; manufacture of five-carbon sugars and sugar
        alcs.)
                                  351900-50-0
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                   351900-49-7
    351900-48-6
ΙT
                                  351900-55-5
                   351900-54-4
     351900-53-3
    RL: PRP (Properties)
        (unclaimed sequence; manufacture of five-carbon sugars and sugar alcs.)
    ANSWER 3 OF 33 HCAPLUS COPYRIGHT 2004 ACS on STN
L49
     2001:545704 HCAPLUS
AN
DN
     135:136473
     Entered STN: 27 Jul 2001
ΕD
     Manufacture of five-carbon sugars and sugar alcohols using microorganisms
TI
     deficient in or transformed with genes involved in
     pentose-phosphate pathway
    Miasnikov, Andrei; Ojamo, Heikki; Povelainen, Mira; Gros, Hakan; Toivari,
IN
     Mervi; Richard, Peter; Ruohonen, Laura; Koivuranta, Kari; Londesborough,
     John; Aristidou, Aristos; Penttilae, Merja; Plazanet-Menut, Claire;
     Deutscher, Josef
     Xyrofin Oy, Finland
PA
     PCT Int. Appl., 205 pp.
SO
     CODEN: PIXXD2
DT
     Patent
LA
     English
IC
     ICM C07H
     16-2 (Fermentation and Bioindustrial Chemistry)
     Section cross-reference(s): 3, 6, 10, 33
FAN.CNT 3
                                            APPLICATION NO. DATE
                      KIND DATE
     PATENT NO.
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                             _____
                     A2
A3
                                            WO 2001-FI51 20010122
                             20010726
PI
     WO 2001053306
                             20020418
     WO 2001053306
         W: AE, AG, AL, AM, AT, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH,
             CN, CR, CU, CZ, CZ, DE, DE, DK, DK, DM, DZ, EE, EE, ES, FI, FI,
             GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR,
             KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SK, SL, TJ, TM, TR,
             TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW, AM, AZ, BY, KG, KZ, MD,
             RU, TJ, TM
         RW: GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZW, AT, BE, CH, CY,
             DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, TR, BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG
                                            AU 2001-31784
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                       A5
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     BR 2001007918
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                                                               20010122
     EP 1254244
                        A2
            AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LI, LU, NL, SE, MC, PT, IE, SI, LT, LV, FI, RO, MK, CY, AL, TR
                                            JP 2001-553780
                                                               20010122
     JP 2003520583
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                             20030708
                                             US 2001-908744
                                                               20010720 <--
     US 2003068791
                             20030410
                        A1
PRAI US 2000-488581
                             20000121
                        Α
     US 1992-973325
                       B2
                             19921105
                                       <--
                       B1
                             19930824
                                       <--
     US 1993-110672
                       A1
     US 1995-368395
                             19950103
                                       <--
     US 1997-790585
                        A2
                             19970129
                        W
                             20010122
     WO 2001-FI51
     The invention relates to the methods of manufacturing five-carbon sugars and
AB
     sugar alcs. as well as other compds. derived from pentose-phosphate
     pathway (PPP) from readily available substrates such a hexoses using
     metabolically engineered microbial hosts. A series of the genes
     involved in the PPP are cloned from various microorganisms or disrupted in
     the host of either Bacillus subtilis or Saccharomyces cerevisiae. This
     strategy is demonstrated to successfully increase the yield of a variety
     of the five-carbon sugar or sugar alcs. for manufacturing purpose.
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five carbon sugar alc fermn Bacillus Saccharomyces PPP gene;

ST

pentose phosphate pathway **gene** mutagenesis overexpression transformation fermn

IT Plasmid vectors

(B1003, gene XYL2 on; manufacture of five-carbon sugars and sugar alcs. using microorganisms deficient in or transformed with genes involved in pentose-phosphate pathway)

IT Plasmid vectors

(B1011, gene IDP2 on; manufacture of five-carbon sugars and sugar alcs. using microorganisms deficient in or transformed with genes involved in pentose-phosphate pathway)

IT **Plasmid** vectors

(B1068, **gene** XYL2 on; manufacture of five-carbon sugars and sugar alcs. using microorganisms deficient in or transformed with **genes** involved in pentose-phosphate pathway)

IT Plasmid vectors

(B11154, gene XKS1 on; manufacture of five-carbon sugars and sugar alcs. using microorganisms deficient in or transformed with genes involved in pentose-phosphate pathway)

IT Plasmid vectors

(B1187, gene PGI1 on; manufacture of five-carbon sugars and sugar alcs. using microorganisms deficient in or transformed with genes involved in pentose-phosphate pathway)

IT **Plasmid** vectors

(B1449, gene LTP1 on; manufacture of five-carbon sugars and sugar alcs. using microorganisms deficient in or transformed with genes involved in pentose-phosphate pathway)

IT Plasmid vectors

(B995, **gene** XYL2 on; manufacture of five-carbon sugars and sugar alcs. using microorganisms deficient in or transformed with **genes** involved in pentose-phosphate pathway)

IT Klebsiella terrigena

(D-xylulose-forming arabitol dehydrogenase **gene**; manufacture of five-carbon sugars and sugar alcs. using microorganisms deficient in or transformed with **genes** involved in pentose-phosphate pathway)

IT Gene, microbial

RL: BUU (Biological use, unclassified); BIOL (Biological study); USES (Uses)

(DOG1, for 2-deoxyglucose-6-phosphate phosphatase; manufacture of five-carbon sugars and sugar alcs. using microorganisms deficient in or transformed with **genes** involved in pentose-phosphate pathway)

IT Gene, microbial

RL: BUU (Biological use, unclassified); BIOL (Biological study); USES (Uses)

(DOG2, for 2-deoxyglucose-6-phosphate phosphatase; manufacture of five-carbon sugars and sugar alcs. using microorganisms deficient in or transformed with **genes** involved in pentose-phosphate pathway)

IT Gene, microbial

RL: BUU (Biological use, unclassified); BIOL (Biological study); USES (Uses)

(GDH2, expression in Pichia stipitis of; manufacture of five-carbon sugars and sugar alcs. using microorganisms deficient in or transformed with genes involved in pentose-phosphate pathway)

IT Gene, microbial

RL: BSU (Biological study, unclassified); BUU (Biological use, unclassified); BIOL (Biological study); USES (Uses)

(LTP, for Low Mol. Weight Protein-Tyrosine Phosphatase; manufacture of five-carbon sugars and sugar alcs. using microorganisms deficient in or transformed with **genes** involved in pentose-phosphate pathway)

IT Gene, microbial

RL: BSU (Biological study, unclassified); BUU (Biological use, unclassified); BIOL (Biological study); USES (Uses)

(LTP1, for Low Mol. Weight Protein-Tyrosine Phosphatase; manufacture of five-carbon sugars and sugar alcs. using microorganisms deficient in or

```
transformed with genes involved in pentose-phosphate pathway)
    Gene, microbial
IT
    RL: BPR (Biological process); BSU (Biological study, unclassified); BUU
     (Biological use, unclassified); BIOL (Biological study); PROC (Process);
    USES (Uses)
        (MAE1, overexpression and carbon source utilization in Saccharomyces
        cerevisiae; manufacture of five-carbon sugars and sugar alcs. using
        microorganisms deficient in or transformed with genes
        involved in pentose-phosphate pathway)
     Peptoniphilus asaccharolyticus
IT
        (NAD-glutamate dehydrogenase of; manufacture of five-carbon sugars and sugar
        alcs. using microorganisms deficient in or transformed with
        genes involved in pentose-phosphate pathway)
     Gene, microbial
IT
     RL: BSU (Biological study, unclassified); BUU (Biological use,
     unclassified); BIOL (Biological study); USES (Uses)
        (PFK26, for 6-phosphofructo-L-kinase; manufacture of five-carbon sugars and
        sugar alcs. using microorganisms deficient in or transformed with
        genes involved in pentose-phosphate pathway)
IT
     Gene, microbial
     RL: BSU (Biological study, unclassified); BUU (Biological use,
     unclassified); BIOL (Biological study); USES (Uses)
        (PFK27, for 6-phosphofructo-L-kinase; manufacture of five-carbon sugars and
        sugar alcs. using microorganisms deficient in or transformed with
        genes involved in pentose-phosphate pathway)
IT
     Gene, microbial
     RL: BSU (Biological study, unclassified); BUU (Biological use,
     unclassified); BIOL (Biological study); USES (Uses)
        (PGI1, for phosphoglucoisomerase; manufacture of five-carbon sugars and
        sugar alcs. using microorganisms deficient in or transformed with
        genes involved in pentose-phosphate pathway)
IT
     Gene, microbial
     RL: BSU (Biological study, unclassified); BUU (Biological use,
     unclassified); PRP (Properties); BIOL (Biological study); USES (Uses)
        (PPPase1; manufacture of five-carbon sugars and sugar alcs. using
        microorganisms deficient in or transformed with genes
        involved in pentose-phosphate pathway)
     Gene, microbial
IT
     RL: BSU (Biological study, unclassified); BUU (Biological use,
     unclassified); PRP (Properties); BIOL (Biological study); USES (Uses)
        (PPPase2; manufacture of five-carbon sugars and sugar alcs. using
        microorganisms deficient in or transformed with genes
        involved in pentose-phosphate pathway)
     Gene, microbial
IT
     RL: BUU (Biological use, unclassified); BIOL (Biological study); USES
        (TKL1; manufacture of five-carbon sugars and sugar alcs. using
        microorganisms deficient in or transformed with genes
        involved in pentose-phosphate pathway)
     Gene, microbial
IT
     RL: BUU (Biological use, unclassified); BIOL (Biological study); USES
        (TKL2; manufacture of five-carbon sugars and sugar alcs. using
        microorganisms deficient in or transformed with genes
        involved in pentose-phosphate pathway)
ΙT
     Morganella morganii
        (XDH (xylitol dehydrogenase) gene cloned
        from; manufacture of five-carbon sugars and sugar alcs. using microorganisms
        deficient in or transformed with genes involved in
        pentose-phosphate pathway)
     Gene, microbial
IT
     RL: BUU (Biological use, unclassified); BIOL (Biological study); USES
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(Uses)

IT

IT

IT

IT

IT

IT

IT

IT

ΙT

IT

IT

Operon

(XK, expression in Pichia stipitis of; manufacture of five-carbon sugars and sugar alcs. using microorganisms deficient in or transformed with genes involved in pentose-phosphate pathway) Gene, microbial RL: BSU (Biological study, unclassified); BUU (Biological use, unclassified); BIOL (Biological study); USES (Uses) (XYL1, for xylose reductase; manufacture of five-carbon sugars and sugar alcs. using microorganisms deficient in or transformed with genes involved in pentose-phosphate pathway) Gene, microbial RL: BSU (Biological study, unclassified); BUU (Biological use, unclassified); BIOL (Biological study); USES (Uses) (XYL2, for xylose dehydrogenase; manufacture of five-carbon sugars and sugar alcs. using microorganisms deficient in or transformed with genes involved in pentose-phosphate pathway) Fermentation (anaerobic; manufacture of five-carbon sugars and sugar alcs. using microorganisms deficient in or transformed with genes involved in pentose-phosphate pathway) Hexoses Pentoses Polysaccharides, biological studies RL: BPR (Biological process); BSU (Biological study, unclassified); BIOL (Biological study); PROC (Process) (as carbon source; manufacture of five-carbon sugars and sugar alcs. using microorganisms deficient in or transformed with genes involved in pentose-phosphate pathway) Redox reaction (biochem., electron transport using; manufacture of five-carbon sugars and sugar alcs. using microorganisms deficient in or transformed with genes involved in pentose-phosphate pathway) Brevibacterium Corynebacterium (fermentation host; manufacture of five-carbon sugars and sugar alcs. using microorganisms deficient in or transformed with genes involved in pentose-phosphate pathway) Pichia Saccharomyces Schizosaccharomyces Schizosaccharomyces pombe Yamadazyma Yamadazyma stipitis (fermentation using; manufacture of five-carbon sugars and sugar alcs. using microorganisms deficient in or transformed with genes involved in pentose-phosphate pathway) Michaelis constant (for nicotinamide coenzymes of malic enzyme of yeast; manufacture of five-carbon sugars and sugar alcs. using microorganisms deficient in or transformed with genes involved in pentose-phosphate pathway) Gene, microbial RL: BSU (Biological study, unclassified); BUU (Biological use, unclassified); BIOL (Biological study); USES (Uses) (fucI, for L-fucose isomerase; manufacture of five-carbon sugars and sugar alcs. using microorganisms deficient in or transformed with genes involved in pentose-phosphate pathway) Ralstonia eutropha (genes for polyhydroxybutyrate biosynthetic enzymes of, expression in yeast of; manufacture of five-carbon sugars and

sugar alcs. using microorganisms deficient in or transformed with

(glcUgdh; manufacture of five-carbon sugars and sugar alcs. using

genes involved in pentose-phosphate pathway)

microorganisms deficient in or transformed with genes involved in pentose-phosphate pathway) Biological transport ΙT (glucose; manufacture of five-carbon sugars and sugar alcs. using microorganisms deficient in or transformed with genes involved in pentose-phosphate pathway) Carboxylic acids, preparation IT RL: BMF (Bioindustrial manufacture); BIOL (Biological study); PREP (Preparation) (hydroxy, polymers, fermentation of; manufacture of five-carbon sugars and sugar alcs. using microorganisms deficient in or transformed with genes involved in pentose-phosphate pathway) Pseudomonas cichorii (ketose 3-epimerase gene of; manufacture of five-carbon sugars and IT sugar alcs. using microorganisms deficient in or transformed with genes involved in pentose-phosphate pathway) Corynebacterium glutamicum IT (lysine fermentation with transgenic; manufacture of five-carbon sugars and sugar alcs. using microorganisms deficient in or transformed with genes involved in pentose-phosphate pathway) Aspergillus nidulans (malic enzyme of, expression in Pichia of gene for; manufacture of IT five-carbon sugars and sugar alcs. using microorganisms deficient in or transformed with genes involved in pentose-phosphate pathway) Bacillus halodurans IT Bacillus subtilis Biomass Clostridium difficile DNA sequences Electron transport system, biological Enterococcus avium Fermentation Lactobacillus rhamnosus Molecular cloning Mutagenesis Pentose phosphate pathway Protein sequences Trichoderma reesei Zygosaccharomyces rouxii (manufacture of five-carbon sugars and sugar alcs. using microorganisms deficient in or transformed with genes involved in pentose-phosphate pathway) Gene, microbial ITRL: BSU (Biological study, unclassified); BUU (Biological use, unclassified); BIOL (Biological study); USES (Uses) (manufacture of five-carbon sugars and sugar alcs. using microorganisms deficient in or transformed with genes involved in pentose-phosphate pathway) Promoter (genetic element) IT RL: BUU (Biological use, unclassified); BIOL (Biological study); USES (Uses) (manufacture of five-carbon sugars and sugar alcs. using microorganisms deficient in or transformed with genes involved in pentose-phosphate pathway) Coenzymes IT RL: BPR (Biological process); BSU (Biological study, unclassified); BIOL (Biological study); PROC (Process) (nicotinamide; manufacture of five-carbon sugars and sugar alcs. using microorganisms deficient in or transformed with genes

involved in pentose-phosphate pathway)

Enzyme kinetics

IT

(of malic enzyme of Saccharomyces cerevisiae; manufacture of five-carbon sugars and sugar alcs. using microorganisms deficient in or transformed with **genes** involved in pentose-phosphate pathway)

IT **Plasmid** vectors

(p131, rpi gene on; manufacture of five-carbon sugars and sugar alcs. using microorganisms deficient in or transformed with genes involved in pentose-phosphate pathway)

IT Plasmid vectors

(p131:Cm-2, rpi gene on; manufacture of five-carbon sugars and sugar alcs. using microorganisms deficient in or transformed with genes involved in pentose-phosphate pathway)

IT Plasmid vectors

(pAOS63, gene XYL2 on; manufacture of five-carbon sugars and sugar alcs. using microorganisms deficient in or transformed with genes involved in pentose-phosphate pathway)

IT Plasmid vectors

(pAOS64, gene XYL2 on; manufacture of five-carbon sugars and sugar alcs. using microorganisms deficient in or transformed with genes involved in pentose-phosphate pathway)

IT Plasmid vectors

(pAOS66, XYL1 and XYL2 genes on, expression in Pichia stipitis of; manufacture of five-carbon sugars and sugar alcs. using microorganisms deficient in or transformed with genes involved in pentose-phosphate pathway)

IT Plasmid vectors

(pAOS67, gene XYL2 on; manufacture of five-carbon sugars and sugar alcs. using microorganisms deficient in or transformed with genes involved in pentose-phosphate pathway)

IT Plasmid vectors

(pBS(AR2T), D-ribulose-5-phosphate epimerase **gene** on; manufacture of five-carbon sugars and sugar alcs. using microorganisms deficient in or transformed with **genes** involved in pentose-phosphate pathway)

IT Plasmid vectors

(pBS(AR2T)-kan, D-ribulose-5-phosphate epimerase **gene** on; manufacture of five-carbon sugars and sugar alcs. using microorganisms deficient in or transformed with **genes** involved in pentose-phosphate pathway)

IT Plasmid vectors

(pBS, cloning vector; manufacture of five-carbon sugars and sugar alcs. using microorganisms deficient in or transformed with genes involved in pentose-phosphate pathway)

IT Plasmid vectors

(pGT21, cloning vector; manufacture of five-carbon sugars and sugar alcs. using microorganisms deficient in or transformed with **genes** involved in pentose-phosphate pathway)

IT Plasmid vectors

(pGT23, cloning vector; manufacture of five-carbon sugars and sugar alcs. using microorganisms deficient in or transformed with **genes** involved in pentose-phosphate pathway)

IT Plasmid vectors

(pGTK24 (MXD2), xylitol dehydrogenase gene on; manufacture of five-carbon sugars and sugar alcs. using microorganisms deficient in or transformed with genes involved in pentose-phosphate pathway)

IT **Plasmid** vectors

(pGTK24, cloning vector; manufacture of five-carbon sugars and sugar alcs. using microorganisms deficient in or transformed with genes involved in pentose-phosphate pathway)

IT **Plasmid** vectors

(pTKT:E1, cloning vector; manufacture of five-carbon sugars and sugar alcs. using microorganisms deficient in or transformed with genes involved in pentose-phosphate pathway)

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IT
     Glycolysis
        (partial blocking through PPP enzymes; manufacture of five-carbon sugars and
        sugar alcs. using microorganisms deficient in or transformed with
        genes involved in pentose-phosphate pathway)
     Alcohols, preparation
IT
     RL: BMF (Bioindustrial manufacture); BIOL (Biological study); PREP
     (Preparation)
        (polyhydric; manufacture of five-carbon sugars and sugar alcs. using
        microorganisms deficient in or transformed with genes
        involved in pentose-phosphate pathway)
     Saccharomyces cerevisiae
IT
        (redox enzymes of, expression in Pichia of genes for; manufacture
        of five-carbon sugars and sugar alcs. using microorganisms deficient in
        or transformed with genes involved in pentose-phosphate
        pathway)
     Klebsiella pneumoniae
IT
        (ribitol dehydrogenase gene of; manufacture of five-carbon sugars
        and sugar alcs. using microorganisms deficient in or transformed with
        genes involved in pentose-phosphate pathway)
IT
     Gene, microbial
     RL: BSU (Biological study, unclassified); BUU (Biological use,
     unclassified); BIOL (Biological study); USES (Uses)
        (rpi, for D-ribose-phosphate isomerase; manufacture of five-carbon sugars
        and sugar alcs. using microorganisms deficient in or transformed with
        genes involved in pentose-phosphate pathway)
     Carbohydrates, preparation
IT
     RL: BMF (Bioindustrial manufacture); BIOL (Biological study); PREP
     (Preparation)
        (sugar phosphates; manufacture of five-carbon sugars and sugar alcs. using
        microorganisms deficient in or transformed with genes
        involved in pentose-phosphate pathway)
     Gene, microbial
IT
     RL: BSU (Biological study, unclassified); BUU (Biological use,
     unclassified); BIOL (Biological study); USES (Uses)
        (tkt, for transketolase; manufacture of five-carbon sugars and sugar alcs.
        using microorganisms deficient in or transformed with genes
        involved in pentose-phosphate pathway)
     Gene, microbial
IT
     RL: BSU (Biological study, unclassified); BUU (Biological use,
     unclassified); BIOL (Biological study); USES (Uses)
        (tsr, or fba, for aldolase; manufacture of five-carbon sugars and sugar
        alcs. using microorganisms deficient in or transformed with
        genes involved in pentose-phosphate pathway)
IT
     Zymomonas mobilis
        (zwf gene of; manufacture of five-carbon sugars and sugar alcs.
        using microorganisms deficient in or transformed with genes
        involved in pentose-phosphate pathway)
IT
     Gene, microbial
     RL: BSU (Biological study, unclassified); BUU (Biological use,
     unclassified); BIOL (Biological study); USES (Uses)
        (zwf, in regulation of PPP oxidative capacity; manufacture of five-carbon
        sugars and sugar alcs. using microorganisms deficient in or transformed
        with genes involved in pentose-phosphate pathway)
                                 351916-74-0
                                               351916-75-1
                                                              351916-76-2
IT
     311826-80-9
                   311826-82-1
     351916-79-5
     RL: BSU (Biological study, unclassified); BUU (Biological use,
     unclassified); PRP (Properties); BIOL (Biological study); USES (Uses)
        (amino acid sequence; manufacture of five-carbon sugars and sugar alcs.
        using microorganisms deficient in or transformed with genes
        involved in pentose-phosphate pathway)
IT
                   351916-83-1
     351916-81-9
     RL: BUU (Biological use, unclassified); BIOL (Biological study); USES
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(Uses)

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(amino acid sequence; manufacture of five-carbon sugars and sugar alcs.
        using microorganisms deficient in or transformed with genes
        involved in pentose-phosphate pathway)
     64-17-5P, Ethanol, preparation
IT
    RL: BMF (Bioindustrial manufacture); BIOL (Biological study); PREP
     (Preparation)
        (fermentation of; manufacture of five-carbon sugars and sugar alcs. using
        microorganisms deficient in or transformed with genes
        involved in pentose-phosphate pathway)
     50-99-7, D-Glucose, biological studies
IT
     RL: BPR (Biological process); BSU (Biological study, unclassified); BIOL
     (Biological study); PROC (Process)
        (five carbon sugar or sugar alc. fermentation from; manufacture of
five-carbon
        sugars and sugar alcs. using microorganisms deficient in or transformed
        with genes involved in pentose-phosphate pathway)
     65187-56-6, 2-Deoxyglucose-6-phosphate phosphatase
IT
     RL: BUU (Biological use, unclassified); BIOL (Biological study); USES
     (Uses)
        (gene DOG1; manufacture of five-carbon sugars and sugar alcs.
        using microorganisms deficient in or transformed with genes
        involved in pentose-phosphate pathway)
     79747-53-8, Phosphatase, phosphoprotein (phosphotyrosine)
IT
     RL: BSU (Biological study, unclassified); BUU (Biological use,
     unclassified); PRP (Properties); BIOL (Biological study); USES (Uses)
        (gene PPPase1 or PPPase2; manufacture of five-carbon sugars and
        sugar alcs. using microorganisms deficient in or transformed with
        genes involved in pentose-phosphate pathway)
     9001-46-1, NAD-dependent glutamate dehydrogenase
IT
     RL: BSU (Biological study, unclassified); PRP (Properties); BIOL
     (Biological study)
        (gene for, of Saccharomyces cerevisiae, cloning and
        expression of; manufacture of five-carbon sugars and sugar alcs. using
        microorganisms deficient in or transformed with genes
        involved in pentose-phosphate pathway)
     9028-18-6, Arabitol dehydrogenase
IT
     RL: BUU (Biological use, unclassified); BIOL (Biological study); USES
     (Uses)
        (gene for; manufacture of five-carbon sugars and sugar alcs. using
        microorganisms deficient in or transformed with genes
        involved in pentose-phosphate pathway)
     60063-83-4, L-Fucose isomerase
IT
     RL: BUU (Biological use, unclassified); BIOL (Biological study); USES
        (gene fucl; manufacture of five-carbon sugars and sugar alcs.
        using microorganisms deficient in or transformed with genes
        involved in pentose-phosphate pathway)
     9001-40-5, Glucose-6-phosphate dehydrogenase
                                                    9001-82-5,
IT
     6-Phosphogluconate dehydrogenase
     RL: BUU (Biological use, unclassified); BIOL (Biological study); USES
     (Uses)
        (in regulation of PPP oxidative capacity; manufacture of five-carbon sugars
        and sugar alcs. using microorganisms deficient in or transformed with
        genes involved in pentose-phosphate pathway)
                             9001-41-6, Phosphoglucoisomerase
                                                                  9001-51-8,
IT
     9001-36-9, Glucokinase
                  37278-03-8, Phosphofructokinase
     Hexokinase
     RL: BUU (Biological use, unclassified); BIOL (Biological study); USES
        (in regulation of glucose uptake and glucose carbon flow into PPP;
        manufacture of five-carbon sugars and sugar alcs. using microorganisms
        deficient in or transformed with genes involved in
        pentose-phosphate pathway)
                        488-81-3P, Ribitol
                                              488-84-6P, D-Ribulose
     87-99-0P, Xylitol
IT
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2152-56-9P, Arabitol
                          1114-34-7P, D-Lyxose
    551-84-8P, Xylulose
    4151-19-3P, Ribulose-5-phosphate 4212-65-1P, Xylulose-5-phosphate
    4300-28-1P, Ribose 5-phosphate 10323-20-3P, D-Arabinose
    RL: BMF (Bioindustrial manufacture); BIOL (Biological study); PREP
     (Preparation)
        (manufacture of five-carbon sugars and sugar alcs. using microorganisms
       deficient in or transformed with genes involved in
       pentose-phosphate pathway)
    58-86-6P, Xylose, biological studies
IT
    RL: BMF (Bioindustrial manufacture); BPR (Biological process); BSU
     (Biological study, unclassified); BIOL (Biological study); PREP
     (Preparation); PROC (Process)
        (manufacture of five-carbon sugars and sugar alcs. using microorganisms
       deficient in or transformed with genes involved in
       pentose-phosphate pathway)
                                                     58-68-4, NADH
                                                                     64-69-7
                                      53-84-9, NAD
     53-57-6, NADPH
                      53-59-8, NADP
IT
     9001-50-7, Glyceraldehyde-3-phosphate dehydrogenase
                                                           79082-92-1,
    Fructose-2,6-bisphosphate
    RL: BPR (Biological process); BSU (Biological study, unclassified); BIOL
     (Biological study); PROC (Process)
        (manufacture of five-carbon sugars and sugar alcs. using microorganisms
       deficient in or transformed with genes involved in
        pentose-phosphate pathway)
                               64886-68-6, Xylitol-phosphate
     9030-58-4, Xylulokinase
IT
     dehydrogenase
     RL: BSU (Biological study, unclassified); BUU (Biological use,
     unclassified); PRP (Properties); BIOL (Biological study); USES (Uses)
        (manufacture of five-carbon sugars and sugar alcs. using microorganisms
        deficient in or transformed with genes involved in
        pentose-phosphate pathway)
     9001-80-3, Kinase (phosphorylating), phosphofructo-
                                                           9014-23-7, Ribitol
IT
                     9014-48-6, Transketolase
                                                9023-83-0, Isomerase, ribose
     dehydrogenase
                 9024-20-8, D-Ribulose 5-phosphate Epimerase 9028-16-4
     phosphate
      Xylitol dehydrogenase
                               9031-25-8, D-Mannose
     isomerase 99775-25-4, Xylose reductase
     150316-09-9, D-Ketohexose 3-epimerase
     RL: BUU (Biological use, unclassified); BIOL (Biological study); USES
     (Uses)
        (manufacture of five-carbon sugars and sugar alcs. using microorganisms
        deficient in or transformed with genes involved in
        pentose-phosphate pathway)
     6917-36-8P, Pentitol
IT
     RL: BMF (Bioindustrial manufacture); BIOL (Biological study); PREP
     (Preparation)
        (manufacture of; manufacture of five-carbon sugars and sugar alcs. using
        microorganisms deficient in or transformed with genes
        involved in pentose-phosphate pathway)
                   351916-82-0
IT
     351916-80-8
     RL: BSU (Biological study, unclassified); BUU (Biological use,
     unclassified); PRP (Properties); BIOL (Biological study); USES (Uses)
        (nucleotide sequence; manufacture of five-carbon sugars and sugar alcs.
        using microorganisms deficient in or transformed with genes
        involved in pentose-phosphate pathway)
IT
     351916-77-3
                   351916-78-4
     RL: BSU (Biological study, unclassified); BUU (Biological use,
     unclassified); PRP (Properties); BIOL (Biological study); USES (Uses)
        (nucleotide sequences; manufacture of five-carbon sugars and sugar alcs.
        using microorganisms deficient in or transformed with genes
        involved in pentose-phosphate pathway)
     9024-52-6, Aldolase
IT
     RL: BUU (Biological use, unclassified); BIOL (Biological study); USES
```

(promoter from the gene for; manufacture of five-carbon sugars and

```
sugar alcs. using microorganisms deficient in or transformed with
       genes involved in pentose-phosphate pathway)
                              9028-86-8, Aldehyde dehydrogenase
     9028-46-0, Malic enzyme
IT
     104118-53-8, Xylose reductase
    RL: BPR (Biological process); BSU (Biological study, unclassified); CAT
     (Catalyst use); BIOL (Biological study); PROC (Process); USES (Uses)
        (redox system using; manufacture of five-carbon sugars and sugar alcs. using
        microorganisms deficient in or transformed with genes
        involved in pentose-phosphate pathway)
                                              351919-62-5
                                                             351919-63-6
                                351919-61-4
                  351919-60-3
     351919-59-0
IT
                                 351919-66-9
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     351919-64-7
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                  351919-70-5
     351919-69-2
                                             351919-77-2
                                                             351919-78-3
                  351919-75-0
                                 351919-76-1
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                                             351919-82-9
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     351919-84-1
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                                                             351919-93-2
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     351919-89-6
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     351919-94-3
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                                 351920-04-2
     351920-01-9
                  351920-03-1
                                                             351920-11-1, 1:
                                              351920-10-0
                                 351920-09-7
     351920-07-5
                  351920-08-6
     PN: WO0153306 SEQID: 1 unclaimed DNA
     RL: PRP (Properties)
        (unclaimed nucleotide sequence; manufacture of five-carbon sugars and sugar
        alcs. using microorganisms deficient in or transformed with
        genes involved in pentose-phosphate pathway)
     351919-99-8
IT
     RL: PRP (Properties)
        (unclaimed protein sequence; manufacture of five-carbon sugars and sugar
        alcs. using microorganisms deficient in or transformed with
        genes involved in pentose-phosphate pathway)
                                               351900-51-1
                                                             351900-52-2
                   351900-49-7
                                351900-50-0
     351900-48-6
IT
                   351900-54-4
                                 351900-55-5
     351900-53-3
     RL: PRP (Properties)
        (unclaimed sequence; manufacture of five-carbon sugars and sugar alcs. using
        microorganisms deficient in or transformed with genes
        involved in pentose-phosphate pathway)
     ANSWER 4 OF 33 HCAPLUS COPYRIGHT 2004 ACS on STN
     1997:746143 HCAPLUS
ΑN
     128:2978
DN
     Entered STN: 27 Nov 1997
ED
     Stable recombinant yeasts for fermenting xylose to ethanol
ΤI
     Ho, Nancy W. Y.; Chen, Zheng-Dao
IN
     Purdue Research Foundation, USA; Ho, Nancy W. Y.; Chen, Zheng-Dao
PA
     PCT Int. Appl., 66 pp.
SO
     CODEN: PIXXD2
     Patent
DT
     English
LA
     ICM C12N001-16
IC
          Cl2N001-18; Cl2N001-19; Cl2N015-09; Cl2N015-68; Cl2N015-69;
          C12N015-81; C12P007-06
     16-5 (Fermentation and Bioindustrial Chemistry)
     Section cross-reference(s): 3
 FAN.CNT 1
                                           APPLICATION NO. DATE
     PATENT NO.
                      KIND DATE
                                           _____
                                           WO 1997-US7663 19970506 <--
                      A1 19971113
ÞΙ
             AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE,
             DK, EE, ES, FI, GB, GE, HU, IL, IS, JP, KE, KG, KP, KR, KZ, LC,
             LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT,
             RO, RU, SD, SE, SG, SI, SK, TJ, TM, TR, TT, UA, UG, US, UZ, VN,
             YU, AM, AZ, BY, KG, KZ, MD, RU, TJ, TM
         RW: GH, KE, LS, MW, SD, SZ, UG, AT, BE, CH, DE, DK, ES, FI, FR, GB,
             GR, IE, IT, LU, MC, NL, PT, SE, BF, BJ, CF, CG, CI, CM, GA, GN,
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ML, MR, NE, SN, TD, TG
                                                            19970506 <--
                                          AU 1997-28301
                           19971126
    AU 9728301
                      A1
                            20010322
                      B2
    AU 731102
                                          EP 1997-922698
                                                          19970506 <--
                            19990303
    EP 898616
                      Α1
        R: AT, BE, DE, DK, ES, FR, GB, GR, IT, NL, SE, PT, IE, FI
                                          CN 1997-196195
                                                          19970506 <--
                     Α
                            19990804
    CN 1225125
                                                            19970506 <--
                                           JP 1997-540153
                            20000808
    JP 2000509988
                      T2
                                           BR 1997-10963
                                                            19970506 <--
    BR 9710963
                      Α
                            20010731
                            19960506 <--
PRAI US 1996-16865P
                      Р
                      W
                                     <--
                            19970506
    WO 1997-US7663
    Described are recombinant yeast which ferment xylose to EtOH and
AΒ
    which maintain their ability to do so when cultured for numerous
    generations in non-selective media. The preferred yeast contain
    multiple copies of integrated genes encoding xylose
    reductase, xylitol dehydrogenase, and
    xylulokinase fused to promoters which are non-glucose inhibited
     and which do not require xylose for induction. Also described are
    preferred methods for integrating multiple copies of exogenous DNA
     into host cells by transforming cells with replicative/integrative
    vectors, and then replicating the cells a number of times under selective
    pressure to promote retention of the vector in subsequent generations.
    The replicated vectors thus serve to integrate multiple copies of the
     exogenous DNA into the host cells throughout the
     replication/selection phase. Thereafter the selective pressure can be
     removed to promote loss of the vector in subsequent generations, leaving
     stable integrants of the exogenous DNA.
    Saccharomyces recombinant ethanol fermn xylose
ST
    Genetic engineering
IT
       Saccharomyces cerevisiae
        (stable recombinant yeasts for fermenting xylose to ethanol)
     64-17-5P, Ethanol, preparation
IT
     RL: BMF (Bioindustrial manufacture); BPN (Biosynthetic preparation); BIOL
     (Biological study); PREP (Preparation)
        (stable recombinant yeasts for fermenting xylose to ethanol)
IT
     9028-16-4, Xylitol dehydrogenase
     9030-58-4, Xylulokinase 99775-25-4,
     Xylose reductase
     RL: BOC (Biological occurrence); BPR (Biological process); BSU (Biological
     study, unclassified); BUU (Biological use, unclassified); BIOL (Biological
     study); OCCU (Occurrence); PROC (Process); USES (Uses)
        (stable recombinant yeasts for fermenting xylose to ethanol)
     58-86-6, D-Xylose, biological studies
TТ
     RL: BPR (Biological process); BSU (Biological study, unclassified); RCT
     (Reactant); BIOL (Biological study); PROC (Process); RACT (Reactant or
     reagent)
        (stable recombinant yeasts for fermenting xylose to ethanol)
     ANSWER 5 OF 33 HCAPLUS COPYRIGHT 2004 ACS on STN
L49
     1997:464186 HCAPLUS
AN
     127:94156
DN
     Entered STN: 24 Jul 1997
ED
     Fermentation of corn fiber sugars by an engineered xylose utilizing
TI
     Saccharomyces yeast strain
     Moniruzzaman, M.; Dien, B.S.; Skory, C.D.; Chen, Z.D.; Hespell,
ΑU
     R.B.; Ho, N.W.Y.; Dale, B.E.; Bothast, R.J.
     Department of Chemical Engineering, Texas AandM University, College
CS
     Station, TX, 77843, USA
     World Journal of Microbiology & Biotechnology (1997), 13(3), 341-346
SO
     CODEN: WJMBEY; ISSN: 0959-3993
     Rapid Science Publishers
PB
DT
     Journal
```

LΑ

CC

English

16-5 (Fermentation and Bioindustrial Chemistry)

The ability of a recombinant Saccharomyces yeast strain to AB ferment the sugars glucose, xylose, arabinose and galactose which are the predominant monosaccharides found in corn fiber hydrolyzates has been examined Saccharomyces strain 1400 (pLNH32) was genetically engineered to ferment xylose by expressing genes encoding a xylose reductase, a xylitol dehydrogenase and a xylulose kinase. The recombinant efficiently fermented xylose alone or in the presence of glucose. Xylose-grown cultures had very little difference in xylitol accumulation, with only 4 to 5 g/L accumulating, in aerobic, micro-aerated and anaerobic conditions. Highest production of ethanol with all sugars was achieved under anaerobic conditions. From a mixture of glucose (80 g/L) and xylose (40 g/L), this strain produced 52 g/L ethanol, equivalent to 85% of theor. yield, in less than 24 h. Using a mixture of glucose (31 g/L), xylose (15.2 g/L), arabinose (10.5 g/L) and galactose (2 g/L), all of the sugars except arabinose were consumed in 24 h with an accumulation of 22 g ethanol/L, a 90% yield (excluding the arabinose in the calcn. since it is not fermented). Approx. 98% theor. yield, or 21 g ethanol/L, was achieved using an enzymic hydrolyzate of ammonia fiber exploded corn fiber containing an estimated 47.0 g mixed sugars/L. In all mixed sugar fermns., less than 25% arabinose was consumed and converted into arabitol. corn fiber sugar fermn Saccharomyces xylose; ethanol fermn corn fiber ST xylose yeast IT Fermentation (fermentation of corn fiber sugars by engineered xylose utilizing Saccharomyces **yeast** strain) Carbohydrates, biological studies IT RL: BPR (Biological process); BSU (Biological study, unclassified); BIOL (Biological study); PROC (Process) (fermentation of corn fiber sugars by engineered xylose utilizing Saccharomyces **yeast** strain) ITCorn (fiber; fermentation of corn fiber sugars by engineered xylose utilizing Saccharomyces **yeast** strain) IT 64-17-5P, Ethanol, preparation RL: BMF (Bioindustrial manufacture); BIOL (Biological study); PREP (Preparation) (fermentation; fermentation of corn fiber sugars by engineered xylose utilizing Saccharomyces yeast strain) 50-99-7, Glucose, biological studies 58-86-6, Xylose, biological studies IT 59-23-4, Galactose, biological studies 147-81-9, Arabinose RL: BPR (Biological process); BSU (Biological study, unclassified); BIOL (Biological study); PROC (Process) (fermentation of corn fiber sugars by engineered xylose utilizing Saccharomyces yeast strain) ANSWER 6 OF 33 HCAPLUS COPYRIGHT 2004 ACS on STN L49 1996:671095 HCAPLUS AN126:2045 DNEntered STN: 13 Nov 1996 EDYeast Sequencing Reports: Sequence and analysis of an aldose (ΤI xylose) reductase gene from the xylose-fermenting yeast Pachysolen tannophilus Bolen, Paul L.; Hayman, G. Thomas; Shepherd, Hurley S. ΑU Microbial Properties Res., National Center Agricultural Utilization Res., CS Agricultural Res. Serv., U.S. Dep. Agriculture, Peoria, IL, 61604, USA Yeast (1996), 12(13), 1367-1375 SO

LA English
CC 3-3 (Biochemical Genetics)

PB

DT

Wiley Journal

CODEN: YESTE3; ISSN: 0749-503X

Section cross-reference(s): 7, 10 A xylose reductase gene was isolated from AB the xylose-fermenting yeast Pachysolen tannophilus as a cDNA clone by selecting clones that hybridized specifically to xylose-inducible mRNA. Use of the cDNA clone as a probe in Northern hybridizations identified a xylose-inducible mRNA. Use of the cDNA clone as a probe in Northern hybridizations identified a xylose-inducible mRNA species large enough to encode a 36 kDa xylose reductase protein known to be produced by this yeast. A corresponding genomic clone was isolated as a 3 kb EcoRI fragment that specifically hybridized to the cDNA clone. The sequence of the cDNA and the largest open reading frame of the genomic clone are identical. The predicted translation product exhibits: (1) significant sequence identity with a previously published N-terminal amino acid sequence from purified P. tannophilus xylose (aldose) reductase protein exhibiting NADH/NADHP-dependent activities (aldose reductase, EC 1.1.1.21); (2) identity with a protein composed of 317 amino acid residues with a calculated mol. mass of 36.2 kDa, equivalent to that reported for purified P. tannophilus xylose reductase; and (3) considerable sequence similarity to, and features of, a superfamily of oxidoreductases. This sequence is deposited as GenBank Accession Number U40706. DNA sequence Pachysolen aldose reductase ST gene; aldose reductase protein sequence Pachysolen TT **DNA** sequences Pachysolen tannophilus Protein sequences cDNA sequences (sequence and anal. of an aldose (xylose) reductase gene from the xylose-fermenting yeast Pachysolen tannophilus) IT 183327-22-2 RL: BOC (Biological occurrence); BSU (Biological study, unclassified); PRP (Properties); BIOL (Biological study); OCCU (Occurrence) (amino acid sequence; anal. of an aldose (xylose) reductase gene from the xylose-fermenting yeast Pachysolen tannophilus) 58-86-6, Xylose, biological studies IT RL: BAC (Biological activity or effector, except adverse); BSU (Biological study, unclassified); BIOL (Biological study) (anal. of an aldose (xylose) reductase gene from the xylose-fermenting yeast Pachysolen tannophilus) IT 183399-64-6 RL: BOC (Biological occurrence); BSU (Biological study, unclassified); PRP (Properties); BIOL (Biological study); OCCU (Occurrence) (nucleotide sequence; anal. of an aldose (xylose) reductase gene from the xylose-fermenting yeast Pachysolen tannophilus) IT 9028-31-3, Aldose reductase RL: BSU (Biological study, unclassified); PRP (Properties); BIOL (Biological study) (sequence and anal. of an aldose (xylose) reductase gene from the xylose-fermenting yeast Pachysolen tannophilus) ANSWER 7 OF 33 HCAPLUS COPYRIGHT 2004 ACS on STN 1996:606013 HCAPLUS AN125:270220 DNED Entered STN: 11 Oct 1996 A glycerol-3-phosphate dehydrogenase-deficient mutant of Saccharomyces ТT cerevisiae expressing the heterologous XYL1 gene

Liden, G.; Walfridsson, M.; Ansell, R.; Anderlund, M.; Adler, L.;

ΑU

Hahn-Haegerdal, B.

- CS Dep. Chem. Reaction Eng., Chalmers Univ. Technol., Goeteborg, S-412 96, Swed.
- SO Applied and Environmental Microbiology (1996), 62(10), 3894-3896 CODEN: AEMIDF; ISSN: 0099-2240
- PB American Society for Microbiology
- DT Journal
- LA English
- CC 10-4 (Microbial, Algal, and Fungal Biochemistry) Section cross-reference(s): 3
- The gene XYL1, encoding a xylose reductase, from Pichia stipitis was transformed into a mutant of Saccharomyces cerevisiae incapable of glycerol production because of deletion of the genes GPD1 and GPD2. The transformed strain was capable of anaerobic glucose conversion in the presence of added xylose, indicating that the xylose reductase reaction can fulfill the role of the glycerol-3-phosphate dehydrogenase reaction as a redox sink. The specific xylitol production rate obtained was 0.38 g g-1 h-1.
- ST Pichia xylose reductase gene cloning Saccharomyces; yeast gene XYL1 cloning glycerol dehydrogenase
- IT Molecular cloning

Saccharomyces cerevisiae

Yamadazyma stipitis

(glycerol-3-phosphate dehydrogenase-deficient mutant of Saccharomyces cerevisiae expressing heterologous XYL1 gene)

Saccharomyces cerevisiae expressing heterologous XYL1 gene)

IT 95829-40-6, Xylose reductase

RL: BSU (Biological study, unclassified); BIOL (Biological study)
(glycerol-3-phosphate dehydrogenase-deficient mutant of Saccharomyces
cerevisiae expressing heterologous XYL1 gene)

- L49 ANSWER 8 OF 33 HCAPLUS COPYRIGHT 2004 ACS on STN
- AN 1996:308481 HCAPLUS
- DN 124:340999
- ED Entered STN: 25 May 1996
- TI A metabolic engineering view on molecular breeding of an alcohol fermenting yeast from xylose
- AU Seki, Tatsuji; Tantirungkij, Manee; Fujiyama, Kazuhito; Yoshida, Toshiomi
- CS International Center Cooperative Research Biotechnology, Osaka University, Suita, 565, Japan
- Environmental Biotechnology: Principles and Applications, [Papers presented at the International Symposium on Environmental Biotechnology], Waterloo, Ont., July 4-8, 1994 (1996), Meeting Date 1994, 114-124. Editor(s): Moo-Young, Murray; Anderson, William A.; Chakrabarty, Ananda M. Publisher: Kluwer, Dordrecht, Neth. CODEN: 62UGA4
- DT Conference
- LA English
- CC 16-5 (Fermentation and Bioindustrial Chemistry) Section cross-reference(s): 3
- AB Xylose-assimilating S. cerevisiae was constructed by introducing the xylose reductase and xylitol dehydrogenase genes originating from P. stipitis. Good

growth of the transformant in xylose medium was observed under aerobic conditions. Under a limited oxygen condition, the transformant produced a lesser amount of ethanol than P. stipitis, and a remarkable amount of xylitol was accumulated. A mutant, IM2, in which the ratio of xylose reductase to xylitol dehydrogenase activities was lower than the parental strain, exhibited an improved fermentation with

accumulation of xylitol and a higher yield. The limited feeding of xylose could also improve the fermentation, with reduced xylitol accumulation as well as increased ethanol yield. The facts suggest strongly that the path of the conversion from xylitol to xylulose is the "bottleneck" due to a poor regeneration of NAD essential for the conversion. An appropriate oxygen supply also improved the ethanol production and the production rate,

suggesting it

less

may contribute to the NAD recycle from NADH.

ST ethanol manuf Saccharomyces xylose

IT Fermentation

Genetic engineering

Saccharomyces cerevisiae

(genetic engineering of yeast for ethanol fermentation from xylose)

IT 58-86-6, Xylose, biological studies RL: BAC (Biological activity or effector, except adverse); BSU (Biological study, unclassified); BIOL (Biological study)

(genetic engineering of yeast for ethanol fermentation from xylose)

IT 64-17-5P, Ethanol, preparation
 RL: BMF (Bioindustrial manufacture); BIOL (Biological study); PREP
 (Preparation)

(genetic engineering of yeast for ethanol fermentation from xylose)

IT 87-99-0P, Xylitol

RL: BYP (Byproduct); PREP (Preparation)
(genetic engineering of yeast for ethanol fermentation from xylose)

L49 ANSWER 9 OF 33 HCAPLUS COPYRIGHT 2004 ACS on STN

AN 1996:158446 HCAPLUS

DN 124:252209

ED Entered STN: 19 Mar 1996

TI Sequencing and analysis of 51 kb on the right arm of **chromosome**XV from Saccharomyces cerevisiae reveals 30 open reading frames

AU Wieman, Stefan; Rechmann, Stefanie; Benes, Vladimir; Voss, Hartmut; Schwager, Christian; Vlcek, Cestmir; Stegemann, Josef; Zimmermann, Jurgen; Erfle, Holger; et al.

CS EMBL, Heidelberg, D-69117, Germany

SO Yeast (1996), 12(3), 281-8 CODEN: YESTE3; ISSN: 0749-503X

PB Wiley

DT Journal

LA English

CC 3-3 (Biochemical Genetics)

- AB We have sequenced a region of 51 kb of the right arm from chromosome XV of Saccharomyces cerevisiae. The sequence contains 30 open reading frames (ORFs) of more than 100 amino acid residues. Thirteen new genes have been identified. Thirteen ORFs correspond to known yeast genes. One delta element and one tRNA gene were identified. Upstream of the RPO31 gene, encoding the largest subunit of RNA polymerase III, lies a Abflp binding site. The nucleotide sequence data reported in this paper are available in the EMBL, GenBank and DDBJ nucleotide sequence databases under the Accession Number X90518.
- ST Saccharomyces chromosome XV gene sequence

IT Deoxyribonucleic acid sequences
Protein sequences

Saccharomyces cerevisiae

(sequencing and anal. of 51 kb on right arm of chromosome XV

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from Saccharomyces cerevisiae reveals 30 open reading frames)
IT
     Gene, microbial
     RL: BSU (Biological study, unclassified); BIOL (Biological study)
        (sequencing and anal. of 51 kb on right arm of chromosome XV
        from Saccharomyces cerevisiae reveals 30 open reading frames)
IT
     Chromosome
        (Saccharomyces cerevisiae XV, sequencing and anal. of 51 kb on right
        arm of chromosome XV from Saccharomyces cerevisiae reveals 30
        open reading frames)
     Ribonucleic acids, transfer
IT
     RL: BSU (Biological study, unclassified); BIOL (Biological study)
        (aspartic acid-specific, gene for; sequencing and anal. of 51
        kb on right arm of chromosome XV from Saccharomyces
        cerevisiae reveals 30 open reading frames)
     Protein formation elongation factors
IT
     RL: BSU (Biological study, unclassified); BIOL (Biological study)
        (eEF-2, gene for; sequencing and anal. of 51 kb on right arm
        of chromosome XV from Saccharomyces cerevisiae reveals 30
        open reading frames)
IT
     9014-24-8
     RL: BSU (Biological study, unclassified); BIOL (Biological study)
        (III, gene for; sequencing and anal. of 51 kb on right arm of
        chromosome XV from Saccharomyces cerevisiae reveals 30 open
        reading frames)
     121548-71-8, Protein (Saccharomyces cerevisiae gene GCY
IT
                122178-43-2, Profilin (Saccharomyces cerevisiae gene
     reduced)
                          146313-69-1, Protein formation elongation factor EF 2
            133758-72-2
     PFY)
     (Saccharomyces cerevisiae strain YM213 gene EFT1 reduced)
     156656-75-6
                   157712-16-8
                                 159521-43-4
                                               172929-93-0
                                                              172929-97-4
                                 174763-95-2
                                                174763-96-3
                                                              174763-97-4
     174763-93-0
                   174763-94-1
                                 174764-00-2
                                               174764-01-3
                                                              174764-02-4
     174763-98-5
                   174763-99-6
                                 174764-05-7
                                               174764-06-8
     174764-03-5
                   174764-04-6
     RL: PRP (Properties)
        (amino acid sequence; sequencing and anal. of 51 kb on right arm of
        chromosome XV from Saccharomyces cerevisiae reveals 30 open
        reading frames)
     86480-67-3
ΙT
     RL: BSU (Biological study, unclassified); BIOL (Biological study)
        (gene for; sequencing and anal. of 51 kb on right arm of
        chromosome XV from Saccharomyces cerevisiae reveals 30 open
        reading frames)
     170321-17-2, GenBank X90518
IT
     RL: PRP (Properties)
        (nucleotide sequence; sequencing and anal. of 51 kb on right arm of
        chromosome XV from Saccharomyces cerevisiae reveals 30 open
        reading frames)
    ANSWER 10 OF 33 HCAPLUS COPYRIGHT 2004 ACS on STN
L49
     1995:814692 HCAPLUS
AN
     124:2028
DN
                   27 Sep 1995
ED
     Entered STN:
     Isolation and characterization of the gene encoding
TI
     xylose reductase from Kluyveromyces lactis
     Billard, Patrick; Menart, Sandrine; Fleer, Reinhard; Bolotin-Fukuhara,
ΑU
     Monique
     Institut de Genetique et Microbiologie, Bat. 400, Universite Paris-Sud,
CS
     91405, Orsay, Fr.
     Gene (1995), 162(1), 93-7
SO
     CODEN: GENED6; ISSN: 0378-1119
     Elsevier
PB
     Journal
DT
     English
LΑ
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3-3 (Biochemical Genetics)

CC

```
Section cross-reference(s): 7, 10
     The identification of a xylose reductase (XR) - encoding
AB
     gene (XYL1) from the xylose-assimilating yeast
Kluyveromyces lactis (Kl) is described. XYL1 was isolated as a highly
     expressed fusion clone from a 'lacZ translational fusion library.
     DNA sequence anal. revealed an open reading frame (ORF) of 987bp
     capable of encoding a polypeptide of 329 amino acids (aa). The deduced aa
     sequence displays a 62% overall identity to that of XR from Pichia
     stipitis. Gene disruption studies indicate that XYL1 exists as
     a single copy in the yeast genome and is essential for growth on
     xylose. Northern blot anal. of the XYL1 transcript and measurement of the
     XR enzymic activities show, in contrast to other known XR-encoding
     genes, a constitutive expression of Kl XYL1.
ST
     Kluyveromyces xylose reductase gene
     sequence; XYL1 gene Kluyveromyces xylose
     reductase sequence
IT
     Kluyveromyces lactis
        (isolation and characterization of gene encoding
        xylose reductase from Kluyveromyces lactis)
     Deoxyribonucleic acid sequences
IT
        (of gene XYL1 from Kluyveromyces lactis)
     Protein sequences
IT
        (of xylose reductase from Kluyveromyces lactis)
IT
     Gene, microbial
     RL: PRP (Properties)
        (XYL1, isolation and characterization of gene encoding
        xylose reductase from Kluyveromyces lactis)
     171043-09-7
IT
     RL: PRP (Properties)
        (amino acid sequence; isolation and characterization of gene
        encoding xylose reductase from Kluyveromyces
        lactis)
     58-86-6, Xylose, biological studies
TI
     RL: BPR (Biological process); BSU (Biological study, unclassified); BIOL
     (Biological study); PROC (Process)
        (isolation and characterization of gene encoding
        xylose reductase from Kluyveromyces lactis)
     104118-53-8, Xylose reductase
ΤT
     RL: PRP (Properties)
        (isolation and characterization of gene encoding
        xylose reductase from Kluyveromyces lactis)
     158764-02-4, GenBank L36993
IT
     RL: PRP (Properties)
        (nucleotide sequence; isolation and characterization of gene
        encoding xylose reductase from Kluyveromyces
        lactis)
     53-57-6, NADPH
TТ
     RL: BPR (Biological process); BSU (Biological study, unclassified); BIOL
     (Biological study); PROC (Process)
        (xylose reductase from Kluyveromyces lactis
        dependence on NADPH)
     ANSWER 11 OF 33 HCAPLUS COPYRIGHT 2004 ACS on STN
L49
     1995:756362 HCAPLUS
AN
     123:196764
DN
     Entered STN: 25 Aug 1995
ED
     Recombinant yeasts for effective fermentation of glucose and
ΤI
     xylose
     Ho, Nancy W. Y.; Tsao, George T.
IN
     Purdue Research Foundation, USA
PΑ
     PCT Int. Appl., 62 pp.
SO
     CODEN: PIXXD2
```

Patent

DΤ

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LA
    English
IC
     ICM C12N001-14
     ICS C12N009-00; C12N009-12; C12N015-00; C12P007-08
     16-5 (Fermentation and Bioindustrial Chemistry)
CC
FAN.CNT 2
                                           APPLICATION NO.
     PATENT NO.
                      KIND DATE
                           _____
                      _ _ _ _
     _____
                                     WO 1994-US12861 19941108 <--
    WO 9513362 A1 19950518
PΤ
        W: AM, AU, BB, BG, BR, BY, CA, CN, CZ, EE, FI, GE, HU, JP, KG, KP, KR, KZ, LK, LR, LT, LV, MD, MG, MN, NO, NZ, PL, RO, RU, SI, SK,
             TJ, TT, UA, UZ, VN
         RW: KE, MW, SD, SZ, AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU,
             MC, NL, PT, SE, BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN,
             TD, TG
                                           US 1993-148581
                                                            19931108 <--
                       Α
                           19980804
     US 5789210
     CA 2176038
                                           CA 1994-2176038
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                       A1
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    AU 695930
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                                                            19941108 <--
    EP 728192
                      A1
                          19960828
        R: AT, BE, DE, DK, ES, FR, GB, GR, IE, IT, NL, SE
    BR 9408010 A 19961217
                                           BR 1994-8010
                                                            19941108 <--
     CN 1141057
                      Α
                            19970122
                                           CN 1994-194767
                                                            19941108 <--
                     В
                         20031126
    CN 1128873
                      T2
                          19970603
                                           JP 1994-513948
                                                            19941108 <--
     JP 09505469
                                                            19941108 <--
     PL 176399
                      B1 19990531
                                           PL 1994-314297
                    A 19960704
                                                            19960507 <--
     FI 9601926
                                           FI 1996-1926
PRAI US 1993-148581 A
                           19931108 <--
                     A 19931108
W 19941108
                                     <--
    US 1993-148541
    WO 1994-US12861 W
                            19941108 <--
    Described are recombinant yeasts containing genes encoding
AB
     xylose reductase, xylitol
     dehydrogenase and xylulokinase, and DNA mols.,
     vectors and methods useful for producing such yeasts. The
     recombinant yeasts effectively ferment xylose to EtOH, and
     preferred yeasts are capable of simultaneously fermenting
     glucose and xylose to EtOH, thereby taking full advantage of these 2 sugar
     sources as they are found in agricultural biomass.
     recombinant yeast ethanol fermn glucose xylose
ST
     Deoxyribonucleic acid sequences
IT
        (for xylulokinase gene of Saccharomyces cerevisiae)
     Protein sequences
IT
        (for xylulokinase of Saccharomyces cerevisiae)
IT
     Fermentation
       Saccharomyces cerevisiae
        (recombinant yeasts for effective fermentation of glucose and
        xylose)
IT
     Gene, microbial
     RL: PRP (Properties)
        (xylulokinase; sequence of xylulokinase
        gene of Saccharomyces cerevisiae)
ΙT
     167078-89-9
     RL: PRP (Properties)
        (amino acid sequence; recombinant yeasts for effective fermentation
        of glucose and xylose)
IT
     167974-35-8
     RL: PRP (Properties)
        (nucleotide sequence; recombinant yeasts for effective fermentation
        of glucose and xylose)
     9028-16-4, Xylitol dehydrogenase
IT
     9030-58-4, Xylulokinase 99775-25-4,
     Xylose reductase
     RL: CAT (Catalyst use); USES (Uses)
        (recombinant yeasts containing cloned enzyme genes for
```

effective fermentation of glucose and xylose) 64-17-5P, Ethanol, preparation IT RL: BMF (Bioindustrial manufacture); BIOL (Biological study); PREP (Preparation) (recombinant yeasts for effective fermentation of glucose and 58-86-6, Xylose, biological studies 50-99-7, Glucose, biological studies RL: BPR (Biological process); BSU (Biological study, unclassified); RCT (Reactant); BIOL (Biological study); PROC (Process); RACT (Reactant or (recombinant yeasts for effective fermentation of glucose and xylose) ANSWER 12 OF 33 HCAPLUS COPYRIGHT 2004 ACS on STN 1994:650857 HCAPLUS AN DN 121:250857 Entered STN: 26 Nov 1994 ED Utilization of xylose with recombinant Saccharomyces cerevisiae harboring TIgenes for xylose metabolism from Pichia stipitis Meinander, Nina; Hallborn, Johan; Keranen, Sirkka; Ojamo, Heikki; AU Penttila, Merja; Walfridsson, Mats; Hahn-Haegerdal, Barbel Chemical center, University Lund, Lund, S-22100, Swed. CS Progress in Biotechnology (1994), 9(ECB6: PROCEEDINGS OF THE 6TH SO EUROPEAN CONGRESS ON BIOTECHNOLOGY, 1993, PT. 2), 1143-6 CODEN: PBITE3; ISSN: 0921-0423 DTJournal English LA 10-2 (Microbial, Algal, and Fungal Biochemistry) CC Section cross-reference(s): 3 Normally, S. cerevisiae lacks the enzymes necessary to convert xylose into ΔB xylulose and is thereby unable to utilize xylose in its metabolism An S. cerevisiae strain expressing both the xylose reductase (converting xylose to xylitol) and xylitol dehydrogenase (converting xylitol to xylulose) genes of P. stipitis was constructed. This strain was able to grow on and ferment xylose. xylose metab Saccharomyces recombinant STMolecular cloning TT Pichia stipitis Saccharomyces cerevisiae (xylose utilization by recombinant Saccharomyces cerevisiae harboring genes for xylose metabolism from Pichia stipitis) TΤ Gene, microbial RL: BAC (Biological activity or effector, except adverse); BSU (Biological study, unclassified); BIOL (Biological study) (xylose utilization by recombinant Saccharomyces cerevisiae harboring genes for xylose metabolism from Pichia stipitis) IT9028-16-4, Xylitol dehydrogenase 95829-40-6, Xylose reductase RL: BAC (Biological activity or effector, except adverse); BSU (Biological study, unclassified); BIOL (Biological study) (xylose utilization by recombinant Saccharomyces cerevisiae harboring genes for xylose metabolism from Pichia stipitis) 58-86-6, Xylose, biological studies ΙT RL: BPR (Biological process); BSU (Biological study, unclassified); BIOL (Biological study); PROC (Process) (xylose utilization by recombinant Saccharomyces cerevisiae harboring genes for xylose metabolism from Pichia stipitis)

L49 ANSWER 13 OF 33 HCAPLUS COPYRIGHT 2004 ACS on STN

1994:650854 HCAPLUS AN

121:250854 DN

Entered STN: 26 Nov 1994 ED

Bioconversion of xylose to xylitol with in situ generation of NAD(P)H in TI

```
recombinant Saccharomyces cerevisiae
     Carlsen, Helle N.; Hallborn, Johan; Gorwa, Marie-Francoise;
AU
     Hahn-Haegerdal, Baerbel
     Chemical Center, University Lund, Lund, S-221 00, Swed.
CS
     Progress in Biotechnology (1994), 9(ECB6: PROCEEDINGS OF THE 6TH
SO
     EUROPEAN CONGRESS ON BIOTECHNOLOGY, 1993, PT. 1), 313-16
     CODEN: PBITE3; ISSN: 0921-0423
DT
     Journal
     English
LA
     10-2 (Microbial, Algal, and Fungal Biochemistry)
CC
     Section cross-reference(s): 3
     The xylose reductase gene of Pichia stipitis
AΒ
     was cloned into S. cerevisiae. The recombinant S. cerevisiae was thus
     able to convert xylose to xylitol. The cofactor NAD(P)H, used for xylose
     reduction, could be generated in situ through the oxidation of ethanol,
acetate,
     or glucose.
     xylose metab Saccharomyces recombinant
ST
IT
     Molecular cloning
       Saccharomyces cerevisiae
        (bioconversion of xylose to xylitol with in situ generation of NAD(P)H
        in recombinant Saccharomyces cerevisiae)
     58-86-6, Xylose, biological studies
IT
     RL: BPR (Biological process); BSU (Biological study, unclassified); BIOL
     (Biological study); PROC (Process)
        (bioconversion of xylose to xylitol with in situ generation of NAD(P)H
        in recombinant Saccharomyces cerevisiae)
                     58-68-4, NADH
     53-57-6, NADPH
IT
     RL: BPR (Biological process); BSU (Biological study, unclassified); MFM
     (Metabolic formation); BIOL (Biological study); FORM (Formation,
     nonpreparative); PROC (Process)
        (bioconversion of xylose to xylitol with in situ generation of NAD(P)H
        in recombinant Saccharomyces cerevisiae)
     87-99-0, Xylitol
IT
     RL: BSU (Biological study, unclassified); MFM (Metabolic formation); BIOL
     (Biological study); FORM (Formation, nonpreparative)
        (bioconversion of xylose to xylitol with in situ generation of NAD(P)H
        in recombinant Saccharomyces cerevisiae)
     ANSWER 14 OF 33 HCAPLUS COPYRIGHT 2004 ACS on STN
L49
     1994:532375 HCAPLUS
AN
DN
     121:132375
     Entered STN: 17 Sep 1994
ED
     Manufacture of xylitol from carbon sources other than xylose or xylulose
TI
     using yeasts expressing foreign genes
     Harkki, Anu Marjukka; Myasnikov, Andrey Novomirovich; Apajalahti, Juha
IN
     Heikki Antero; Pastinen, Ossi Antero
     Xyrofin Oy, Finland
PΑ
SO
     PCT Int. Appl., 90 pp.
     CODEN: PIXXD2
DT
     Patent
     English
LΑ
     ICM C12P007-18
IC
     ICS C12N015-52
     16-2 (Fermentation and Bioindustrial Chemistry)
CC
     Section cross-reference(s): 10, 17
FAN.CNT 3
                                           APPLICATION NO. DATE
     PATENT NO.
                      KIND DATE
                           -----
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                                          WO 1993-FI450 19931105 <--
     WO 9410325
                A1 19940511
PΙ
         W: AT, AU, BB, BG, BR, BY, CA, CH, CZ, DE, DK, ES, FI, GB, HU, JP,
             KP, KR, KZ, LK, LU, LV, MG, MN, MW, NL, NO, NZ, PL, PT, RO, RU,
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SD, SE, SK, UA, US, UZ, VN

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RW: AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE,
            BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG
                                          AU 1994-54215
                                                           19931105 <--
    AU 9454215
                      A1
                           19940524
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                           19950920
                                          EP 1993-924615
    EP 672161
                      A1
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                      B1
    EP 672161
        R: AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LI, LU, NL, PT, SE
                                          HU 1995-1288
                                                           19931105 <--
                      A2
                           19960328
    HU 72187
                      В
                           20010129
    HU 219016
                                          JP 1994-510748
                                                           19931105 <--
    JP 08505522
                      T2 19960618
                      B2 20030804
    JP 3433295
                                                           19931105 <--
                           19990831
                                          BR 1993-7391
    BR 9307391
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                                                           19931105 <--
    AT 184917
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                          19991015
                                          AT 1993-924615
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                                          RU 1995-113172
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    ES 2139024
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                                          ES 1993-924615
                                                           19931105 <--
                     B1 20000229
                                          PL 1993-308742
    PL 178040
                                                           19931105 <--
    FI 9502148
                     A 19950704
                                          FI 1995-2148
                                                           19950504 <--
                     A 19950705
                                          NO 1995-1747
                                                           19950504 <--
    NO 9501747
PRAI US 1992-973325
                           19921105 <--
                      Α
                           19930824
    US 1993-110672
                                    <--
    WO 1993-FI450
                      W
                           19931105
                                     <--
    Novel methods for the fermentation of xylitol from sugars using a yeast
AΒ
    with modified D-arabitol metabolism are described. Yeasts
    synthesizing D-arabitol were modified by the introduction of expression
    cassettes for D-arabitol dehydrogenase and xylitol
    dehydrogenase and by inactivation of the host genes for
    transketolase and D-xylulokinase and increasing levels of
    expression of genes for enzymes of the oxidative branch of the
    pentose phosphate pathway. Plasmid pSRT(AX) -9 carrying
    expression cassettes for D-arabitol dehydrogenase and xylitol
    dehydrogenase was introduced into Zygosaccharomyces rouxii.
    Transformants showing significant activities of the 2 enzymes were used to
    manufacture xylitol with yields of 7.7 g/L obtained after 48 h. Yields
    depended upon nutritional conditions with higher yields coming from a
    medium enriched with yeast extract
    xylitol fermn transgenic yeast; arabitol metab xylitol fermn
ST
    yeast; pentose phosphate pathway xylitol fermn yeast
IT
    Gene, microbial
    RL: BIOL (Biological study)
        (XYL2, for xylitol dehydrogenase of Pichia
        stipitis, cloning and expression of, xylitol manufacture in fungi with
        altered arabitol metabolism in relation to)
    Gene, microbial
IT
    RL: BIOL (Biological study)
        (for transketolase of Saccharomyces cerevisiae, cloning and
        inactivation of, xylitol manufacture in relation to)
    Gene, microbial
TТ
    RL: BIOL (Biological study)
        (for D-arabitol dehydrogenase of Klebsiella terrigena, cloning and
        expression of, xylitol manufacture in fungi with altered arabitol
metabolism in
        relation to)
     Pentose phosphate pathway
TΤ
        (oxidative branch of, increasing activity of, in xylitol manufacture with
        fungi with altered arabitol metabolism)
IT
    Plasmid and Episome
        (pCPU(AX), genes for D-arabitol dehydrogenase and
        xylitol dehydrogenase on, expression in Candida
       polymorpha of, altered arabitol metabolism and xylitol manufacture in
relation
IT
    Plasmid and Episome
        (pSRT(AX)-9, genes for D-arabitol dehydrogenase and
```

xylitol dehydrogenase on, expression in

```
Zygosaccharomyces rouxii of, altered arabitol metabolism and xylitol
manufacture
        in relation to)
    Plasmid and Episome
TT
        (pSRT(ZG), genes for D-glucose-6-phosphate dehydrogenase and
        6-phospho-D-gluconate dehydrogenase on, expression in Zygosaccharomyces
        rouxii of)
    Plasmid and Episome
IT
        (pTC(AX), integrating dominant selection vector for Torulopsis candida,
        genes for D-arabitol dehydrogenase and xylitol
        dehydrogenase on, altered arabitol metabolism and xylitol manufacture in
        relation to)
TT
    Candida diddensii
    Dendryphiella salina
    Fungi
    Pichia farinosa
       Saccharomyces rouxii
    Schizophyllum commune
    Torulaspora hansenii
    Torulopsis candida
       Yeast
        (xylitol manufacture with, with altered arabitol metabolism)
    Fermentation
IT
        (xylitol, with transgenic fungi with altered arabitol metabolism)
IT
     Klebsiella terrigena
        (D-arabitol dehydrogenase gene of, cloning and expression of,
        xylitol manufacture in fungi with altered arabitol metabolism in relation
to)
     Gene, microbial
TT
     RL: BIOL (Biological study)
        (URA3, of Candida polymorpha, cloning of, transformation vectors for C.
        polymorpha and xylitol manufacture in relation to)
     Gene, microbial
IT
     RL: BIOL (Biological study)
        (gnd, of Escherichia coli, cloning of, alteration of pentose phosphate
        pathway in xylitol manufacture in relation to)
     Gene, microbial
IT
    RL: BIOL (Biological study)
        (zwf, of Saccharomyces cerevisiae, cloning of, alteration of pentose
        phosphate pathway in xylitol manufacture in relation to)
IT
     87-99-0, Xylitol
     RL: BIOL (Biological study)
        (fermentation of, with transgenic yeast, modified anabitol metabolism
        and pentose phosphate pathway in)
     9001-40-5, D-Glucose-6-phosphate dehydrogenase
                                                       9024-20-8,
ΙT
                                          9026-40-8, D-Ribulokinase
     D-Ribulose-5-phosphate-3-epimerase
     9073-95-4, 6-Phospho-D-gluconate dehydrogenase
     RL: BIOL (Biological study)
        (gene for, expression of, in fungal hosts with altered
        arabitol metabolism for manufacture of xylitol)
     9028-16-4, Xylitol dehydrogenase
                                        9028-18-6,
IT
     D-Arabitol dehydrogenase
     RL: BIOL (Biological study)
        (gene for, expression of, in transgenic yeast,
        alteration of arabitol metabolism in xylitol manufacture in relation to)
     9014-48-6, Transketolase 9030-58-4, D-Xylulokinase
IT
     RL: BIOL (Biological study)
        (gene for, inactivation of, in fungal hosts with altered
        arabitol metabolism for manufacture of xylitol)
     2152-56-9, Arabitol
IT
     RL: BPR (Biological process); BSU (Biological study, unclassified); BIOL
     (Biological study); PROC (Process)
```

(metabolism of, alteration of, in manufacture of xylitol with transgenic

yeasts)

L49

AN

1993:226769 HCAPLUS

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ANSWER 15 OF 33 HCAPLUS COPYRIGHT 2004 ACS on STN
     1993:510092 HCAPLUS
ΑN
DN
     119:110092
ED
     Entered STN: 18 Sep 1993
     Cloning and improveing the expression of Pichia stipitis xylose
TT
     reductase gene in Saccharomyces cerevisiae
     Chen, Zhengdao; Ho, Nancy W. Y.
ΑU
     Lab. Renewable Resourc. Eng., Purdue Univ., West Lafayette, IN,
CS
     47907-1295, USA
     Applied Biochemistry and Biotechnology (1993), 39-40, 135-47
SO
     CODEN: ABIBDL; ISSN: 0273-2289
DТ
     Journal
     English
LA
     3-2 (Biochemical Genetics)
CC
     Section cross-reference(s): 16
     The intact Pichia stipitis xylose reductase
AB
     gene (XR) has been cloned and expressed in Saccharomyces
     cerevisiae. The possible further improvement of the expression of the
     Pichia gene in the new host was studied. To improve the
     expression of the XR gene in yeast (Saccharomyces
     cerevisiae), its 5'-noncoding sequence containing the genetic elements for
     transcription and translation was systematically replaced by that from the
     yeast genes. It was found that the Pichia genetic
     signal for transcription of XR is more effective than the yeast
     TRP5 promoter, but is about half as effective as the yeast
     strong promoter of the alc. dehydrogenase gene (ADC1). However,
     the nucleotide sequence immediately adjacent to the initiation codon of
     XR, which controls the translation of the gene product, seemed
     to be five times less effective than the corresponding sequence of the
     ADC1 gene. By totally replacing its 5'-noncoding sequence with
     that of the yeast ADC1 gene, the expression of XR in
     yeast was nearly ten times higher. Furthermore, the cloned Pichia
     XR described in this article contains very little of its 3'-noncoding sequence. In order to study whether the 3'-noncoding sequence is
     important to its expression in S. cerevisiae, the intact 3'-noncoding
     sequences of the yeast xylulokinase gene was
     spliced to the 3' end of the PADC1-XR structural gene. This
     latter modification has resulted in a 2-fold further increase in the
     expression of the Pichia XR in yeast.
     Pichia xylose reductase gene cloning
ST
     Saccharomyces
     Saccharomyces cerevisiae
IT
         (cloning and expression in, of xylose reductase
        gene of Pichia stipitis)
     Gene, microbial
IT
     RL: BIOL (Biological study)
         (for xylose reductase, of Pichia stipitis, cloning
        and expression in Saccharomyces cerevisiae of)
IT
     Molecular cloning
         (of xylose reductase gene, of Pichia
        stipitis, in Saccharomyces cerevisiae)
IT
     Pichia stipitis
         (xylose reductase gene of, cloning and
        expression of, in Saccharomyces cerevisiae)
     95829-40-6, Xylose reductase
ΙT
     RL: BIOL (Biological study)
         (gene for, of Pichia stipitis, cloning and expression of, in
        Saccharomyces cerevisiae)
     ANSWER 16 OF 33 HCAPLUS COPYRIGHT 2004 ACS on STN
```

```
DN
     118:226769
ED
     Entered STN: 12 Jun 1993
     Isolation of xylose reductase gene of Pichia
TI
     stipitis and its expression in Saccharomyces cerevisiae
     Takuma, Shinya; Nakashima, Noriyuki; Tantirungkij, Manee; Kinoshita,
AU
     Shinichi; Okada, Hirosuke; Seki, Tatsuji; Yoshida, Toshiomi
     Fac. Eng., Osaka Univ., Suita, 565, Japan
CS
     Applied Biochemistry and Biotechnology (1991), 28-29, 327-40
SO
     CODEN: ABIBDL; ISSN: 0273-2289
דת
     Journal
LΑ
     English
     3-2 (Biochemical Genetics)
CC
     Section cross-reference(s): 7, 10
AB
     A NADPH/NADH-dependent xylose reductase gene
     was isolated from the xylose-assimilating yeast, Pichia stipits.
     DNA sequence anal. showed that the gene consists of 951
          The gene introduced in Saccharomyces cerevisiae was
     transcribed to mRNA, and a considerable amount of enzyme activity was observed
     constitutively, whereas transcription and translation in P. stipitis were
     inducible. S. cerevisiae carrying the xylose reductase
     gene could not, however, grow on xylose medium, and could not
     produce ethanol from xylose. Since xylose uptake and accumulation of
     xylitol by S. cerevisiae were observed, the conversion of xylitol to xylulose
     seemed to be limited.
ST
     Pichia xylose reductase gene cloning
     sequence; Saccharomyces cloning xylose reductase
     gene Pichia
     Saccharomyces cerevisiae
IT
        (cloning and expression in, of xylose reductase
        gene, of Pichia stipitis)
IT
     Gene, microbial
     RL: BIOL (Biological study)
        (for xylose reductase, of Pichia stipitis, cloning
        and expression and sequencing of)
     Deoxyribonucleic acid sequences
IT
        (of xylose reductase gene, of Pichia
        stipitis)
IT
     Molecular cloning
        (of xylose reductase gene, of Pichia
        stipitis, for expression in yeast)
IT
     Protein sequences
        (of xylose reductase, of Pichia stipitis)
TΤ
     Pichia stipitis
        (xylose reductase gene of, sequence and
        expression in yeast of)
IT
     138263-97-5
     RL: BAC (Biological activity or effector, except adverse); BSU (Biological
     study, unclassified); PRP (Properties); BIOL (Biological study)
        (amino acid sequence of, complete)
TT
     95829-40-6, Xylose reductase
     RL: BIOL (Biological study)
        (gene for, of Pichia stipitis, cloning and expression and
        sequencing of)
ΙT
     147651-00-1
     RL: PRP (Properties); BIOL (Biological study)
        (nucleotide sequence of)
    ANSWER 17 OF 33 HCAPLUS COPYRIGHT 2004 ACS on STN
L49
AN
     1993:190040 HCAPLUS
DN
     118:190040
ED
     Entered STN:
                   14 May 1993
     Secretion of a xylanase from Cryptococcus albidus by Saccharomyces
TI
```

cerevisiae and Pichia stipitis

```
Morosoli, Rolf; Zalce, Eugenia; Moreau, Alain; Durand, Serge
ΑU
     Cent. Rech. Microbiol. Appl., Inst. Armand-Frappier, Ville de Laval, QC,
CS
     H7N 4Z3, Can.
     Progress in Biotechnology (1992), 7(Xylans Xylanases), 247-58
SO
     CODEN: PBITE3; ISSN: 0921-0423
DT
LΑ
     English
     16-4 (Fermentation and Bioindustrial Chemistry)
CC
     Section cross-reference(s): 3
     The xylanase gene of Cryptococcus albidus and its cDNA were each
AΒ
     inserted in the vector pVT100 and in the vector pJHS to transform
     Saccharomyces cerevisiae and Pichia stipitis, resp. The xylanase
     gene was under the control of its own promoter for expts. in S.
     cerevisiae, while in P. stipitis it was under the control of the
     xylose reductase promoter of the same strain.
     Yeasts transformed with plasmids containing the cDNA of the
     structural xylanase gene produced active extracellular xylanase.
     The enzyme secreted by S. cerevisiae had an apparent mol. mass of 48-kDa,
     which corresponds to that of the native xylanase produced by C. albidus.
     The enzyme synthesized by P. stipitis, however, had an apparent mol. mass
     of 50-kDa, probably reflecting a different protein glycosylation level by
     this strain. With plasmids bearing the genomic xylanase
     gene, transcription occurred, but the seven introns interrupting
     the xylanase gene were neither spliced out by S. cerevisiae nor
     by P. stipitis and no enzyme was produced. Expression of the xylanase
     gene by P. stipitis, resulted in a yeast able to grow on
     xylan as carbon source, directly fermenting it to ethanol under anaerobic
     conditions.
     Cryptococcus xylanase gene cloning Saccharomyces Pichia
st
IT
     Pichia stipitis
       Saccharomyces cerevisiae
        (cloning and expression in, of xylanase gene of Cryptococcus
        albidus)
IT
     Gene, microbial
     RL: BIOL (Biological study)
         (for xylanase, of Streptococcus albidus, cloning and expression in
        Saccharomyces cerevisiae and Pichia stipitis of)
IT
     Molecular cloning
        (of xylanase gene, of Cryptococcus albidus, in Saccharomyces
        cerevisiae and Pichia stipitis)
     Cryptococcus albidus
IT
         (xylanase gene of, cloning and expression of, in
        Saccharomyces cerevisiae and Pichia stipitis)
IT
     37278-89-0, Xylanase
     RL: BIOL (Biological study)
         (gene for, of Cryptococcus albidus, cloning and expression in
        Saccharomyces cerevisiae and Pichia stipitis of)
     ANSWER 18 OF 33 HCAPLUS COPYRIGHT 2004 ACS on STN
     1993:123016 HCAPLUS
AN
DN
     118:123016
     Entered STN: 30 Mar 1993
ED
     Xylitol production by recombinant Saccharomyces cerevisiae
TI
     Hallborn, Johan; Walfridsson, Mats; Airaksinen, Ulla; Ojamo, Heikki;
ΑU
     Hahn-Hagerdal, Barbel; Penttila, Merja; Keranen, Sirkka
     VTT Biotech. Lab., Espoo, SF-02151, Finland
CS
     Bio/Technology (1991), 9(11), 1090-5
CODEN: BTCHDA; ISSN: 0733-222X
SO
DT
     Journal
     English
LA
     16-2 (Fermentation and Bioindustrial Chemistry)
CC
```

Efficient conversion of xylose to xylitol was obtained by transforming

Section cross-reference(s): 3, 7, 10

AΒ

ST

IT

IT

TТ

TT

TΤ

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TT

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IT

ΙT

IT

AN DN

ED

TI

IN PA

SO

CODEN: PIXXD2

Saccharomyces cerevisiae with the gene encoding the xylose reductase (XR) of Pichia stipitis CBS 6054. Comparison of the chromosomal and cDNA copies of the XYL1 gene showed that the genomic XYL1 contains no introns, and an XR monomer of 318 amino acids (35,985 Da) is encoded by an open reading frame The amino acid sequence of the P. stipitis XR is similar to of 954 bp. several aldose reductases, suggesting that P. stipitis XR is part of the aldoketo reductase superfamily. S. cerevisiae transformed with the XYL1 gene gave over 95% conversion of xylose into xylitol, a yield not obtainable with natural xylose utilizing yeasts. xylitol fermn xylose recombinant Saccharomyces; xylose reductase gene XYL1 sequence Pichia Saccharomyces cerevisiae (cloning and expression in, of xylose reductase gene of Pichia stipitis) Molecular cloning (of xylose reductase gene, of Pichia stipitis, in Saccharomyces cerevisiae) Protein sequences (of xylose reductase, of Pichia stipitis) Fermentation (xylitol, from xylose by recombinant Saccharomyces cerevisiae) Pichia stipitis (xylose reductase of, gene for, cloning and sequence of) Deoxyribonucleic acid sequences (complementary, for xylose reductase of Pichia stipitis) Gene, microbial RL: BIOL (Biological study) (XYL1, for xylose reductase, of Pichia stipitis, cloning and sequence of) 138263-97-5, Xylose reductase (Pichia stipitis reduced) RL: PRP (Properties); BIOL (Biological study) (amino acid sequence of, complete) 95829-40-6, Xylose reductase RL: BIOL (Biological study) (gene for, of Pichia stipitis, cloning and sequence of) 87-99-0P, Xylitol RL: BMF (Bioindustrial manufacture); BIOL (Biological study); PREP (Preparation) (manufacture of, from xylose by recombinant Saccharomyces cerevisiae) 146409-21-4 RL: PRP (Properties) (nucleotide sequence of) 146409-22-5 RL: PRP (Properties) (nucleotide sequence of, complete) 58-86-6, D-Xylose, uses RL: USES (Uses) (xylitol manufacture from, by recombinant Saccharomyces cerevisiae) ANSWER 19 OF 33 HCAPLUS COPYRIGHT 2004 ACS on STN 1993:100558 HCAPLUS 118:100558 Entered STN: 19 Mar 1993 Xylitol manufacture with yeast mutants Apajalahti, Juha; Leisola, Matti Xyrofin Oy, Finland PCT Int. Appl., 33 pp.

```
DT
     Patent
LA
     English
IC
     ICM C12P007-18
     16-5 (Fermentation and Bioindustrial Chemistry)
     Section cross-reference(s): 10
FAN.CNT 1
     PATENT NO.
                      KIND DATE
                                            APPLICATION NO.
                                                              DATE
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PΙ
     WO 9301299
                      A1
                             19930121
                                            WO 1992-FI203
                                                              19920630 <--
         W: CA, DE, FI, GB, JP, NL, NO
         RW: AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LU, MC, NL, SE
                             19930121
                                            CA 1992-2112374 19920630 <--
     CA 2112374
                       AA
     CA 2112374
                       С
                             20021029
                             19940706
                                            EP 1992-912764
                                                              19920630 <--
     EP 604429
                       Α1
                             19991020
     EP 604429
                       В1
         R: AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LI, LU, MC, NL, SE
                                            JP 1993-501989
                                                              19920630 <--
     JP 07500492
                       T2
                           19950119
                       B2
                             20021007
     JP 3331343
                                            AT 1992-912764
                                                              19920630 <--
     AT 185841
                       E
                             19991115
                       Т3
                                            ES 1992-912764
                                                              19920630 <--
     ES 2140411
                             20000301
     FR 2678637
                       A1
                             19930108
                                            FR 1992-8109
                                                              19920701 <--
     FR 2678637
                       B1
                             19960209
                                            US 1994-194624
                                                              19940207 <--
    US 6271007
                       В1
                             20010807
PRAI FI 1991-3197
                       Α
                             19910701
                                      <--
     US 1992-905870
                       В1
                             19920630
                                       <--
                             19920630 <--
     WO 1992-FI203
                       W
     Xylitol is manufactured with yeast mutants defective in xylose metabolism
AB
     Kluyveromyces marxianus was mutagenized with acriflavine or ethylmethane
     sulfonate the treated with benomyl. Strains unable to metabolize xylose effectively were selected on xylose- and nystatin-containing medium. Xylitol
     production rates of 2.8 g/L/h were achieved with one mutant.
ST
     xylitol manuf yeast mutant
IT
     Candida
     Candida utilis
     Hansenula
     Kluyveromyces
     Kluyveromyces marxianus
     Kluyveromyces marxianus bulgaricus
     Kluyveromyces marxianus lactis
     Kluyveromyces marxianus marxianus
     Pichia
        (xylose metabolism mutants of, for xylitol manufacture)
IT
     Ribozymes
     RL: BIOL (Biological study)
        (yeast transformant containing, directed against xylose metabolism,
        xylitol manufacture with)
     Ribonucleic acids
IT
     RL: BIOL (Biological study)
        (antisense, yeast transformant containing, directed against
        xylose metabolism, xylitol manufacture with)
     9028-16-4, Xylitol dehydrogenase
TΤ
     9030-58-4
     RL: BIOL (Biological study)
        (inactivating mutation in gene for, yeast for
        xylitol manufacture containing)
     87-99-0P, Xylitol
IT
     RL: BMF (Bioindustrial manufacture); BIOL (Biological study); PREP
     (Preparation)
        (manufacture of, yeast mutants for)
IT
     58-86-6, Xylose, uses
     RL: BIOL (Biological study)
        (yeast deficient in metabolism of, in preparation of, xylitol manufacture
        in relation to)
```

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ANSWER 20 OF 33 HCAPLUS COPYRIGHT 2004 ACS on STN
AN
     1993:96990 HCAPLUS
DN
     118:96990
ED
     Entered STN: 19 Mar 1993
     Cloning and expression in Saccharomyces cerevisiae of the
TI
     NAD(P)H-dependent xylose reductase-encoding
     gene (XYL1) from the xylose-assimilating yeast Pichia
     stipitis
     Amore, Rene; Koetter, Peter; Kuester, Christina; Ciriacy, Michael;
ΑU
     Hollenberg, Cornelis P.
     Inst. Mikrobiol., Heinrich-Heine-Univ., Duesseldorf, 4000, Germany
CS
     Gene (1991), 109(1), 89-97
SO
     CODEN: GENED6; ISSN: 0378-1119
DT
     Journal
     English
LA
CC
     7-5 (Enzymes)
     Section cross-reference(s): 3
     The XYL1 gene of the yeast P. stipitis was isolated
AΒ
     from a genomic library using a specific cDNA probe, and its nucleotide
     (nt) sequence was determined In the 5' noncoding region of the P. stipitis
     XYL1 gene, a TATAAA element (known to be necessary for
     transcription initiation in most yeast genes) is found
     at nt -81, and two CCAAT recognition motifs (often referred to as the
     CCAAT box) are present at nt -146 and -106. The XYL1 encodes a
     polypeptide of 35,927 Da that constitutes a NADH/NADPH-dependent
     xylose reductase (XR). The enzyme is part of the
     xylose-xylulose pathway that is absent or only weakly expressed in S.
     cerevisiae. Extensive homol. is found to the N terminus of the XR of
     Pachysolen tannophilus and Candida shehatae. None of the known cofactor
     binding domains found in many NAD-dependent dehydrogenases are present in
     the protein. Transformants of S. cerevisiae containing XYL1 of P. stipitis
     synthesize an active XR. Fusion of XYL1 with the prokaryotic tac promoter
     leads to a gene that can be expressed in S. cerevisiae and
     Escherichia coli.
     xylose reductase gene Pichia sequence
ST
     cloning
IT
     Pichia stipitis
        (NADH/NADPH-dependent xylose reductase gene
        of, cloning and sequence and expression of)
IT
     Escherichia coli
       Saccharomyces cerevisiae
        (cloning in, of XYL1 gene of Pichia stipitis)
IT
     Deoxyribonucleic acid sequences
        (for NADH/NADPH-dependent xylose reductase
        gene, of Pichia stipitis)
IT
     Protein sequences
        (for NADH/NADPH-dependent xylose reductase, of
        Pichia stipitis)
IT
     Molecular cloning
        (of XYL1 gene of Pichia stipitis, in Escherichia coli and
        Saccharomyces cerevisiae)
IT
     Gene, microbial
     RL: BIOL (Biological study)
         (XYL1, of Pichia stipitis, cloning and sequence and expression of)
IT
     138263-97-5, NADH/NADPH-dependent xylose
     reductase (Pichia stipitis λgt11 clone reduced)
     RL: PRP (Properties); BIOL (Biological study)
         (amino acid sequence of, complete)
     95829-40-6
IT
     RL: BIOL (Biological study)
         (gene for, of Pichia stipitis, cloning and sequence and
        expression of)
```

138575-99-2, DNA (Pichia stipitis λgt11 clone XYL1 IT gene and flanking region) RL: PRP (Properties); BIOL (Biological study) (nucleotide sequence of) IT 138575-98-1 RL: PRP (Properties); BIOL (Biological study) (nucleotide sequence of, complete) ANSWER 21 OF 33 HCAPLUS COPYRIGHT 2004 ACS on STN L49 AN 1992:52957 HCAPLUS DM116:52957 EDEntered STN: 21 Feb 1992 Cloning of yeast xylose reductase and TIxylitol dehydrogenase genes and their use Strasser, Alexander W. M.; Hollenberg, Cornelis P.; Von Ciriacy-Wantrup, IN Michael; Koetter, Peter; Amore, Rene; Piontek, Michael; Hagedorn, Jutta Rhein Biotech Gesellschaft fuer neue Biotechnologische Prozesse und PA Produkte m.b.H., Germany Ger. Offen., 51 pp. SO CODEN: GWXXBX Patent DTGerman LA ICM C12N001-19 IC ICS C12N015-63; C12P019-34; C07H021-04; C07K015-04 CC 3-4 (Biochemical Genetics) FAN.CNT 1 KIND DATE APPLICATION NO. DATE PATENT NO. ----_____ 19911002 DE 1990-4009676 19900326 <--DE 4009676 A1 ΡI C2 19930909 DE 4009676 A2 19911009 EP 1991-104558 19910322 <--EP 450430 EP 450430 A3 19920102 EP 450430 B1 19970625 R: AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LI, LU, NL, SE E 19970715 AT 1991-104558 19910322 <--AT 154829 19971016 ES 2104626 Т3 ES 1991-104558 19910322 <--AA 19910927 CA 2039021 CA 1991-2039021 19910325 <--A2 19941213 JP 1991-62160 19910326 <--JP 06339383 B2 20010109 JP 3122153 JP 2000139486 A2 20000523 JP 2001103988 A2 20010417 JP 2000-589 19910326 <--JP 2000-276227 19910326 <--B2 20010730 JP 3193917 PRAI DE 1990-4009676 A 19900326 <--JP 1991-62160 A3 19910326 <--The XYL1 gene encoding xylose reductase and AB the XYL2 gene encoding xylitol dehydrogenase of Pichia stipitis are cloned, sequenced, and expressed in other microorganisms. Yeast transformants expressing these genes can be used to prepare EtOH, alc. beverages, or biomass. promoters of these genes can be used to express genes in yeast. A Saccharomyces cerevisiae mutant containing both genes was prepared and used to prepare EtOH in .apprx.80% yield from xylose. Plasmids containing Clostridium thermocellum cellulase gene linked to the promoter of XYL1 or XYL2 were prepared and P. stipitis transformed with them. These transformants produced the enzyme in response to xylose induction. XYL1 XYL2 gene Pichia cloning; xylose streductase gene Pichia; xylitol dehydrogenase gene Pichia; Saccharomyces transformant ethanol manuf xylose

(alc., yeast expressing XYL1 and/or XYL2 genes of

Fermentation

Pichia stipitis for)

IT

```
IT
     Paecilomyces
       Saccharomyces cerevisiae
     Schizosaccharomyces
     Schizosaccharomyces pombe
     Zymomonas
        (expression in, of XYL1 and XYL2 genes of Pichia stipitis)
IT
     Protein sequences
        (of xylitol dehydrogenase of Pichia stipitis,
        complete)
IT
     Protein sequences
        (of xylose reductase of Pichia stipitis, complete)
     Molecular cloning
IT
        (of XYL1 and XYL2 genes of Pichia stipitis, in yeast
     Plasmid and Episome
IT
        (pMPGC1-2, cellulase gene of Clostridium on, expression in
        Pichia stipitis of)
ΙT
     Plasmid and Episome
        (pR2, xylose reductase gene XYL1 of
        Pichia stipitis on, expression in Saccharomyces cerevisiae of)
ΙT
     Plasmid and Episome
        (pXDH, xylitol dehydrogenase gene XYL2
        fragment of Pichia stipitis on)
     Plasmid and Episome
IT
        (pXDH-HIS3, xylitol dehydrogenase gene
        XYL2 of Pichia stipitis on, expression in Schizosaccharomyces pombe of)
IT
     Plasmid and Episome
        (pXR, xylose reductase gene XYL1 of
        Pichia stipitis on, expression in Saccharomyces cerevisiae of)
IT
     Plasmid and Episome
        (pXR-LEU2, xylose reductase gene XYL1 of
        Pichia stipitis on, expression in Schizosaccharomyces pombe of)
IT
     Plasmid and Episome
        (pXRa, xylose reductase gene, XYL1
        fragment of Pichia stipitis on)
IT
     Plasmid and Episome
        (pXRb, xylose reductase gene XYL1
        fragment of Pichia stipitis on)
IT
     Biomass
        (preparation of, yeast expressing XYL1 and/or XYL2 genes
        of Pichia stipitis for)
     Deoxyribonucleic acid sequences
ΙT
        (xylitol dehydrogenase-specifying, of Pichia
        stipitis, complete)
IT
     Candida
     Debaryomyces -
     Hansenula
     Kluyveromyces
     Metschnikowia
     Pachysolen (fungus)
     Pichia
       Saccharomyces
     Schwanniomyces
        (xylose reductase and xylitol
        dehydrogenase genes of, cloning of, cloning of XYL1
        and XYL2 genes of Pichia stipitis in relation to)
IT
     Deoxyribonucleic acid sequences
        (xylose reductase-specifying, of Pichia stipitis,
        complete)
IT
     Pichia stipitis
        (XYL1 and XYL2 genes of, cloning and expression in
        yeast of)
```

IT

Plasmid and Episome

```
(pD1, xylitol dehydrogenase gene XYL2 of
        Pichia stipitis on, expression in Saccharomyces cerevisiae of)
IT
    Plasmid and Episome
        (pD2, xylitol dehydrogenase gene XYL2 of
        Pichia stipitis on, expression in Saccharomyces cerevisiae of)
IT
     Plasmid and Episome
        (pR1, xylose reductase gene XYL1 of
        Pichia stipitis on, expression in Saccharomyces cerevisiae of)
IT
     Plasmid and Episome
        (pRD1, xylose reductase gene XYL1 and
        xylitol dehydrogenase gene XYL2 of Pichia
        stipitis on, expression in Saccharomyces cerevisiae of)
TT
     Genetic element
     RL: BIOL (Biological study)
        (promoter, of XYL1 and XYL2 genes of Pichia stipitis,
        heterologous gene expression in yeast using)
     Gene, microbial
TΤ
     RL: BIOL (Biological study)
        (XYL1, cloning and expression of, of Pichia stipitis, in yeast
     Gene, microbial
IT
     RL: BIOL (Biological study)
        (XYL2, cloning and expression of, of Pichia stipitis, in yeast
IT
     136511-83-6 138263-97-5
     RL: BIOL (Biological study)
        (amino acid sequence of and expression in Saccharomyces of gene
     136510-54-8, Deoxyribonucleic acid (Pichia stipitis clone pD1 gene
IT
             136510-55-9, Deoxyribonucleic acid (Pichia stipitis clone pD1
     gene XYL2 plus 5'- and 3'-flanking region fragment)
                                                           138575-98-1,
     Deoxyribonucleic acid (Pichia stipitis clone pR1 gene XYL1)
     138575-99-2, Deoxyribonucleic acid (Pichia stipitis clone pR1 gene
     XYL1 plus 5'- and 3'-flanking region fragment)
     RL: BIOL (Biological study)
        (cloning and expression in Saccharomyces and nucleotide sequence of)
     138575-97-0, Deoxyribonucleic acid (Pichia stipitis clone pD1 gene
IT
     XYL2 promoter region-containing fragment)
                                                 138576-00-8, Deoxyribonucleic
     acid (Pichia stipitis clone pR1 gene XYL1 promoter
     region-containing fragment)
     RL: PRP (Properties)
        (gene expression in yeast using and nucleotide
        sequence of)
TΤ
     9028-16-4, Xylitol dehydrogenase
     95829-40-6, Xylose reductase
     RL: BIOL (Biological study)
        (gene for, of Pichia stipitis, cloning and expression in
        yeast of)
ΙT
     64-17-5P, Ethanol, preparation
     RL: PREP (Preparation)
        (manufacture of, yeast transformants expressing XYL1 and XYL2
        genes of Pichia stipitis for)
     53-57-6P, NADPH
                       53-59-8P, NADP+
IT
     RL: PREP (Preparation)
        (preparation of, from NADPH, xylose reductase of Pichia
        stipitis for)
     551-84-8, Xylulose
ΙŢ
     RL: BIOL (Biological study)
        (yeast mutants growing on, XYL1 and XYL2 genes of
        Pichia stipitis expression in and enzyme manufacture with)
     58-86-6, Xylose, biological studies
IT
     RL: BIOL (Biological study)
```

(yeast transformed with XYL1 and/or XYL2 genes of

Pichia growth on, for biomass preparation)

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ANSWER 22 OF 33 HCAPLUS COPYRIGHT 2004 ACS on STN
    1992:35663 HCAPLUS
AN
DN
    116:35663
ED
    Entered STN: 08 Feb 1992
    Recombinant yeasts containing DNA sequences coding for
TI
    xylose reductase and xylitol
     dehydrogenase
    Hallborn, Johan; Penttila, Merja; Ojamo, Heikki; Walfridsson, Mats;
IN
    Airaksinen, Ulla; Keranen, Sirkka; Hahn-Hagerdal, Barbel
    Valtion Teknillinen Tutkimuskeskus, Finland
PΑ
SO
    PCT Int. Appl., 47 pp.
     CODEN: PIXXD2
     Patent
DT
    English
LΑ
    ICM C12N015-53
ICS C12N009-04
IC
CC
     3-4 (Biochemical Genetics)
FAN.CNT 1
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                     KIND DATE
     PATENT NO.
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     ______
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                           19911017
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                                         FI 1990-1771
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     CA 2090122
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                           19911007
     CA 2090122
                      С
                           20020618
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                     В1
                           19980107
     EP 527758
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                                          JP 1991-506907
     JP 05507843
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                                                           19910408 <--
                                          AT 1991-906996
                      E
                           19980115
     AT 161886
                      Т3
                                          ES 1991-906996
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                           19980501
     ES 2113373
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                           19921006
                                          NO 1992-3880
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                                          US 1994-336198
                                                          19941103 <--
                      Α
                           19990202
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                                          US 1999-184965
                      В1
                                                          19990108 <--
     US 6582944
                           20030624
                      A
                                          FI 1999-2153
                                                          19991006 <--
     FI 9902153
                           19991006
PRAI FI 1990-1771
                           19900406
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                      Α
     US 1990-527775
                      A2
                           19900524
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     WO 1991-FI103
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     US 1992-848694
                      B1
                           19920309
                                     <--
     FI 1992-4461
                      Α
                           19921002
                                     <--
                           19941103 <--
     US 1994-336198
                      A3
     A cDNA for yeast xylose reductase is cloned
AB
     and sequenced. This cDNA is expressed in recombinant yeast,
     optionally along with that for xylitol dehydrogenase.
     These recombinant yeast can be used to prepare xylitol, or ethanol
     (when both genes are expressed), from xylose or xylose-containing
     materials. The xylose reductase cDNA of Pichia
     stipitis was cloned. Saccharomyces cerevisiae transformants expressing
     this cDNA were used to prepare xylitol. S. cerevisiae expressing both
     xylose reductase and xylitol
     dehydrogenase produced EtOH, xylitol, and biomass from spent
     sulfite liquor.
     xylose reductase cDNA Pichia cloning; xylitol ethanol
ST
     manuf recombinant Saccharomyces
IT
     Gene, microbial
     RL: BIOL (Biological study)
        (cDNA, for xylose reductase of Pichia stipitis,
```

cloning and expression in Saccharomyces cerevisiae of) IT Kluyveromyces Pichia Saccharomyces cerevisiae Schizosaccharomyces pombe (expression in, of xylose reductase cDNA of Pichia stipitis) ΙT Molecular cloning (of xylose reductase cDNA of Pichia stipitis, in Saccharomyces cerevisiae) IT Protein sequences (of xylose reductase of Pichia stipitis, complete) ΙT Plasmid and Episome (pJHXDH60, xylitol dehydrogenase cDNA of Pichia stipitis on, expression in Saccharomyces cerevisiae of) IT Plasmid and Episome (pJHXDH70, xylitol dehydrogenase cDNA of Pichia stipitis on, expression in Saccharomyces cerevisiae of) IT Plasmid and Episome (pJHXR22, xylose reductase cDNA of Pichia stipitis on, expression in Saccharomyces cerevisiae of) IT Plasmid and Episome (pMW22, xylitol dehydrogenase cDNA of Pichia stipitis on, expression in Saccharomyces cerevisiae of) IT Plasmid and Episome (pUA103, xylose reductase cDNA of Pichia stipitis on, expression in Saccharomyces cerevisiae of) IT Plasmid and Episome (pUA107, xylose reductase cDNA of Pichia stipitis on, expression in Saccharomyces cerevisiae of) IT Pichia stipitis (xylose reductase cDNA of, cloning and expression in Saccharomyces cerevisiae of) ΙT Deoxyribonucleic acid sequences (xylose reductase-specifying, of Pichia stipitis, complete) 138263-97-5 IT RL: PRP (Properties); BIOL (Biological study) (amino acid sequence of and cloning of cDNA for) 95829-40-6, Xylose reductase IT RL: BIOL (Biological study) (cDNA for, of Pichia stipitis, cloning and expression in Saccharomyces cerevisiae of) ΙT 9028-16-4, Xylitol dehydrogenase RL: BIOL (Biological study) (cDNA for, recombinant yeast expressing xylose reductase cDNA and, ethanol manufacture with) 138263-60-2, Deoxyribonucleic acid (Pichia stipitis clone pUA103 IT gene xrd minus terminator fragment) RL: PRP (Properties); BIOL (Biological study) (cloning and nucleotide sequence of) 87-99-0P, Xylitol IT RL: PREP (Preparation) (manufacture of, from xylose, recombinant yeast xylose reductase for) 64-17-5P, Ethanol, preparation IT RL: PREP (Preparation) (manufacture of, recombinant yeast expressing xylose reductase and xylitol dehydrogenase cDNAs for) IT 58-86-6, Xylose, biological studies

RL: BIOL (Biological study)

(xylitol manufacture from, recombinant yeast expressing cloned xylose reductase cDNA for)

```
ANSWER 23 OF 33 HCAPLUS COPYRIGHT 2004 ACS on STN
L49
AN
     1991:576285 HCAPLUS
DN
     115:176285
ED
     Entered STN: 01 Nov 1991
     Isolation and characterization of the Pichia stipitis xylitol
TI
     dehydrogenase gene, XYL2, and construction of a
     xylose-utilizing Saccharomyces cerevisiae transformant
ΑU
     Koetter, Peter; Amore, Rene; Hollenberg, Cornelis P.; Ciriacy, Michael
CS
     Inst. Mikrobiol., Heinrich-Heine-Univ., Duesseldorf, W-4000, Germany
     Current Genetics (1990), 18(6), 493-500
SO
     CODEN: CUGED5; ISSN: 0172-8083
DT
     Journal
LA
     English
CC
     3-3 (Biochemical Genetics)
     A P. stipitis cDNA library in Agt11 was screened using antisera
AB
     against P. stipitis xylose reductase and
     xylitol dehydrogenase, resp. The resulting cDNA clones
     served as probes for screening a P. stipitis genomic library. The genomic
     XYL2 gene was isolated and the nucleotide sequence of the
     1089-bp structural gene, and of adjacent non-coding regions, was
     determined The XYL2 open-reading frame codes for a protein of 363 amino acids
     with a predicted mol. mass of 38.5 kDA. The XYL2 gene is
     actively expressed in S. cerevisiae transformants. S. cerevisiae cells
     transformed with a plasmid, pRD1, containing both the xylose
     reductase gene (XYL1) and the xylitol
     dehydrogenase gene (XYL2), were able to grow on xylose
     as a sole carbon source. In contrast to aerobic glucose metabolism, S.
     cerevisiae XYL1-XYL2 transformants utilize xylose almost entirely
     oxidatively.
st
     Pichia xylitol dehydrogenase gene XYL2
     sequence; transformation Saccharomyces xylitol
     dehydrogenase gene Pichia
IT
     Saccharomyces cerevisiae
        (cloning and expression in, of xylitol dehydrogenase
        and xylose reductase genes of Pichia
        stipitis)
     Protein sequences
IT
        (of xylitol dehydrogenase, of Pichia stipitis,
        complete)
IT
     Transformation, genetic
        (of Saccharomyces cerevisiae, with Pichia stipitis xylose metabolism
        genes XYL1 and XYL2)
IΤ
     Codon
     RL: BIOL (Biological study)
        (usage of, in xylitol dehydrogenase gene
        XYL2, of Pichia stipitis)
IT
     Pichia stipitis
        (xylitol dehydrogenase gene XYL2 of,
        structure and cloning and expression in Saccharomyces cerevisiae of)
     Deoxyribonucleic acid sequences
IT
        (xylitol dehydrogenase-specifying, of Pichia
        stipitis, complete)
IT
     Plasmid and Episome
        (pRD1, cloning vector for xylitol dehydrogenase and
        xylose reductase genes, expression in
        Saccharomyces cerevisiae of)
ΙT
     Gene and Genetic element, microbial
     RL: BIOL (Biological study)
        (XYL1, for xylose reductase, of Pichia stipitis,
        cloning and expression in Saccharomyces cerevisiae of)
```

IΤ Gene and Genetic element, microbial RL: BIOL (Biological study) (XYL2, for xylitol dehydrogenase, of Pichia stipitis, structure and cloning and expression in Saccharomyces cerevisiae of) IT136511-83-6 RL: PRP (Properties) (amino acid sequence of) IT 95829-40-6, Xylose reductase RL: PRP (Properties) (gene for, of Pichia stipitis, cloning and expression in Saccharomyces cerevisiae of) 9028-16-4, Xylitol dehydrogenase IT RL: PRP (Properties) (gene for, of Pichia stipitis, sequence and expression in Saccharomyces cerevisiae of) IT 58-86-6, Xylose, biological studies RL: BPR (Biological process); BSU (Biological study, unclassified); BIOL (Biological study); PROC (Process) (metabolism of, by Saccharomyces cerevisiae transformant containing Pichia stipitis genes XYL1 and XYL2) 136510-55-9, Deoxyribonucleic acid (Pichia stipitis clone pD1 gene ΙT XYL2 plus 5'- and 3'-flanking region fragment) RL: PRP (Properties) (nucleotide sequence in) 136510-54-8, Deoxyribonucleic acid (Pichia stipitis clone pD1 gene TT XYL2) RL: PRP (Properties); BIOL (Biological study) (nucleotide sequence of) ANSWER 24 OF 33 HCAPLUS COPYRIGHT 2004 ACS on STN L49 1990:492753 HCAPLUS ANDN 113:92753 ED Entered STN: 16 Sep 1990 Isolation of cDNA clones using yeast artificial ΤI chromosome probes Elvin, P.; Slynn, G.; Black, D.; Graham, A.; Butler, R.; Riley, J.; Anand, ΑU R.; Markham, A. F. Eep. Biotechnol., ICI Pharm., Cheshire, SK10 4TG, UK CS Nucleic Acids Research (1990), 18(13), 3913-17 SO CODEN: NARHAD; ISSN: 0305-1048 DTJournal English LA3-4 (Biochemical Genetics) CC The cloning of large DNA fragments of hundreds of kilobases in AB yeast artificial chromosomes has simplified the anal. of regions of the genome previously cloned by cosmid walking. The mapping of expressed sequences within cosmid contigs has relied on the association of genes with sequence motifs defined by rare-cutting endonucleases, and the identification of sequence conservation between species. reasoned that if the contribution of repetitive sequences to filter hybridizations could be minimized, then the use of large cloned DNAs as hybridization probes to screen cDNA libraries would greatly simplify the characterization of hitherto unidentified genes. The use of this approach is demonstrated by using a YAC, containing 180kb of human genomic DNA including the aldose reductase gene, as a probe to isolate an aldose reductase cDNA from a Agt11 human fetal liver cDNA library. human aldose reductase cDNA detection YAC; cDNA STdetection cosmid YAC gene probe; yeast artificial chromosome probe cDNA identification Gene and Genetic element, animal TT

RL: BIOL (Biological study)

```
(for aldose reductase, of human, yeast
        artificial chromosome vector for isolation of cDNA for)
IT
     Gene and Genetic element
     RL: BIOL (Biological study)
        (isolation of, yeast artificial chromosome containing
        genomic inserts as probes for)
IT
     Molecular cloning
        (yeast artificial chromosome as probe for, cDNA
        library screening with)
IT
     Chromosome
        (yeast artificial, isolation of cDNA clones using
        chromosomal probes cloned in)
IT
     9028-31-3
     RL: PRP (Properties)
        (cDNA library containing sequence for human, homologous gene in
        yeast artificial chromosome as probe for detection
        of)
L49
     ANSWER 25 OF 33 HCAPLUS COPYRIGHT 2004 ACS on STN
ΑN
     1990:422145 HCAPLUS
DN
     113:22145
ED
     Entered STN: 21 Jul 1990
TI
     Xylulokinase activity in various yeasts including
     Saccharomyces cerevisiae containing the cloned xylulokinase
     gene
ΑU
     Deng, Xue Xing; Ho, Nancy W. Y.
CS
     A. A. Potter Eng. Cent., Purdue Univ., West Lafayette, IN, 47907, USA
     Applied Biochemistry and Biotechnology (1990), 24-25, 193-9
SO
     CODEN: ABIBDL; ISSN: 0273-2289
DT
     Journal
LA
     English
     16-5 (Fermentation and Bioindustrial Chemistry)
CC
     Section cross-reference(s): 3, 10
AB
     D-Xylose is a major constituent of hemicellulose, which makes up 20-30% of
     the renewable biomass in nature. D-Xylose can be fermented by most
     yeasts, including S. cerevisiae, by a 2-stage process. In this
     process, xylose is 1st converted to xylulose in vitro by xylose (glucose)
     isomerase, and the latter sugar is then fermented by yeast to
     EtOH. With the availability of an inexpensive source of xylose isomerase
     produced by recombinant Escherichia coli, this process of fermenting
     xylose to EtOH can become quite effective. Yeast xylose and
     xylulose fermentation was further improved by cloning and overexpression of the
     xylulokinase gene. For instance, the level of
     xylulokinase activity in S. cerevisiae was increased 230-fold by
     cloning its xylulokinase gene on a high copy-number
     plasmid, coupled with fusion of the gene with an
     effective promoter. The resulting genetically engineered yeasts
     can ferment xylose and xylulose more than twice as fast as the parent
    yeast.
ST
     xylulokinase gene cloning yeast ethanol
     fermn; Saccharomyces xylulose fermn xylulokinase gene
IT
     Fermentation
        (ethanol, from xylose by yeast, xylulokinase
        gene cloning in)
     Gene and Genetic element, microbial
IT
     RL: PROC (Process)
        (for xylulokinase, cloning of, in yeast for ethanol
        fermentation)
    Molecular cloning
TT
        (of xylulokinase gene, in yeast for
        ethanol fermentation)
IT
     Yeast
```

(xylulokinase activities in)

```
IT
     Saccharomyces cerevisiae
     (xylulokinase gene cloning in, for ethanol fermentation) 58-86-6, Xylose, biological studies
IT
     RL: BIOL (Biological study)
        (ethanol fermentation of, by yeast, xylulokinase
        gene cloning in)
IT
     9030-58-4, Xylulokinase
     RL: BIOL (Biological study)
        (gene for, cloning of, in yeast for ethanol fermentation)
IT
     64-17-5P, Ethanol, preparation
     RL: BMF (Bioindustrial manufacture); BIOL (Biological study); PREP
     (Preparation)
        (manufacture of, from xylose by yeast, xylulokinase
        gene cloning in)
     ANSWER 26 OF 33 HCAPLUS COPYRIGHT 2004 ACS on STN
L49
     1990:94337 HCAPLUS
AN
DN
     112:94337
     Entered STN: 18 Mar 1990
ED
     Purification, characterization, and amino terminal sequence of
TI
     xylose reductase from Candida shehatae
     Ho, N. W. Y.; Lin, F. P.; Huang, S.; Andrews, P. C.; Tsao, G. T.
ΑU
     Lab. Renewable Resourc. Eng., Purdue Univ., West Lafayette, IN, 47907, USA
CS
SO
     Enzyme and Microbial Technology (1990), 12(1), 33-9
     CODEN: EMTED2; ISSN: 0141-0229
DT
     Journal
LA
     English
CC
     7-2 (Enzymes)
     Section cross-reference(s): 16
AB
     A convenient and reliable procedure for the purification of xylose
     reductase from C. shehatae to near homogeneity was developed. The
     amino acid composition and N-terminal sequence of the enzyme were also
     analyzed. C. shehatae seems to contain only 1 xylose
     reductase, but the enzyme has a dual coenzyme specificity for both
     NADPH and NADH. The enzyme is remarkably stable at room temperature and
     4°. Xylose fermentation is by NADH-dependent activity.
     xylose reductase Candida; xylose fermn xylose
ST
     reductase NADH Candida
     Protein sequences
IT
        (of xylose reductase N-terminus, of Candida
        shehatae)
IT
     Amino acids, biological studies
     RL: BIOL (Biological study)
        (of xylose reductase, of Candida shehatae)
     Fermentation
TТ
        (of xylose, by xylose reductase of yeast,
        NADH-dependent activity in)
IT
     Candida shehatae
        (xylose reductase of, purification and N-terminal amino
        acid sequence and other properties of)
     58-86-6, D-Xylose, biological studies
IT
     RL: BIOL (Biological study)
        (fermentation of, by yeast xylose reductase,
        NADH-dependent activity in)
IT
     95829-40-6P, Xylose reductase
     RL: PREP (Preparation)
        (of Candida shehatae, purification and N-terminal amino acid sequence and
        other properties of)
     53-57-6, NADPH
IT
     RL: BIOL (Biological study)
        (xylose reductase of Candida shehatae requirement
        for NADH and)
     58-68-4, NADH
```

ΙT

RL: BIOL (Biological study) (xylose reductase of Candida shehatae requirement for NADPH and) ANSWER 27 OF 33 HCAPLUS COPYRIGHT 2004 ACS on STN AN 1989:611729 HCAPLUS DN111:211729 ED Entered STN: 09 Dec 1989 Metabolism of D-xylose in Schizosaccharomyces pombe cloned with a xylose ΤI isomerase gene Chan, Err Cheng; Ueng, Peter Pear; Chen, Li Fu AU Dep. Food Sci., Purdue Univ., West Lafayette, IN, 47907, USA CS Applied Microbiology and Biotechnology (1989), 31(5-6), 524-8 SO CODEN: AMBIDG; ISSN: 0175-7598 DT Journal LA English CC 10-2 (Microbial Biochemistry) Section cross-reference(s): 3 The Escherichia coli xylose isomerase gene was transformed into AΒ S. pombe for direct D-xylose utilization. In order to understand D-xylose metabolism and determine the limiting factors on D-xylose utilization by the transformed yeast, D-xylose transport, xylose isomerization, and xylulose phosphorylation were investigated. The results indicated that low activity of xylose isomerization in the cloned yeast was the limiting step for D-xylose fermentation An in vitro study showed that yeast proteases decreased xylose isomerase activity. Xylitol, a byproduct of D-xylose fermentation, had no effect on the activity of xylose isomerase. xylose metab Schizosaccharomyces gene cloning; Escherichia xylose isomerase gene cloning yeast Gene and Genetic element, microbial IT RL: BIOL (Biological study) (for xylose isomerase, of Escherichia coli, xylose metabolism by recombinant Schizosaccharomyces pombe containing) Biological transport IT (of xylose, by recombinant xylose isomerase-containing Schizosaccharomyces pombe) Schizosaccharomyces pombe IT(xylose metabolism by recombinant xylose isomerase-containing) 551-84-8, D-Xylulose 58-86-6, D-Xylose, biological studies IT RL: BPR (Biological process); BSU (Biological study, unclassified); BIOL (Biological study); PROC (Process) (metabolism of, by Schizosaccharomyces pombe with xylose isomerase gene) IT9030-58-4, Xylulokinase RL: BIOL (Biological study) (of transformed Schizosaccharomyces pombe containing xylose isomerase) IT 9023-82-9, Xylose isomerase RL: BIOL (Biological study) (of Escherichia coli, xylose metabolism by Schizosaccharomyces pombe with) ANSWER 28 OF 33 HCAPLUS COPYRIGHT 2004 ACS on STN L49 1989:510301 HCAPLUS AN DN111:110301 Entered STN: 01 Oct 1989 ED ΤI Construction of yeast xylulokinase mutant by recombinant DNA techniques ΑU Stevis, Panayiotis E.; Ho, Nancy W. Y. Dep. Foods Nutr., Purdue Univ., West Lafayette, IN, 47907, USA CS Applied Biochemistry and Biotechnology (1989), Volume Date 1988, SO 20-21, 327-34 CODEN: ABIBDL; ISSN: 0273-2289

DT

Journal

```
LA
     English
CC
     3-5 (Biochemical Genetics)
AΒ
     A Saccharomyces cerevisiae xylulokinase mutant was constructed
     by using the cloned yeast xylulokinase gene,
     XYK-Sc, and the gene disruption technique. The S. cerevisiae
     LEU2 gene was used to disrupt the XYK-Sc gene cloned
     on pLSK4 by insertion into the unique HindIII site of the gene.
     The disrupted gene was liberated from the remainder of the
     plasmid with XhoI digestion, yielding a 4.4 kb DNA fragment. Transformation of a S. cerevisiae leu2 mutant with this
     fragment and selection for Leu+ complementation resulted in the isolation
     of transformants that were unable to grow in pure xylulose medium.
     ability to grow in xylulose medium and increased xylulokinase
     activity were obtained by transforming the mutant with a plasmid
     -borne wild-type XYK-Sc gene. Insertional inactivation of the
     chromosomal XYK-Sc gene was also demonstrated by
     xululokinase assays.
ST
     Saccharomyces xylulokinase gene mutation recombinant
     DNA
TT
     Mutation
        (in xylulokinase gene, of Saccharomyces cerevisiae,
        gene disruption technique in construction of)
IT
     Saccharomyces cerevisiae
        (xylulokinase gene of, mutation construction in, by
        gene disruption technique)
IT
     Gene and Genetic element, microbial
     RL: BIOL (Biological study)
        (XYK, for xylulokinase, of Saccharomyces cerevisiae, mutation
        in, gene disruption technique for)
IT
     9030-58-4, Xylulokinase
     RL: PRP (Properties)
        (gene for, of Saccharomyces cerevisiae, construction of
        mutation in, by gene disruption technique)
    ANSWER 29 OF 33 HCAPLUS COPYRIGHT 2004 ACS on STN
     1989:472444 HCAPLUS
AN
DN
     111:72444
ED
     Entered STN: 03 Sep 1989
ΤI
     Cloning of yeast xylulokinase gene by
     complementation of E. coli and yeast mutations
ΑU
     Ho, Nancy W. Y.; Chang, Sue Fen
CS
     Lab. Renew. Resour. Eng., Purdue Univ., West Lafayette, IN, USA
     Enzyme and Microbial Technology (1989), 11(7), 417-21
     CODEN: EMTED2; ISSN: 0141-0229
DT
     Journal
LΑ
     English
CC
     3-4 (Biochemical Genetics)
     The gene encoding yeast (Saccharomyces cerevisiae)
AB
     xylulokinase has been isolated by complementation of Escherichia
     coli xylulokinase mutations. Through subcloning, the
     gene has been localized on two HindIII fragments (1.2 and 2.4 bp).
     Within these HindIII fragments, there lies a 2.2-kb Xho fragment which
     contains the structural gene of yeast
     xylulokinase. Upon insertion of a selectable gene into
     the XhoI fragment, the resulting recombination fragment has been used to
     construct a yeast xylulokinase mutant by the
     gene disruption technique. The cloned xylulokinase
     gene was found to be able to complement such a
     xylulokinase mutant.
     Saccharomyces xylulokinase gene cloning Escherichia
ST
     Gene and Genetic element, microbial
IT
     RL: PROC (Process)
```

(for xylulokinase, of yeast, cloning of)

```
IT
     Complementation, genetic
        (of xylulokinase gene mutation in Escherichia coli,
        by yeast gene)
ΙT
     Molecular cloning
        (of xylulokinase gene, of yeast)
     Saccharomyces cerevisiae
IT
        (xylulokinase gene of, cloning of)
IT
     Escherichia coli
        (xylulokinase gene of, mutation in, yeast
        gene complementation of)
IT
     9030-58-4, Xylulokinase
     RL: PRP (Properties)
        (gene for, of yeast, cloning of)
     ANSWER 30 OF 33 HCAPLUS COPYRIGHT 2004 ACS on STN
L49
     1989:434409 HCAPLUS
AN
DN
     111:34409
ED
     Entered STN: 05 Aug 1989
     A nuclear yeast gene (GCY) encodes a polypeptide with
TI
     high homology to a vertebrate eye lens protein
     Oechsner, Ulrich; Magdolen, Viktor; Bandlow, Wolfhard
AII
CS
     Inst. Genet. Microbiol., Munich, D-8000, Fed. Rep. Ger.
     FEBS Letters (1988), 238(1), 123-8
SO
     CODEN: FEBLAL; ISSN: 0014-5793
DT
     Journal
     English
LA
     3-3 (Biochemical Genetics)
CC
     Section cross-reference(s): 6
AΒ
     The nuclear gene for a yeast protein is described
     which shows unexpectedly high homol. with mammalian aldo/keto reductases
     as well as with \rho\text{-crystallin,} one of the prominent proteins of the
     frog eye lens. Although it could be proven that the gene occurs
     as a single copy in the haploid yeast genome, replacement of the
     intact by a disrupted, nonfunctional allele led to no obvious phenotype,
     indicating that the gene is dispensable. The gene was
     assigned to chromosome XV. It is transcribed in vivo into an
     mRNA of about 1300 bases with a coding capacity for a protein of 312 amino
     acids (estimated Mr, 35,000).
st
     Saccharomyces gene GCY protein sequence
IT
     Saccharomyces cerevisiae
        (gene GCY protein of, homol. of, to vertebrate eye lens
        protein)
IT
     Protein sequences
        (of gene GCY protein, of Saccharomyces cerevisiae, complete)
ΙT
     Eye, composition
        (protein of, of vertebrate, gene GCY protein of Saccharomyces
        cerevisiae homol. with)
IT
     Proteins, specific or class
     RL: BIOL (Biological study)
        (gene GCY, gene for, of Saccharomyces cerevisiae,
        nucleotide and encoded peptide sequences of)
IT
     Deoxyribonucleic acid sequences
        (gene GCY protein-specifying, of Saccharomyces cerevisiae,
        complete)
IT
     Gene and Genetic element, microbial
     RL: BIOL (Biological study)
        (GCY, of Saccharomyces cerevisiae, nucleotide and encoded peptide
        sequences of)
IT
     Crystallins
     RL: BIOL (Biological study)
        (p-, gene \bar{\text{GCY}} protein of Saccharomyces cerevisiae homol. with, of frog)
```

IT

Chromosome

```
(Saccharomyces cerevisiae XV, gene GCY localization on)
    121548-71-8, Protein (Saccharomyces cerevisiae gene GCY
IT
    reduced)
    RL: PRP (Properties)
        (amino acid sequence of)
     121547-66-8, Deoxyribonucleic acid (Saccharomyces cerevisiae gene
IΤ
     RL: PRP (Properties); BIOL (Biological study)
        (nucleotide sequence of)
     9028-31-3
TΤ
     RL: PRP (Properties)
        (of eye lens, of rat, gene GCY protein of Saccharomyces
        cerevisiae homol. with)
     9028-12-0, Aldehyde reductase
TΤ
     RL: PRP (Properties)
        (of liver of human, gene GCY protein of Saccharomyces
        cerevisiae homol. with)
     55976-95-9
IT
    RL: PRP (Properties)
        (of lung of cattle, gene GCY protein of Saccharomyces
        cerevisiae homol. with)
L49 ANSWER 31 OF 33 HCAPLUS COPYRIGHT 2004 ACS on STN
NΑ
     1989:129846 HCAPLUS
DN
     110:129846
ED
    Entered STN: 15 Apr 1989
    Cloning the yeast xylulokinase gene for the
TТ
     improvement of xylose fermentation
    Chang, Sue Feng; Ho, Nancy W. Y.
ΑU
    Lab. Renew. Resour. Eng., Purdue Univ., West Lafayettye, IN, 47907, USA
CS
    Applied Biochemistry and Biotechnology (1988), 17, 313-18
SO
    CODEN: ABIBDL; ISSN: 0273-2289
דת
    Journal
     English
LΑ
     3-4 (Biochemical Genetics)
CC
     Section cross-reference(s): 16
    Plasmids pLSK1 or pLSK3 containing a Saccharomyces cerevisiae
AB
    DNA fragment complemented Escherichia coli xylulokinase
     mutations. The cloned yeast DNA probably contains all
     the necessary structural elements of a yeast gene
     encoding a yeast protein (enzyme). E. coli xylulokinase
     mutants harboring either pLSK1 or pLSK3 synthesized xylulokinase
     in the absence of xylose induction as well as in the presence of glucose
     (insensitive to glucose inhibition). Thus, the yeast
    DNA fragment cloned on pLSK1 or pLSK3 at least contains the
     structural gene encoding S. cerevisiae xylulokinase.
    xylulokinase gene Saccharomyces complementation
ST
    Escherichia
ΙT
    Escherichia coli
        (expression in, of xylulokinase gene of
        Saccharomyces cerevisiae)
     Gene and Genetic element, microbial
IT
     RL: BIOL (Biological study)
        (for xylulokinase, of Saccharomyces cerevisiae, structure and
        expression in Escherichia coli of)
IT
     Complementation, genetic
        (of xylulokinase mutants of Escherichia coli, by
        Saccharomyces cerevisiae gene)
IT
     Saccharomyces cerevisiae
        (xylulokinase gene of, structure and expression in
        Escherichia coli of)
IT
     9030-58-4, Xylulokinase
```

RL: PRP (Properties)

(gene for, of yeast, structure and expression in Escherichia coli of)

```
ANSWER 32 OF 33 HCAPLUS COPYRIGHT 2004 ACS on STN
L49
     1988:33023 HCAPLUS
AN
DN
     108:33023
     Entered STN: 06 Feb 1988
ED
     Cloning of the Pachysolen tannophilus xylulokinase gene
ТT
     by complementation in Escherichia coli
     Stevis, Panayiotis E.; Huang, James J.; Ho, Nancy W. Y.
ΑU
     Lab. Renewable Resour. Eng., Purdue Univ., West Lafayette, IN, 47907, USA
CS
     Applied and Environmental Microbiology (1987), 53(12), 2975-7
SO
     CODEN: AEMIDF; ISSN: 0099-2240
DT
     Journal
LA
     English
     3-4 (Biochemical Genetics)
CC
     The gene coding for xylulokinase has been isolated
AB
     from the yeast P. tannophilus by complementation of E. coli
     xylulokinase (xylB) mutants. Through subcloning, the gene
     has been localized at one end of a 3.2-kilobase EcoRI-PstI fragment.
     Expression of the cloned gene was insensitive to glucose
     inhibition. Further, the cloned gene did not cross-hybridize
     with E. coli and Saccharomyces cerevisiae xylulokinase
     Pachysolen xylulokinase gene cloning Escherichia
st
IT
     Escherichia coli
        (cloning in, of xylulokinase gene of Pachysolen
        tannophilus)
     Molecular cloning
IT
        (of xylulokinase gene of Pachysolen tannophilus, in
        Escherichia coli)
IT
     Pachysolen tannophilus
        (xylulokinase gene of, cloning in Escherichia coli
     Gene and Genetic element, microbial
ΙT
     RL: BIOL (Biological study)
        (XYK, for xylulokinase, of Pachysolen tannophilus, cloning in
        Escherichia coli of)
IT
     9030-58-4, Xylulokinase
     RL: PRP (Properties)
        (gene for, of Pachysolen tannophilus, cloning in Escherichia
        coli of)
     ANSWER 33 OF 33 HCAPLUS COPYRIGHT 2004 ACS on STN
     1986:419795 HCAPLUS
AN
     105:19795
DN
     Entered STN: 26 Jul 1986
ED
     Towards a xylose fermenting yeast
TI
     Hollenberg, Cornelis P.; Wilhelm, Martin
ΑU
     Inst. Mikrobiol., Univ. Duesseldorf, Duesseldorf, 4000, Fed. Rep. Ger.
CS
     Eur. Congr. Biotechnol., 3rd (1984), Volume 3, 175-9 Publisher:
SO
     Verlag Chemie, Weinheim, Fed. Rep. Ger.
     CODEN: 55BBA6
DT
     Conference
LΑ
     English
     3-4 (Biochemical Genetics)
CC
     As a 1st step towards the cloning and expression of the xylose isomerase
AΒ
     [9023-82-9] gene of Bacillus subtilis in Saccharomyces
     cerevisiae, a DNA fragment for B. subtilis was isolated which
```

encodes xylose isomerase and xylulokinase [9030-58-4]. The DNA fragment was cloned on recombinant plasmids

and used to transform an Escherichia coli strain that was isomerase deficient. The **plasmids** pMW1 and pMW11 were largely identical

except for a region of .apprx.1 kilobase absent from pMW11.

Plasmid pMW11 contained the originally cloned B. subtilis

DNA fragment which had been changed in pMW1 by the addition of an insertion element. The presence of pMW1 in E. coli transformants led to wild-type levels of isomerase activity, whereas in pMW11 transformants almost no activity could be detected. Thus, the expression of the B. subtilis gene was observed only after the insertion of an IS element. Furthermore, the expression of the xylose isomerase from pMW1 was not subjected to regulation in the E. coli transformants. The transformants also expressed high levels of xylulokinase.

ST xylose isomerase **gene** Bacillus cloning; **yeast** xylose fermn cloning

IT Escherichia coli

(cloning in, of xylose isomerase and xylulokinase genes of Bacillus subtilis)

IT Gene and Genetic element, microbial

RL: BIOL (Biological study)

(for xylose isomerase and xylulokinase, of Bacillus subtilis, cloning and expression in Escherichia coli of)

IT Molecular cloning

(of xylose isomerase and xylulokinase genes, of Bacillus subtilis in Escherichia coli)

IT Bacillus subtilis

(xylose isomerase and xylulokinase genes of, cloning in Escherichia coli of)

IT Gene and Genetic element, microbial

(insertion sequence, xylose isomerase **gene** containing, of Bacillus subtilis, Escherichia coli expression of)

IT Plasmid and Episome

(pMW1, xylose isomerase and xylulokinase genes of

Bacillus subtilis cloning on, for expression in Escherichia coli)

IT 9023-82-9 **9030-58-4**

RL: PRP (Properties)

(gene for, of Bacillus subtilis, cloning and expression in Escherichia coli of)

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 THE TIME RANGE CODE WILL ALSO CHANGE FROM 018 TO 2004.

 SDIS USING THE TIME RANGE CODE WILL NEED TO BE UPDATED.

 FOR FURTHER DETAILS: http://thomsonderwent.com/chem/polymers/ <<<

=> d all abeq tech abex tot L66 ANSWER 1 OF 11 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN **2003-748653** [70] WPIX DNC C2003-205320 Improvement of ethanol production from xylose employs strain of Saccharomyces cerevisiae additionally comprising specific genes that are over-expressed. DC D16 E17 H06 HAHN-HAEGERDAL, B; JOENSSON, L; WAHLBOM, F (FORS-N) FORSKARPATENT I SYD AB CYC 102 PΙ WO 2003078642 A1 20030925 (200370) * EN 28p C12P007-10 RW: AT BE BG CH CY CZ DE DK EA EE ES FI FR GB GH GM GR HU IE IT KE LS LU MC MW MZ NL OA PT RO SD SE SI SK SL SZ TR TZ UG ZM ZW W: AE AG AL AM AT AU AZ BA BB BG BR BY BZ CA CH CN CO CR CU CZ DE DK DM DZ EC EE ES FI GB GD GE GH GM HR HU ID IL IN IS JP KE KG KP KR KZ LC LK LR LS LT LU LV MA MD MG MK MN MW MX MZ NO NZ OM PH PL PT RO RU SC SD SE SG SK SL TJ TM TN TR TT TZ UA UG US UZ VC VN YU ZA ADT WO 2003078642 A1 WO 2003-SE397 20030311 PRAI SE 2002-855 20020319 ICM C12P007-10 ICS C12N001-19 AB WO2003078642 A UPAB: 20031030 NOVELTY - Improving ethanol production from xylose employs strain of Saccharomyces cerevisiae additionally comprising over-expressed PET18 (YCR020c), HXT5 (YHR096c), GAL2 (YLR081w), SOL3 (YHR163w), GND1 (YHR183w), TAL1 (YLR354c), TKL1 (YPR074c), PCK1 (YKR097w), ICL1 (YER065c), MLS1 (YNL117w), GAL1 (YBR020c), GAL7 (YBR018c), GAL10 (YBR019c), and/or CAT8 (YMR280c) genes. Open reading frames are given in brackets. DETAILED DESCRIPTION - Improving ethanol production from xylose employs strain of Saccharomyces cerevisiae comprising genes for over-expression of xylose reductase, xylitol dehydrogenase and xylulokinase and additionally over-expressed PET18 (YCR020c), HXT5 (YHR096c), GAL2 (YLR081w), SOL3 (YHR163w), GND1 (YHR183w), TAL1 (YLR354c), TKL1 (YPR074c), PCK1 (YKR097w), ICL1 (YER065c), MLS1 (YNL117w), GAL1 (YBR020c), GAL7 (YBR018c), GAL10 (YBR019c), and/or CAT8 (YMR280c) genes. Open reading frames are given in brackets. USE - For improving ethanol production from xylose. ADVANTAGE - The invention improves xylose utilization. Dwq.0/0CPI FS FΑ AB; DCN MC CPI: D05-B03; E10-E04E2; E11-M; H06-B TECH UPTX: 20031030 TECHNOLOGY FOCUS - BIOTECHNOLOGY - Preferred Method: TEC1 (YBR083w), ARR1 (YPR199c), MIG1 (YGL035c), and/or MIG2 (YGL209w) are deleted. One or more of the genes is over expressed and one or more of the genes is deleted. L66 ANSWER 2 OF 11 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN 2002-097582 [13] WPIX DNC C2002-030382 ΤI Obtaining recombinant yeast of Saccharomyces cerevisiae for fermenting lignocellulose raw materials to ethanol, comprises introducing deoxyribonucleic acid into yeast DC D16 D17 E17 IN CORDERO OTERO, R R; HAHN-HAEGERDAL, B; VAN ZYL, W H; HAHN-HAGERDAL, B;

HAHNAEGERDAL, B

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(FORS-N) FORSKARPATENT I SYD; (FORS-N) FORSKARPATENT I SYD AB; (OTER-I)
PA
     CORDERO OTERO R R; (HAHN-I) HAHN-HAGERDAL B; (VZYL-I) VAN ZYL W H
CYC
     WO 2001088094 A1 20011122 (200213)* EN
                                              18p
                                                     C12N001-19
PΤ
        RW: AT BE CH CY DE DK EA ES FI FR GB GH GM GR IE IT KE LS LU MC MW MZ
            NL OA PT SD SE SL SZ TR TZ UG ZW
         W: AE AG AL AM AT AU AZ BA BB BG BR BY BZ CA CH CN CO CR CU CZ DE DK
            DM DZ EE ES FI GB GD GE GH GM HR HU ID IL IN IS JP KE KG KP KR KZ
            LC LK LR LS LT LU LV MA MD MG MK MN MW MX MZ NO NZ PL PT RO RU SD
            SE SG SI SK SL TJ TM TR TT TZ UA UG US UZ VN YU ZA ZW
     AU 2001058985 A 20011126 (200222)
                                                     C12N001-19
     EP 1282686
                  A1 20030212 (200312) EN
                                                     C12N001-19
         R: AL AT BE CH CY DE DK ES FI FR GB GR IE IT LI LT LU LV MC MK NL PT
            RO SE SI TR
     US 2003157675 A1 20030821 (200356)
                                                     C12P007-06
     ZA 2002009448 A 20031231 (200408)
                                              29p
                                                     C12N000-00
ADT WO 2001088094 A1 WO 2001-SE1061 20010515; AU 2001058985 A AU 2001-58985
     20010515; EP 1282686 A1 EP 2001-932462 20010515, WO 2001-SE1061 20010515;
     US 2003157675 A1 Cont of WO 2001-SE1061 20010515, US 2002-293255 20021114;
     ZA 2002009448 A ZA 2002-9448 20021120
FDT AU 2001058985 A Based on WO 2001088094; EP 1282686 A1 Based on WO
     2001088094
PRAI ZA 2000-2363
                      20000515
     ICM C12N000-00; C12N001-19; C12P007-06
     ICS C12N001-18; C12N015-74; C12P007-10
AB
     WO 200188094 A UPAB: 20020226
     NOVELTY - Obtaining recombinant yeast of Saccharomyces
     cerevisiae, comprising introducing DNA into a yeast,
     where the obtained yeast introduces genes encoding
     xylose reductase, xylithol dehydrogenase and
     xylulokinase, is new.
          USE - For obtaining recombinant yeast of
     Saccharomyces cerevisiae useful for fermenting lignocellulose raw
     materials to produce ethanol.
          ADVANTAGE - The obtained recombinant yeast is efficiently
     capable of fermenting lignocellulose raw materials to produce ethanol.
     Dwg.0/2
FS
     CPI
FΑ
     AB; DCN
     CPI: D05-B03; D05-C03B; D05-H05; D05-H08;
MC
          D05-H12A; D05-H17A3; E10-E04E2
                    UPTX: 20020226
TECH
     TECHNOLOGY FOCUS - BIOLOGY - Preferred Product: The yeast is
     capable of producing one or more lignocellulose utilizing enzymes of
     xylose reductase, xylithol dehydrogenase, or
     xylulokinase. Preferred Enzymes: The enzymes of the yeast
     is of the genus Saccharomyces cerevisiae and Pichia stipitis.
     The xylose reductase or xylitol
     dehydrogenase lignocellulose utilizing enzyme can be obtained from
     Pichia stipitis. The xylulokinase enzyme is obtained from
     Saccharomyces cerevisiae. Preferred Medium: The growth medium by
     the recombinant yeast comprises glucose and xylose. Preferred
     Method: The method includes isolating mutants by ethyl methanesulfonate
     treatment. The mutants show a growth rate over basic strain of more than
     30%. The recombinant strain is maintained in continuous culture on xylose
     as carbon source at dilution rate of 0.1/h with growth rate on xylose of
     0.14-0.15/h and biomass yield of 0.4 g/g on xylose at aerobic growth. It
     utilizes 20 g/L and 15-16 g/L of xylose (4-5 g/L residual) in a continuous
     culture from a 20 g/L xylose and 20 g/L of glucose feed. Preferred Strain:
     The Saccharomyces cerevisiae strain is Saccharomyces
     cerevisiae USM21, which has been deposited under CBS 102678. It is
     (non-)detoxified lignocellulose hydrolysates, or (soft or hard)wood
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derived hydrolysate. Preferred Mutants: The mutant is a xylose-fermenting

mutant XYLUSM125, which is deposited under CBS 102679 or XYLUSM145, which is deposited under CBS 102680.

ABEX UPTX: 20020226

 ${\tt EXAMPLE}$ - ${\tt XYLUSM125}$ mutant was grown in 20 g/L xylose in minimal medium and established XYLUSM125 in a continuous culture on 20 g/L xylose using dilution rate of 0.1/h (aerobic fermentation condition). The growth rate obtained on xylose as carbon source was 0.14-0.15/h and the biomass yield was 0.4 g/g to have 8 g/L biomass on 20 g/L xylose as carbon source. When the feed was changed to 20 g/L xylose and 20 g/L glucose the biomass had raised to 18 g/L and the result was only 4-5 g/L xylose remained. The XYLUSM125 mutant utilized 20 g/L glucose and 15-16 g/L xylose in continuous fermentation.

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L66 ANSWER 3 OF 11 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN
                       WPTX
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AΝ 2002-097498 [13]

DNC C2002-030322

Site-specific insertion in Zymomonas mobilis, comprises transforming Zymomonas through homologous recombination with deoxyribonucleic acid fragment having interrupted sequence.

DC

CHOU, Y; ZHANG, M IN

(MIDE) MIDWEST RES INST PA

CYC 95

C12N015-63 WO 2001083784 A2 20011108 (200213)* EN 27p PΙ

RW: AT BE CH CY DE DK EA ES FI FR GB GH GM GR IE IT KE LS LU MC MW MZ NL OA PT SD SE SL SZ TR TZ UG ZW

W: AE AG AL AM AT AU AZ BA BB BG BR BY BZ CH CN CO CR CU CZ DE DK DM DZ EE ES FI GB GD GE GH GM HR HU ID IL IN IS JP KE KG KP KR KZ LC LK LR LS LT LU LV MA MD MG MK MN MW MX MZ NO NZ PL PT RO RU SD SE SG SI SK SL TJ TM TR TT TZ UA UG UZ VN YU ZA ZW

C12N001-21 CA 2304927 A1 20011102 (200213) $\mathbf{E}\mathbf{N}$ C12N015-63 AU 2001051397 A 20011112 (200222) C12N015-09 JP 2003531620 W 20031028 (200373) 33p A2 20031203 (200380) EN C12N015-90

R: AL AT BE CH CY DE DK ES FI FR GB GR IE IT LI LT LU LV MC MK NL PT RO SE SI TR

BR 2001010676 A 20031230 (200409) C12N015-63

ADT WO 2001083784 A2 WO 2001-US11239 20010406; CA 2304927 A1 CA 2000-2304927 20000502; AU 2001051397 A AU 2001-51397 20010406; JP 2003531620 W JP 2001-580391 20010406, WO 2001-US11239 20010406; EP 1366178 A2 EP 2001-924773 20010406, WO 2001-US11239 20010406; BR 2001010676 A BR 2001-10676 20010406, WO 2001-US11239 20010406

FDT AU 2001051397 A Based on WO 2001083784; JP 2003531620 W Based on WO 2001083784; EP 1366178 A2 Based on WO 2001083784; BR 2001010676 A Based on WO 2001083784

PRAI CA 2000-2304927 20000502; US 2000-562613 20000501 ICM C12N001-21; C12N015-09; C12N015-63; C12N015-90 IC

C12N015-11

WO 200183784 A UPAB: 20020226 AΒ

NOVELTY - Site-specific insertion in Zymomonas, comprising interrupting a sequence in a Zymomonas deoxyribonucleic acid (DNA) fragment and transforming the Zymomonas through homologous recombination

with the interrupted fragment, is new.

USE - For insertion inactivation of specific gene products in recombinant Z. mobilis strains which ferment xylose and/or arabinose

ADVANTAGE - The method eliminates the formation of by-products in a Z. mobilis fermentation through the construction of stable recombinant strains to be eliminated.

Dwg.0/10

CPI FS

AB; DCN FA

CPI: D05-B03; D05-H05; D05-H09; D05-H12A; MC

D05-H17A6; E10-E04E2

TECH

UPTX: 20020226 TECHNOLOGY FOCUS - BIOTECHNOLOGY - Preferred Method: Interrupting is by inserting a DNA sequence inside the DNA fragment or by deleting a DNA sequence inside the DNA fragment. The DNA fragment (e.g., ldh) is encoding a structural protein in a metabolic pathway of the by-product (e.g., lactic acid) to be eliminated. The interrupted DNA fragment is ligated with a plasmid vector. Transforming the Zymomonas mobilis organism is through homologous recombination with the interrupted fragment of the plasmid vector. The Z. mobilis of the plasmid is cured.

TECHNOLOGY FOCUS - BIOLOGY - Preferred Plasmid: The plasmid is pZB101, pZB102 or pZB121. It may be pZB1962-ldhL-ara. Preferred Insertion: The insertion is ldhL, a selection marker or an operon. The operon encodes structural gene(s) including xylose isomerase, xylulokinase, L-arabinose isomerase, L-ribulokinase, L-ribulose-5-phosphate-4-epimerase, transaldolase or transketolase, and a promoter for expression of the structural gene in Z. mobilis.

ABEX UPTX: 20020226

> EXAMPLE - Plasmids containing 1dh gene of Zymomonas having a tetracycline (Tc) resistant gene insert (1dh::Tc) cassette were used to transform Z. mobilis. The resultant Tc resistance transformants were analyzed by Southern hybridization. Results showed that the ldh::Tc cassette had been inserted into the ldh region of the Zymomonas genome. The gene integration based on homologous recombination in Zymomonas, together with the targeted integration resulting in an inactivated ldh gene, eliminated lactic acid by-product formation in ethanol fermentation.

L66 ANSWER 4 OF 11 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN

2002-089743 [12] WPIX AN

CR 2003-731102 [69]

DNC C2002-027635

Transposon for plasmid vector for constructing strains of Zymomonas mobilis for conversion of pentose sugars into ethanol, comprises operon(s) having structural genes encoding enzymes and promoter(s).

DC D16 E17 H06

IN CHOU, Y; ZHANG, M

(MIDE) MIDWEST RES INST; (CHOU-I) CHOU Y; (ZHAN-I) ZHANG M PΑ

CYC 96

WO 2001083786 A2 20011108 (200212)* EN 49p C12N015-74 PΤ

RW: AT BE CH CY DE DK EA ES FI FR GB GH GM GR IE IT KE LS LU MC MW MZ NL OA PT SD SE SL SZ TR TZ UG ZW

W: AE AG AL AM AT AU AZ BA BB BG BR BY BZ CH CN CO CR CU CZ DE DK DM DZ EE ES FI GB GD GE GH GM HR HU ID IL IN IS JP KE KG KP KR KZ LC LK LR LS LT LU LV MA MD MG MK MN MW MX MZ NO NZ PL PT RO RU SD SE SG SI SK SL TJ TM TR TT TZ UA UG UZ VN YU ZA ZW

C12N001-21 A1 20011102 (200212) EN AU 2001049926 A 20011112 (200222) C12N015-74 US 2002151034 A1 20021017 (200270) C12N001-20 EP 1278876 A2 20030129 (200310) EN C12N015-74

R: AL AT BE CH CY DE DK ES FI FR GB GR IE IT LI LT LU LV MC MK NL PT RO SE SI TR

ADT WO 2001083786 A2 WO 2001-US11334 20010406; CA 2304929 A1 CA 2000-2304929 20000502; AU 2001049926 A AU 2001-49926 20010406; US 2002151034 A1 US 2000-565233 20000501; EP 1278876 A2 EP 2001-923213 20010406, WO 2001-US11334 20010406

FDT AU 2001049926 A Based on WO 2001083786; EP 1278876 A2 Based on WO 2001083786

PRAI CA 2000-2304929 20000502; US 2000-565233 20000501

ICM C12N001-20; C12N001-21; C12N015-74

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ICS C12N015-09; C12N015-11; C12N015-52; C12N015-63; C12N015-90;
          C12P007-04; C12P007-06
    WO 200183786 A UPAB: 20031027
AΒ
    NOVELTY - A transposon for stable insertion of foreign genes
    into a bacterial genome, comprising operon(s) having xylAxylB, araBAD or
     tal/tkt as structural genes encoding enzymes, and promoter(s)
     for expression of the structural genes in the bacterium, where
     the operons are contained inside a pair of inverted insertion sequences,
     and a transposase gene is located outside of the insertion
     sequences, is new.
          USE - The transposon, e.g., Tn5 or Tn10 derivative, is for a
    plasmid vector (e.g., pZB1862-1dhL-ara ATCC Accession Number
     PTA-1798) which transforms a bacterium, i.e. Zymomonas mobilis (claimed).
     The strains of Z. mobilis (e.g., G8 ATCC Accession Number PTA-1796, C25 ATCC
     Accession Number PTA-1799, and AX ATCC Accession Number PTA-1797) are useful in
     the conversion of the cellulose derived pentose sugars, e.g., xylose and
     arabinose, into fuels and chemicals, e.g. ethanol.
          ADVANTAGE - The invention provides for the construction of Z. mobilis
     strains which are capable of fermenting xylose and/or arabinose to ethanol
     through the generation of stable genomic inserts which encode the enzymes
     necessary for xylose and arabinose catabolism. The strains are free of
     antibiotic resistance and stable for more than 40 (at least 45)
     generations in a non-selection media. The strains also demonstrate a high
     specific rate of product formation at close to maximum theoretical product
     yield.
     Dwq.0/18
FS
     CPI
FA
     AB; DCN
     CPI: D05-B03; D05-C07; D05-H12D5; D05-H12E; E10-E04L2;
MC
          H06-B
                    UPTX: 20020221
TECH
     TECHNOLOGY FOCUS - BIOTECHNOLOGY - Preferred Component: The promoter is
     Peno or Pgap.
                    UPTX: 20020221
ABEX
     EXAMPLE - Two operons containing Pgap-xylA/xylB and Peno-talB/tktA were
     assembled in mini-Tn5 and the resulting plasmid was conjugated
     into Z. mobilis. With the help of the transposase located outside of the
     mini-Tn5 cassette, single copies of the two operons were inserted into the

    mobilis genome. Enzymatic analysis of xylose isomerase,

     xylulokinase, transaldolase, and transketolase indicated that all
     the genes coordinately expressed and that the integrated strains
     produced 30-70% of the enzymatic activities of the plasmid
     -bearing strains. These enzymatic levels were enough for the organism to
     grow and ferment xylose to ethanol.
L66 ANSWER 5 OF 11 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN
                        WPIX
     2000-526176 [48]
DNC C2000-156444
     Genetically modified microorganism useful for producing xylitol
     from D-xylulose comprises an exogenous xylitol
     dehydrogenase gene.
DC
     B05 D13 D16 D17 E17
     TAKENAKA, Y; TONOUCHI, N; YOKOZEKI, K
IN
     (AJIN) AJINOMOTO KK; (AJIN) AJINOMOTO CO INC
PΔ
CYC 30
                   A1 20000823 (200048)* EN
                                               21p
                                                      C12P007-18
PΙ
     EP 1029925
         R: AL AT BE CH CY DE DK ES FI FR GB GR IE IT LI LT LU LV MC MK NL PT
            RO SE SI
                                                      C12N015-09
     JP 2000295994 A 20001024 (200059)
                                               16p
                                                      C12P007-18
                  A 20001025 (200104)
     CN 1271017
                                                      C12N001-00
     KR 2000076625 A 20001226 (200134)
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BR 2000000338 A 20010821 (200155)

US 2003148482 A1 20030807 (200358)

C12P007-18

C12P007-18

ADT EP 1029925 A1 EP 2000-102566 20000207; JP 2000295994 A JP 1999-197621 19990712; CN 1271017 A CN 2000-105396 20000209; KR 2000076625 A KR 2000-5838 20000208; BR 200000338 A BR 2000-338 20000208; US 2003148482 A1 Cont of US 2000-500908 20000209, US 2002-277706 20021023

PRAI JP 1999-197621 19990712; JP 1999-31464 19990209

IC ICM C12N001-00; C12N015-09; C12P007-18

ICS C12N001-20; C12N009-04; C12N015-00; C12N015-74

ICA C12N001-21

AB EP 1029925 A UPAB: 20001001

NOVELTY - Microorganism (I) that has been transformed with a **gene** encoding **xylitol dehydrogenase** (XDH) and is capable of supplying reducing power to its culture medium (i.e. of producing sufficient reduced nicotinamide adenine **dinucleotide** (NADH) for XDH-catalyzed conversion of D-xylulose to xylitol) is new.

DETAILED DESCRIPTION - An INDEPENDENT CLAIM is also included for a process for producing xylitol by reacting a microorganism of type (I) with D-xylulose and collecting the resulting xylitol.

USE - The xylitol is useful as a low-calorie, non-carcinogenic sweetener and for fluid therapy in the treatment of diabetes.

ADVANTAGE - (I) is capable of converting D-xylulose to xylitol in the absence of added carbon sources or NADH.

Dwg.0/0

FS CPI

FA AB; DCN

MC CPI: B04-F10A3E; B10-A07; B14-S04; D03-H01A; D05-C03B; D05-C08; D05-H12A; D05-H14A1; D05-H17A3; D06-G; E10-A07

TECH UPTX: 20001001

TECHNOLOGY FOCUS - BIOTECHNOLOGY - Preferred Process: The D-xylulose is produced by means of a microorganism (II) capable of converting D-arabitol to D-xylulose or a microorganism (III) capable of producing D-xylulose from glucose. Preferred Microorganisms: (I) is an Escherichia coli transformant. (II) is a microorganism of the genus Gluconobacter, Achromobacter, Agrobacterium, Alcaligenes, Arthrobacter, Azotobacter, Brevibacterium, Corynebacterium, Enterobacter, Erwinia, Flavobacterium, Micrococcus, Morganella, Nocardia, Planococcus, Proteus, Propionibacterium, Pseudomonas, Rhodococcus, Sporosarcina, Staphylococcus, Vibrio, Actinomadura, Actinomyces, Kitasatosporia, Streptomyces, Aeromonas, Aureobacterium, Bacillus, Escherichia, Microbacterium, Serratia, Salmonella or Xanthomonas. (III) is a microorganism of the genus Gluconobacter, Acetobacter or Frateuria.

ABEX UPTX: 20001001

EXAMPLE - A 1.2 kilobase XDH DNA fragment was amplified from Morganella morganii genomic DNA, digested with EcoRI and BamHI, and ligated into EcoRI/BamHI-digested pUC18. The product was used to transform Escherichia coli JM109. The transformants were cultured overnight in L medium (1 percent tryptone, 0.5 percent yeast extract, 0.5 percent sodium chloride). The cultures were sonicated and the supernatants (100 microliters) were incubated at 30 degrees Centigrade for 1 minute in a reaction mixture (1 milliliter) containing 100 milliMolar glycine buffer (pH 9.5), 100 milliMolar xylitol and 2 milliMolar nicotinamide adenine dinucleotide. One transformant had an xylitol dehydrogenase activity of 3.9 Units/milligram (1 unit being the activity required for oxidizing 1 micromole of xylitol to generate 1 micromole of NADH per minute).

L66 ANSWER 6 OF 11 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN

AN 1997-558974 [51] WPIX

DNC C1997-178545

Yeast which ferments xylose to ethanol - comprising xylitol reductase, xylitol dehydrogenase and xylulokinase genes integrated at each of its multiple reiterated ribosomal DNA sites.

```
DC
    D16 D17 E17 H06
    CHEN, Z; HO, N W Y
IN
     (PURD) PURDUE RES FOUND
PA
CYC
                   A1 19971113 (199751)* EN
     WO 9742307
                                            66p
                                                     C12N001-16
PΤ
        RW: AT BE CH DE DK EA ES FI FR GB GH GR IE IT KE LS LU MC MW NL OA PT
            SD SE SZ UG
        W: AL AM AT AU AZ BA BB BG BR BY CA CH CN CU CZ DE DK EE ES FI GB GE
           HU IL IS JP KE KG KP KR KZ LC LK LR LS LT LU LV MD MG MK MN MW MX
           NO NZ PL PT RO RU SD SE SG SI SK TJ TM TR TT UA UG US UZ VN YU
                  A 19971126 (199813)
                                                     C12N001-16
     AU 9728301
                  A1 19990303 (199913) EN
                                                     C12N001-16
    EP 898616
        R: AT BE DE DK ES FI FR GB GR IE IT NL PT SE
                A 19990804 (199949)
                                                     C12N001-16
     CN 1225125
     JP 2000509988 W 20000808 (200043)
                                              50p
                                                     C12N015-09
    MX 9809223 A1 19990701 (200061)
                                                     C12N001-16
                  B 20010322 (200122)
    AU 731102
                                                     C12N001-16
                  A 20010731 (200146)
                                                     C12N001-16
    BR 9710963
ADT WO 9742307 A1 WO 1997-US7663 19970506; AU 9728301 A AU 1997-28301
     19970506; EP 898616 A1 EP 1997-922698 19970506, WO 1997-US7663 19970506;
     CN 1225125 A CN 1997-196195 19970506; JP 2000509988 W JP 1997-540153
     19970506, WO 1997-US7663 19970506; MX 9809223 A1 MX 1998-9223 19981105; AU
     731102 B AU 1997-28301 19970506; BR 9710963 A BR 1997-10963 19970506, WO
     1997-US7663 19970506
FDT AU 9728301 A Based on WO 9742307; EP 898616 A1 Based on WO 9742307; JP
     2000509988 W Based on WO 9742307; AU 731102 B Previous Publ. AU 9728301,
     Based on WO 9742307; BR 9710963 A Based on WO 9742307
PRAI US 1996-16865P
                      19960506
REP 6.Jnl.Ref; WO 9513362
     ICM C12N001-16; C12N015-09
         C12N001-18; C12N001-19; C12N015-68; C12N015-69; C12N015-81;
          C12P007-06
ICI C12N001-19; C12N001-19; C12N001-19; C12R001:72; C12R001:84; C12R001:85
          9742307 A UPAB: 19991020
     Novel yeast which ferments xylose to ethanol, comprises: (a)
     xylose reductase (XR), xylitol
     dehydrogenase (XD) and xylulokinase (XK) genes
     integrated at each of its multiple reiterated ribosomal DNA
     sites; (b) multiple copies of exogenous DNA, including XR, XD,
     and XK genes, fused to non-glucose inhibited promoters
     integrated into its chromosomal DNA, where the
     yeast simultaneously ferments glucose and xylose to ethanol; or
     (c) multiple copies of an introduced DNA containing XR, XD and
     XK genes, where the yeast ferments xylose to ethanol,
     where the yeasts of (b) and (c) retain their capacity for
     fermenting xylose to ethanol when cultured under non-selective conditions
     for at least 20 generations.
          USE - The methods can produce yeast, which even upon
     culture in non-selective medium for multiple generations, e.g. up to 20,
     retain their full capability to ferment xylose to ethanol.
     Dwg.0/12
FS
     CPI
FA
     AB; DCN
     CPI: D05-B03; D05-H12E; D05-H14A2; D06-G; E10-E04E2;
MC
L66 ANSWER 7 OF 11 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN
AN
     1995-194082 [25]
                        WPIX
DNC
    C1995-089834
TI
     Recombinant yeast encoding xylose reductase,
     xylitol dehydrogenase and xylulokinase - can
     effectively ferment xylose alone, or simultaneously with glucose, to
     produce ethanol e.g. for use as a fuel.
```

```
DC
     D16 E17 H06
IN
     HO, N W Y; TSAO, G T
PA
     (PURD) PURDUE RES FOUND
CYC
    59
PΙ
                   A1 19950518 (199525)* EN
                                             63p
                                                     C12N001-14
     WO 9513362
        RW: AT BE CH DE DK ES FR GB GR IE IT KE LU MC MW NL OA PT SD SE SZ
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            MD MG MN NO NZ PL RO RU SI SK TJ TT UA UZ VN
     AU 9510517
                  A 19950529 (199537)
                                                     C12N001-14
     EP 728192
                   A1 19960828 (199639) EN
                                                     C12N001-14
         R: AT BE DE DK ES FR GB GR IE IT NL SE
     FI 9601926 A 19960704 (199641)
                                                     C12N000-00
     BR 9408010
                  A 19961217 (199705)
                                                     C12N001-14
     JP 09505469 W 19970603 (199732)
                                              56p
                                                     C12N015-09
     US 5789210 A 19980804 (199838)
                                                     C12P007-08
     AU 695930
                 B 19980827 (199846)
                                                     C12N001-14
     CN 1141057
                  A 19970122 (200047)
                                                     C12N001-14
ADT WO 9513362 A1 WO 1994-US12861 19941108; AU 9510517 A AU 1995-10517
     19941108; EP 728192 A1 WO 1994-US12861 19941108, EP 1995-901176 19941108;
     FI 9601926 A WO 1994-US12861 19941108, FI 1996-1926 19960507; BR 9408010 A
     BR 1994-8010 19941108, WO 1994-US12861 19941108; JP 09505469 W WO
     1994-US12861 19941108, JP 1995-513948 19941108; US 5789210 A US
     1993-148581 19931108; AU 695930 B AU 1995-10517 19941108; CN 1141057 A CN
     1994-194767 19941108
FDT AU 9510517 A Based on WO 9513362; EP 728192 A1 Based on WO 9513362; BR
     9408010 A Based on WO 9513362; JP 09505469 W Based on WO 9513362; AU
     695930 B Previous Publ. AU 9510517, Based on WO 9513362
PRAI US 1993-148581
                     19931108
REP
    4.Jnl.Ref
     ICM C12N000-00; C12N001-14; C12N015-09; C12P007-08
IC
         CO7H021-04; C12N001-19; C12N009-00; C12N009-02; C12N009-12;
          C12N015-00; C12N015-81; C12P007-06
    C12N015-09, C12R001:865; C12N001-19, C12R001:865; C12P007-06, C12R001:865
ICI
AB
          9513362 A UPAB: 19951128
     Recombinant yeast (pref. of the genus Saccharomyces)
     contains introduced genes (pref. fused to non-glucose-inhibited
     promoters) encoding xylose reductase, xylitol
     dehydrogenase (XD) and xylulokinase effective for
     fermenting xylose to ethanol. Also claimed are: (1) a recombinant
     DNA molecule comprising genes encoding xylose
     reductase, XD and xylulokinase; and (2) a vector for
     transforming yeast comprising these genes.
          USE - The yeast can effectively ferment xylose, alone or
     simultaneously with glucose, to produce ethanol; the ethanol can be used
     as liquid fuel for cars either as a neat fuel (100% ethanol) or as a blend
     with petroleum.
          ADVANTAGE - The recombinant yeast are suitable for ethanol
     fuel production by fermentation using plant biomass as feedstock.
     Dwg.0/14
FS
     CPI
FA
     AB; GI; DCN
MC
     CPI: D05-B03; D05-H12C; D05-H12E; D05-H14A2;
          D06-B; E10-E04E2; E11-N; H06-B
L66 ANSWER 8 OF 11 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN
AN
     1994-167479 [20] WPIX
     2001-465360 [50]; 2003-777185 [73]
CR
DNC
TI
     Production of xylitol from recombinant hosts - using a metabolic pathway which
     uses a carbon source other than D-xylose, D-xylulose or their oligomers.
DC
     B05 D13 D16 E17
     APAJALAHTI, J H A; HARKKI, A M; MYASNIKOV, A N; PASTINEN, O A; APAJALAHTI,
IN
     J; HARKKI, A; MYASNIKOV, A; PASTINEN, O; APAJALAHTI, J H; MJASNIKOV, A N;
```

NOVOMIROVICH, A; ANDREI, N M PA (XYRO-N) XYROFIN OY CYC 47 A1 19940511 (199420) * EN 90p C12P007-18 PΙ WO 9410325 RW: AT BE CH DE DK ES FR GB GR IE IT LU MC NL OA PT SE W: AT AU BB BG BR BY CA CH CZ DE DK ES FI GB HU JP KP KR KZ LK LU LV MG MN MW NL NO NZ PL PT RO RU SD SE SK UA US UZ VN A 19940524 (199434) C12P007-18 AU 9454215 C12P007-18 NO 9501747 A 19950705 (199538) FI 9502148 A 19950704 (199540) C12N000-00 EP 672161 A1 19950920 (199542) EN C12P007-18 R: AT BE CH DE DK ES FR GB GR IE IT LI LU NL PT SE NZ 257561 A 19960925 (199644) C12P007-18 g88 JP 08505522 W 19960618 (199648) C12N015-09 34p US 5631150 A 19970520 (199726) C12P019-02 HU 72187 T 19960328 (199741) C12P007-18 B1 19990922 (199943) EN EP 672161 C12P007-18 R: AT BE CH DE DK ES FR GB GR IE IT LI LU NL PT SE DE 69326559 E 19991028 (199951) C12P007-18 BR 9307391 A 19990831 (200002) C12P007-18 ES 2139024 T3 20000201 (200013) C12P007-18 RU 2142999 C1 19991220 (200043) C12N015-52 HU 219016 B 20010131 (200112) C12P007-18 FI 108300 B1 20011231 (200214) C12P007-18 B2 20030804 (200354) 39p C12N015-09 JP 3433295 WO 9410325 A1 WO 1993-FI450 19931105; AU 9454215 A WO 1993-FI450 19931105, AU 1994-54215 19931105; NO 9501747 A WO 1993-FI450 19931105, NO 1995-1747 19950504; FI 9502148 A WO 1993-FI450 19931105, FI 1995-2148 19950504; EP 672161 A1 EP 1993-924615 19931105, WO 1993-FI450 19931105; NZ 257561 A NZ 1993-257561 19931105, WO 1993-FI450 19931105; JP 08505522 W WO 1993-FI450 19931105, JP 1994-510748 19931105; US 5631150 A CIP of US 1992-973325 19921105, Cont of US 1993-110672 19930824, US 1995-368395 19950103; HU 72187 T WO 1993-FI450 19931105, HU 1995-1288 19931105; EP 672161 B1 EP 1993-924615 19931105, WO 1993-FI450 19931105; DE 69326559 E DE 1993-626559 19931105, EP 1993-924615 19931105, WO 1993-FI450 19931105; BR 9307391 A BR 1993-7391 19931105, WO 1993-FI450 19931105; ES 2139024 T3 EP 1993-924615 19931105; RU 2142999 C1 WO 1993-FI450 19931105, RU 1995-113172 19931105; HU 219016 B WO 1993-FI450 19931105, HU 1995-1288 19931105; FI 108300 B1 WO 1993-FI450 19931105, FI 1995-2148 19950504; JP 3433295 B2 WO 1993-FI450 19931105, JP 1994-510748 19931105 FDT AU 9454215 A Based on WO 9410325; EP 672161 Al Based on WO 9410325; NZ 257561 A Based on WO 9410325; JP 08505522 W Based on WO 9410325; HU 72187 T Based on WO 9410325; EP 672161 B1 Based on WO 9410325; DE 69326559 E Based on EP 672161, Based on WO 9410325; BR 9307391 A Based on WO 9410325; ES 2139024 T3 Based on EP 672161; RU 2142999 C1 Based on WO 9410325; HU 219016 B Previous Publ. HU 72187, Based on WO 9410325; FI 108300 B1 Previous Publ. FI 9502148; JP 3433295 B2 Previous Publ. JP 08505522, Based on WO 9410325 PRAI US 1992-973325 19921105; US 1993-110672 19930824; US 1995-368395 19950103 01Jnl.Ref; EP 450430; WO 9115588 REP ICM C12N000-00; C12N015-09; C12N015-52; C12P007-18; C12P019-02 ICC12N001-15; C12N001-19; C12N005-10; C12P019-00 AΒ 9410325 A UPAB: 20031112 Production of xylitol from a recombinant host comprises (a) constructing within a microbial host, a novel metabolic pathway leading to the synthesis of xylitol as an end prod. from a carbon source other than D-xylose, D-xylulose or polymers or oligomers containing D-xylose or D-xylulose as major components, (b) growing the recombinant host under conditions that provide for the synthesis of the xylitol using the pathway and on a carbon source other than those in (a), and (c) recovering the xylitol. Pref. the recombinant host may be transformed with a

construct-encoding enzymes such as D-glucose-6-phosphate dehydrogenase

FS

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MC

AN

DC

IN

PA

ΡI

FI 104636

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(GPD), 6-phospho-D-gluconate dehydrogenase (PGD), D-ribulose-5-phosphate-3-
     epimerase (RPE), D-ribulokinase (RK), xylitol
     dehydrogenase (XD) and D-arabitol dehydrogenase (AD).
          USE/ADVANTAGE - The xylitol produced is used as a sweetener. The
    method can be used to produce xylitol using readily available sources such
     as D-glucose.
    Dwq.0/0
    CPI
    AB; DCN
     CPI: B04-F09; B10-E04C; B11-A01; B14-E11; D03-H01A; D05-C08;
         D05-H14A; D05-H17A3; E10-A07
          5631150 A UPAB: 19970626
     A method for production of xylitol from a recombinant microbial host,
          a) growing an arabitol-producing yeast or an
     arabitol-producing fungus under conditions that provide for the synthesis
     of xylitol, where the arabitol-producing yeast or
     arabitol-producing fungus has been modified to synthesize xylitol as an
     end product in a fermentation when grown on D-arabitol or a carbon source
     that the unmodified arabitol-producing yeast or unmodified
     arabitol-producing fungus utilized for D-arabitol biosynthesis, the carbon
     source being other than D-xylose, D-xylulose, mixtures of D-xylose and
     D-xylulose, and polymers and oligomers containing D-xylose or D-xylulose
     as major components under conditions that provide for synthesis of xylitol
     where before the modification the host produced D-arabitol but did not
     utilize D-arabitol for the synthesis of xylitol, and the host has been
     transformed with a DNA encoding a D-xylulose forming D-arabitol
     dehydrogenase (E.C. 1.1.1.11) and with a DNA encoding
     xylitol dehydrogenase (E.C. 1.1.1.9); and
          b) recovering the xylitol produced in step a.
    Dwg.0/11
L66 ANSWER 9 OF 11 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN
     1991-325230 [44]
                       WPIX
DNC
    C1991-140554
    DNA encoding xylose reductase and/or
     xylitol dehydrogenase - useful for transforming
     yeast strains for expression of one enzyme or co-expression of
     both.
     B05 D13 D16 E17 F09
     AIRAKSINEN, U; HAHN-HAGERDAL, B; HALLBORN, J; KERANEN, S; OJAMO, H;
     PENTTILA, M; WALFRIDSSON, M; HAHN-HAEGERDAL, B; KERAENEN, S; PENTTILAE, M;
     HAHNHAGERD, B; WALFRIDSSO, M
     (VALW) VALTION TEKNILLINEN TUTKIMUSKESKUS; (XYRO-N) XYROFIN OY; (VALW)
     VALTION TEKNILLINEN; (HALL-I) HALLBORN J
CYC
     WO 9115588
                  A 19911017 (199144)*
        RW: AT BE CH DE DK ES FR GB GR IT LU NL SE
         W: AU CA FI JP NO US
                  A 19911030 (199205)
     AU 9175657
                  A 19921002 (199302)
                                                     C12N000-00
     FI 9204461
     NO 9203880
                  A 19921006 (199306)
                                                     C12N015-53
     EP 527758
                  A1 19930224 (199308) EN
                                                     C12N015-53
        R: AT BE CH DE DK ES FR GB GR IT LI LU NL SE
     JP 05507843 W 19931111 (199350)
                                                     C12N015-53
                  B 19940317 (199416)
                                                     C12N015-53
     AU 647104
                  B1 19980107 (199806) EN 24p
                                                     C12N015-53
     EP 527758
        R: AT BE CH DE DK ES FR GB GR IT LI LU NL SE
     DE 69128619 E 19980212 (199812)
                                                     C12N015-53
     ES 2113373
                  T3 19980501 (199824)
                                                     C12N015-53
                  A 19990202 (199912)
                                                     C12P007-02
     US 5866382
                 A 19991006 (200003)
                                                    C12N000-00
     FI 9902153
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C12N015-53

B1 20000315 (200020)

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B1 20000925 (200056)
                                                     C12N015-53
     NO 308544
     CA 2090122
                  C 20020618 (200250) EN
                                                     C12N015-53
     JP 3348215
                  B2 20021120 (200282)
                                              21p
                                                     C12P007-18
                  B1 20030624 (200343)
     US 6582944
                                                     C12P007-06
                   B1 20031114 (200377)
                                                     C12P007-06
     FI 112250
ADT FI 9204461 A WO 1991-FI103 19910408, FI 1992-4461 19921002; NO 9203880 A
     WO 1991-FI103 19910408, NO 1992-3880 19921006; EP 527758 A1 EP 1991-906996
     19910408, WO 1991-FI103 19910408; JP 05507843 W JP 1991-506907 19910408,
     WO 1991-FI103 19910408; AU 647104 B AU 1991-75657 19910408; EP 527758 B1
     EP 1991-906996 19910408, WO 1991-FI103 19910408; DE 69128619 E DE
     1991-628619 19910408, EP 1991-906996 19910408, WO 1991-FI103 19910408; ES
     2113373 T3 EP 1991-906996 19910408; US 5866382 A CIP of US 1990-527775
     19900524, Cont of US 1992-848694 19920309, US 1994-336198 19941103; FI
     9902153 A WO 1991-FI103 19910408, Div ex FI 1992-4461 19921002, FI
     1999-2153 19991006; FI 104636 B1 WO 1991-FI103 19910408, FI 1992-4461
     19921002; NO 308544 B1 WO 1991-FI103 19910408, NO 1992-3880 19921006; CA
     2090122 C CA 1991-2090122 19910408, WO 1991-FI103 19910408; JP 3348215 B2
     JP 1991-506907 19910408, WO 1991-FI103 19910408; US 6582944 B1 CIP of US
     1990-527775 19900524, CIP of WO 1991-FI103 19910408, Cont of US
     1992-848694 19920309, Div ex US 1994-336198 19941103, US 1999-184965
     19990108; FI 112250 B1 WO 1991-FI103 19910408, Div ex FI 1992-4461
     19921002, FI 1999-2153 19991006
FDT EP 527758 A1 Based on WO 9115588; JP 05507843 W Based on WO 9115588; AU
     647104 B Previous Publ. AU 9175657, Based on WO 9115588; EP 527758 B1
     Based on WO 9115588; DE 69128619 E Based on EP 527758, Based on WO
     9115588; ES 2113373 T3 Based on EP 527758; FI 104636 B1 Previous Publ. FI
     9204461; NO 308544 B1 Previous Publ. NO 9203880; CA 2090122 C Based on WO
     9115588; JP 3348215 B2 Previous Publ. JP 05507843, Based on WO 9115588; US
     6582944 B1 Div ex US 5866382; FI 112250 B1 Previous Publ. FI 9902153
PRAI FI 1990-1771
                     19900406
    7.Jnl.Ref
REP
     ICM C12N000-00; C12N015-53; C12P007-02; C12P007-06; C12P007-18
TC
         C07H021-04; C12N001-15; C12N001-19; C12N009-02; C12N015-52;
          C12N015-81; C12P019-02
    C12N009-04; C12N015-09
ICA
    C12P007-18, C12R001:865
ICI
          9115588 A UPAB: 20040123
AB
       DNA (I) encoding xylose reductase enzyme (A)
     is new. When (I) is transferred into a yeast strain it renders
     the strain capable of reducing xylose to xylitol.
          Pref. the yeast of (1) is capable of integrating to the
     yeast chromosome when transformed into a yeast
     strain. (I) and/or (II) is expressed under yeast gene
     regulatory regions, e.g. promoters of (A), (B), yeast alcohol
     dehydrogenase gene ADH1 or yeast phosphogylcerate
     kinase gene PGK1, and functional fragments. The yeast
     strain is a Saccharomyces cerevisiae strain (pref.),
     kluyveromyces strain, Schizosacchoromyces pombe strain or Pichia strain.
          The yeast vectors pUA103, pUA107, pJHXR22, pMW22, pJHXDH60
     and pJHXDH70, and the yeast strains. S. cerivisiae H475, H477,
     H479, H481, VTT-C-91181, H949, H495, H496, H497, H492 and H493 are
     specifically claimed.
          USE - The yeast transformants can reduce xylase to xylitol
     for use by diabetics or as a natural sweetener. The co-expression of the
     two enzymes in a yeast strain results in the production of ethanol.
     Dwg.0/8
FS
     CPI
FΑ
     AB; DCN
     CPI: B04-B02B2; B04-B02C2; B04-B04A1; B10-A07; B10-E04D; B11-A; B12-J01;
MC
          B12-L03; D05-B03; D05-C03B; D05-C03D; D05-H03B;
         D05-H12; E10-A07; F05-A02C; F05-B
          527758 A UPAB: 19930928
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DNA (I) encoding xylose reductase enzyme (A)

ABEQ EP

is new. When (I) is transferred into a **yeast** strain it renders the strain capable of reducing xylose to xylitol.

Pref. the **yeast** of (1) is capable of integrating to the **yeast** chromosome when transformed into a **yeast** strain. (I) and/or (II) is expressed under **yeast** gene regulatory regions, e.g. promoters of (A), (B), **yeast** alcohol

dehydrogenase **gene** ADH1 or **yeast** phosphoglycerate kinase **gene** PGK1, and functional fragments. The **yeast** strain is a **Saccharomyces** cerevisiae strain (pref.),

kluyveromyces strain, Schizosaccharomyces pombe strain or Pichia strain.

The yeast vectors pUA103, pUA107, pJHXR22, pMW22, pJHXDH60, and pJHXDH70, and the yeast strains S. cerivisiae H475, H477, H479, H481, VTT-C-91181, H949, H495, H496, H497, H492 and H493 are specifically claimed.

USE - The **yeast** transformants can reduce xylase to xylitol for use by diabetics or as a natural sweetener. The co-expression of the two enzymes in a **yeast** strain results in the prodn. of ethanol EP 527758 B UPAB: 19980209

DNA (I) encoding xylose reductase enzyme (A) is new. When (I) is transferred into a yeast strain it renders the strain capable of reducing xylose to xylitol.

Pref. the yeast of (1) is capable of integrating to the yeast chromosome when transformed into a yeast strain. (I) and/or (II) is expressed under yeast gene regulatory regions, e.g. promoters of (A), (B), yeast alcohol dehydrogenase gene ADH1 or yeast phosphogylcerate kinase gene PGK1, and functional fragments. The yeast strain is a Saccharomyces cerevisiae strain (pref.), kluyveromyces strain, Schizosacchoromyces pombe strain or Pichia strain.

The yeast vectors pUA103, pUA107, pJHXR22, pMW22, pJHXDH60 and pJHXDH70, and the yeast strains. S. cerivisiae H475, H477, H479, H481, VTT-C-91181, H949, H495, H496, H497, H492 and H493 are specifically claimed.

USE - The **yeast** transformants can reduce xylase to xylitol for use by diabetics or as a natural sweetener. The co-expression of the two enzymes in a **yeast** strain results in the prodn. of ethanol. Dwq.0/8

L66 ANSWER 10 OF 11 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN

AN 1991-296506 [41] WPIX

DNC C1991-128227

New DNA encoding xylose reductase and xylitol dehydrogenase - and transformed yeast for production of ethanol and biomass from xylose or recovery of oxidised NADP.

DC B02 B04 D16 E17

IN AMORE, R; HAGEDORN, J; HOLLENBERG, C P; KOTTER, P; PIONTEK, M; STRASSER, A W M; VONCIRIACY, M; KOETTER, P; VON, CIRIACY-WANTRUP M; HAGENDORN, J

PA (RHEI-N) RHEIN BIOTECH NEUE BIOTECHNOLOGISCHE PROZESSE & PROD GMBH; (RHEI-N) RHEIN BIOTECH GES NEUE BIOTECHNOLOGISCHE; (RHEI-N) RHEIN BIOTECH GES B; (RHEI-N) RHEIN BIOTECH GMBH

CYC 16

PΙ DE 4009676 A 19911002 (199141) * 50p A 19911009 (199141) EP 450430 R: AT BE CH DE DK ES FR GB GR IT LI LU NL SE CA 2039021 A 19910927 (199150) A3 19920102 (199320) EP 450430 50p C2 19930909 (199336) C12N001-19 DE 4009676 51p JP 06339383 A 19941213 (199509) 32p C12N015-53 EP 450430 B1 19970625 (199730) EN C12N015-53 R: AT BE CH DE DK ES FR GB GR IT LI LU NL SE

DE 69126632 E 19970731 (199736) C12N015-53 ES 2104626 T3 19971016 (199748) C12N015-53

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C12N015-09
    JP 2000139486 A 20000523 (200033)
                                              21p
                                              32p
                  B2 20010109 (200104)
                                                     C12N015-09
    JP 3122153
    JP 2001103988 A 20010417 (200128)
                                                     C12N015-09
                                              27p
                                              27p
                  B2 20010730 (200146)
                                                     C12N015-09
    JP 3193917
ADT DE 4009676 A DE 1990-4009676 19900326; EP 450430 A EP 1991-104558
    19910322; EP 450430 A3 EP 1991-104558 19910322; DE 4009676 C2 DE
    1990-4009676 19900326; JP 06339383 A JP 1991-62160 19910326; EP 450430 B1
    EP 1991-104558 19910322; DE 69126632 E DE 1991-626632 19910322, EP
    1991-104558 19910322; ES 2104626 T3 EP 1991-104558 19910322; JP 2000139486
    A Div ex JP 1991-62160 19910326, JP 2000-589 19910326; JP 3122153 B2 JP
     1991-62160 19910326; JP 2001103988 A Div ex JP 1991-62160 19910326, JP
     2000-276227 19910326; JP 3193917 B2 Div ex JP 1991-62160 19910326, JP
     2000-276227 19910326
FDT DE 69126632 E Based on EP 450430; ES 2104626 T3 Based on EP 450430; JP
     3122153 B2 Previous Publ. JP 06339383; JP 3193917 B2 Previous Publ. JP
     2001103988
PRAI DE 1990-4009676 19900326
    NoSR.Pub; 6.Jnl.Ref; EP 238023; GB 2151635; JP 60199383; JP 61063291; US
     4840903
     C07H021-04; C07K015-04; C12C011-00; C12N001-19; C12N009-02; C12N015-63;
IC
     C12P007-06; C12P019-34
     ICM C12N001-19; C12N015-09; C12N015-53
         C07H021-04; C07K013-00; C07K015-04; C12C011-00; C12N001-14;
          C12N001-21; C12N009-02; C12N009-04; C12N015-63; C12N015-81;
          C12P007-06; C12P007-10; C12P019-34; C12P021-02
ICI C12N009-02; C12N009-02; C12N015-09; C12N015-09; C12N015-09; C12R001:645;
          C12R001:84; C12R001:84; C12R001:85; C12R001:865; C12N001-19;
          C12N001-19; C12N001-19; C12N001-21; C12N009-04; C12N015-09;
          C12P007-06; C12P007-06; C12P007-06; C12P007-06; C12R001:01;
          C12R001:01; C12R001:645; C12R001:645; C12R001:84; C12R001:84;
          C12R001:84; C12R001:84; C12R001:865; C12R001:865; C12N001-19,
          C12R001:865; C12N009-02, C12R001:865; C12P007-06, C12R001:865;
          Cl2N015-53, Cl2R001:645; Cl2N001-19, Cl2R001:865; Cl2N009-02,
          C12R001:865
AB
     DE
          4009676 A UPAB: 19971030
     New DNA sequence (I) comprises a structural gene
     encoding a xylose reductase (XR) and/or xylyl
     dehydrogenase (XDH) and is able to express the enzyme(s) in a
     microorganism.
          Also new are (1) combinations of (I) with other DNA halogen
     sequences for regulating expression; (2) vectors and microorganisms containing
     (I), and (3) XR and XDH produced by expressing (I).
          More specifically, (I) is derived from a yeast,
     specifically Pichia stipitis CBS 5773 (DSM 5855). The specification
     includes sequences for DNA fragments which encode XR (2040
     bases) and \bar{\text{XDH}} (1950 bases), and the derived structures (318 and 363
     amino acids, respectively).
          USE/ADVANTAGE - XR and XDH are useful (1) for production of ethanol from
     xylose (a waste prod. of cellulose mfr.); (2) for production of biomass and
     (3) for recovery of NADP(+) for NADPH. The microorganisms transformed with
     (I) can ferment highly concentrate carbohydrate solns. prior art and are
     tolerant to EtOH, pH and temperature. Also contemplated is production of
specific
     proteins (II) in P. stipitis by expressing the structural gene
     for (II) under control of the 5'-regulatory region of the XR and XDH
     genes of P. stipitis (these are inducible by xylose) and/or the
     ADH1 promoter of S. cerevisiae and/or the glucoamylase promoter of
     Schwanniomyces occidentalis. P. Stipitis has an efficient secretory
     system and can use xylose as a C source. @(50pp Dwg.No.0/7)
     CPI
FS
     AB; DCN
FA
     CPI: B04-B02B2; B04-B02C2; B04-B04A1; D05-C03B; D05-H03B;
MC
          D05-H05; D05-H12; E10-E04E2
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450430 A UPAB: 19931113
ABEQ EP
     New DNA sequence (I) comprises a structural gene
     encoding a xylose reductase (XR) and/or xylyl
     dehydrogenase (XDH) and is able to express the enzyme(s) in a
     microorganism.
          Also new are (1) combinations of (I) with other DNA halogen
     sequences for regulating expression; (2) vectors and microorganisms contg.
     (I), and (3) XR and XDH produced by expressing (I).
          More specifically, (I) is derived from a yeast,
     specifically Pichia stipitis CBS 5773 (DSM 5855). The specification
     includes sequences for DNA fragments which encode XR (2040
     bases) and XDH (1950 bases), and the derived structures (318 and 363
     amino acids, respectively).
          USE/ADVANTAGE - XR and XDH are useful (1) for prodn. of ethanol from
     xylose (a waste prod. of cellulose mfr.); (2) for prodn. of biomass and
     (3) for recovery of NADP(+) for NADPH. The microorganisms transformed with
     (I) can ferment highly conc. carbohydrate solns. prior art and are
     tolerant to EtOH, pH and temp.. Also contemplated is prodn. of specific
     proteins (II) in P. stipitis by expressing the structural gene
     for (II) under control of the 5'-regulatory region of the XR and XDH
     genes of P. stipitis (these are inducible by xylose) and/or the
     ADH1 promoter of S. cerevisiae and/or the glucoamylase promoter of
     Schwanniomyces occidentalis. P. Stipitis has an efficient secretory
     system and can use xylose as a C source. @(50pp Dwg.No.0/7)
          4009676 C UPAB: 19931122
ABEQ DE
     Recombinant DNA sequence that encodes the prodn. of an
     xylosereductase and/or xylitoldehydrogenase has been utlised in expression
     vectors contg. this DNA to produce the enzymes. Host cells have
     been transformed with these vectors and then propagated to produce the
     exogenous polypeptides. The nucleotide sequence of the
     cDNA and the aminoacid sequences of the polypeptides are defined.
          USE - The prods. facilitate the degradation of xylose from wood pulp,
     leading to the conversion of waste biomass to alcohol.
     Dwq.0/7
ABEQ EP
           450430 B UPAB: 19970723
     New DNA sequence (I) comprises a structural gene
     encoding a xylose reductase (XR) and/or xylyl
     dehydrogenase (XDH) and is able to express the enzyme(s) in a
     microorganism.
L66 ANSWER 11 OF 11 WPIX COPYRIGHT 2004 THOMSON DERWENT on STN
     1990-276784 [37] WPIX
AN
DNN N1990-213906
                        DNC C1990-119559
    DNA coding for placenta-specific protein-9 - and recombinant
TI
     protein with aldose reductase activity.
DC
     B04 D16 P34 S03
ΙŅ
    GRUNDMANN, U; AMANN, E
PA
     (BEHW) BEHRINGWERKE AG
CYC 18
                  A 19900912 (199037)*
PΤ
    EP 386733
        R: AT BE CH DE ES FR GB IT LI LU NL SE
    DE 3907744 A 19900920 (199039)
    AU 9051103 A 19900913 (199044)
    PT 93381 A 19901107 (199047)
CA 2011877 A 19900910 (199048)
     JP 02295486 A 19901206 (199104)
     EP 386733 B1 19950705 (199531) DE
                                                     C12N015-12
                                              11p
        R: AT BE CH DE DK ES FR GB IT LI LU NL SE
     DE 59009360 G 19950810 (199537)
                                                     C12N015-12
     ES 2076238
                  T3 19951101 (199550)
                                                     C12N015-12
     IE 67797
                 B 19960501 (199629)
                                                     C12N015-12
ADT EP 386733 A EP 1990-104352 19900307; DE 3907744 A DE 1989-3907744
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19890310; JP 02295486 A JP 1990-59800 19900309; EP 386733 B1 EP

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1990-104352 19900307; DE 59009360 G DE 1990-509360 19900307, EP
    1990-104352 19900307; ES 2076238 T3 EP 1990-104352 19900307; IE 67797 B IE
    1990-853 19900309
FDT DE 59009360 G Based on EP 386733; ES 2076238 T3 Based on EP 386733
PRAI DE 1989-3907744 19890310
    1.Jnl.Ref; EP 37963; 01Jnl.Ref
    A61K031-70; A61K037-02; A61N037-02; C07H021-04; C07K013-00; C07K015-12;
    C07K015-28; C12N001-21; C12N005-00; C12N015-12; C12P021-02; C12Q001-68;
     G01N033-53; G01N033-68
     ICM C12N015-12
         A61K031-70; A61K037-02; A61N037-02; C07H021-04; C07K013-00;
     ICS
          C07K015-12; C07K015-28; C12N001-21; C12N005-00; C12P021-02;
          C12Q001-68; G01N033-53; G01N033-577; G01N033-68
    C12N001-21, C12R001:
           386733 A UPAB: 19930928
AΒ
     The following are claimed: (a) DNA coding for placenta-specific
     protein 9 (PP9), where the amino acid sequence of PP9 and the coding
     sequence of the DNA are shown in the fig.; (b) DNA and
     RNA which hybridise with DNA (a) under stringent conditions; (c)
     gene structures, vectors and transformed cells containing
     nucleic acids (a) or (b); (d) PP) when produced in E. coli,
     yeast or animal cells by expression of DNA (a); (e)
     polyclonal and monoclonal antibodies specific to PP9, when produced by
     immunisation with recombinant
          USE - PP9 is probably identical to human aldose
     reductase and is thus useful for screening and identifying
     aldose reductase inhibitors for treatment of diabetic
     complications. The nucleic acid and antibodies are useful for
     diagnostic purposes, and the antibodies are also useful for purificn. of
     pp9 by affinity chromatography.
     1/1
     CPI EPI GMPI
FS
FA
     AB; GI
     CPI: B04-B02B1; B04-B02B2; B04-B02C2; B04-B04A; B04-B04C5; B04-B04C6;
MC
          B12-H05; D05-H09; D05-H11; D05-H12
     EPI: S03-E14H
           386733 B UPAB: 19950810
ABEQ EP
     A nucleotide sequence shown in Table 1, or a sequence derived
     therefrom on the basis of the degeneracy of the genetic code,
     coding for placenta-specific protein PP9.
     Dwq.0/1
=> d his
     (FILE 'HOME' ENTERED AT 15:35:08 ON 04 MAR 2004)
     FILE 'REGISTRY' ENTERED AT 15:35:42 ON 04 MAR 2004
                SET COST OFF
                E XYLOSE REDUCTASE/CN
              4 S E3
L1
                E XYLITOL DEHYDROGENASE/CN
              2 S E3
L2
                E XYLULOKINASE/CN
              1 S E3
T.3
             26 S XYLOSE REDUCTASE
L4
              7 S XYLITOL DEHYDROGENASE
L_5
             33 S XYLULOKINASE
L6
     FILE 'HCAPLUS' ENTERED AT 15:37:10 ON 04 MAR 2004
           2967 S L1-L6
L7
            464 S XYLOSE REDUCTASE OR XYLITOL DEHYDROGENASE OR XYLULOKINASE
L8
           3153 S ALDOSE REDUCTASE
L9
```

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68 S XYLULOSE REDUCTASE
L10
             50 S XYLULOSE KINASE
L11
           3760 S L7-L11
L12
            218 S L12 AND YEAST
L13
                E YEAST/CT
             25 S L12 AND E3-E53
L14
                E E53+ALL
          12225 S E1
L15
                E E2+ALL
          34588 S E6, E5+NT
L16
             25 S L12 AND L15, L16
L17
            218 S L13, L14, L17
L18
              1 S YEAST (L) 1400 (L) LNH (L) ST
L19
                E SACCHAROMYCES/CT
              9 S L12 AND E3
L20
            114 S L12 AND E3-E196
L21
                E E3+ALL
              0 S L12 AND E4
L22
            113 S L12 AND E5+NT
L23
            269 S L18, L20-L23
L24
            32 S L24 AND DNA
L25
             31 S L24 AND PLASMID
L26
             20 S L24 AND CHROMOSOM?
L27
            126 S L24 AND GENE
L28
             74 S L24 AND GENETIC?/SC,SX
L29
            130 S L25-L29
L30
L31
            269 S L24-L30
                 E WO97-US7663/AP, PRN
               1 S E3, E4
L32
                 E US96-016865/AP, PRN
               1 S E5
L33
                 E HO N/AU
L34
              20 S E3,E11,E12
              36 S E28, E31, E32
L35
                E CHEN Z/AU
            722 S E3, E7
L36
                E CHEN ZHENG/AU
L37
            261 S E3, E4
              8 S E51
L38
              1 S L31 AND L32,L33
L39
             11 S L31 AND L34-L38
L40
             11 S L32, L33, L39, L40
L41
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L42
L43
              9 S L41 AND L42
             11 S L41, L43
L44
             27 S L42 AND GENETIC?/SC,SX
L45
             31 S L44, L45
L46
            105 S L42 NOT L46
L47
                 SEL DN AN 22 34 L47
               2 S E1-E6 AND L47
L48
              33 S L46, L48 AND L7-L48
L49
                 SEL HIT RN
     FILE 'REGISTRY' ENTERED AT 16:16:43 ON 04 MAR 2004
L50
              14 S E7-E20
              14 S L50 AND L1-L6
L51
     FILE 'REGISTRY' ENTERED AT 16:17:14 ON 04 MAR 2004
     FILE 'HCAPLUS' ENTERED AT 16:17:29 ON 04 MAR 2004
     FILE 'WPIX' ENTERED AT 16:17:48 ON 04 MAR 2004
L52
             784 S L8/BIX OR L9/BIX OR L10/BIX
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L53
           24 S L52 AND YEAST/BIX
L54
            0 S L19/BIX
           11 S L52 AND SACCHAROMYC?/BIX
L55
           21 S L52 AND (?PLASMID? OR ?CHROMOSOM?)/BIX
L56
           38 S L53,L55,L56
L57
           2 S L52 AND (HO N? OR CHEN Z?)/AU
2 S L57 AND L58
L58
L59
           27 S L57 AND D05-H?/MC
L60
           30 S L57 AND (GENE OR GENETIC? OR DNA OR CDNA)/BIX
L61
L62
           15 S L57 AND (?NUCLEIC? OR ?NUCLEO?)/BIX
           32 S L59-L62
L63
            6 S L57 NOT L63
L64
             SEL DN AN 5 15-17 20 25 27-31 L63
            11 S E21-E43 AND L63
L65
            11 S L59, L65
L66
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FILE 'WPIX' ENTERED AT 16:25:58 ON 04 MAR 2004